

ILLUMINA, INC.,
Petitioner,

v.

THE TRUSTEES OF COLUMBIA UNIVERSITY
IN THE CITY OF NEW YORK
Patent Owner.

IPR2018-00291, -00318, -00322, -00385, -00797

Patent Owner's Demonstratives

U.S. Patent Nos.

9,718,852 9,719,139 9,708,385 9,725,480 9,868,985

Columbia Exhibit No. 2141
Illumina v. Columbia
IPR2018-00291, 318, 322, 385, 797

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Columbia's Patents

Relevant to:

IPR2018-00291, 318, 322, 385: Grounds 1-2

IPR2018-00797: Grounds 1-3

Columbia Inventors' Insights

Recognition of Problem & Solution:

Spatial Constraints

(e.g., max. 3.7Å capping group)

Chemical Constraints

(e.g., no ketone, ester, methoxy)



No SBS Success

2000

SBS Success

Before 2000

After 2000

Efforts to Develop SBS

Columbia's Invention (Spatial, Chemical Constraints)

**Tsien/Dower
Filed**

Metzker abstract:
2-nitrobenzyl
incorporated

Metzker 1994:
2-nitrobenzyl
incorporated

Metzker 1998:
2-nitrobenzyl
incorporated

Stemple:
reiterating
2-nitrobenzyl
incorporated

▶▶▶ Dr. Romesberg: "[A] large number of companies and research groups pursued [SBS]" ▶▶▶

1990	1991	1992	1993	1994	1995	1996	1997	1998	1999	2000
------	------	------	------	------	------	------	------	------	------	------

**Provide no guidance for
the capping group**

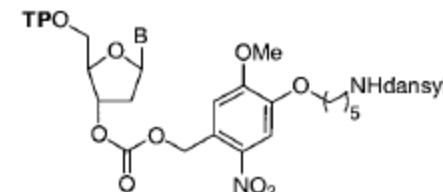
- (i) Large groups
- (ii) Labels on groups
- (iii) Ester; ketone; methoxy

"formidable
obstacles"

"major
challenge"

Welch testing

- (i) Non-ether
- (ii) Large group



Columbia's Patent Claims Are Narrowly Tailored

'852 patent claim vs. '869 patent claim 16

■ Narrower terms

■ New terms

UNITED STATES PATENT AND

BEFORE THE PATENT TRI

ILLUMINATING
Petition

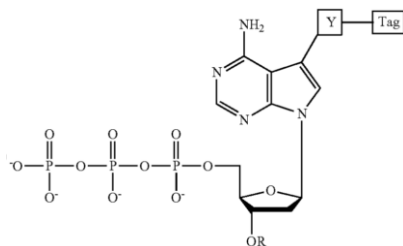
THE TRUSTEES OF COLUMBIA UNIVERSITY
IN THE CITY OF NEW YORK
Patent Owner

IPR2018-00291 (Patent)
IPR2018-00318 (Patent)
IPR2018-00322 (Patent)
IPR2018-00385 (Patent)

PATENT OWNER

Paper No. _____
Filed: October 26, 2018

1. An **adenine** deoxyribonucleotide analogue having the structure:



wherein R (a) represents a **small, chemically cleavable,** chemical group capping the oxygen at the 3' position of the deoxyribose of the deoxyribonucleotide analogue, (b) **does not interfere with recognition of the analogue as a substrate by a DNA polymerase,** (c) **is stable during a DNA polymerase reaction,** and (d) **does not contain a ketone group;**

wherein OR is not a methoxy group or an ester group;

wherein the covalent bond between the 3'-oxygen and R is stable during a DNA polymerase reaction;

wherein tag represents a detectable **fluorescent** moiety;

wherein Y represents a **chemically cleavable, chemical** linker which (a) **does not interfere with recognition of the analogue as a substrate by a DNA polymerase** and (b) **is stable during a DNA polymerase reaction;**

and wherein the adenine deoxyribonucleotide analogue:

- i) **is recognized as a substrate by a DNA polymerase,**
- ii) **is incorporated at the end of a growing strand of DNA during a DNA polymerase reaction,**
- iii) **produces a 3'-OH group on the deoxyribose upon cleavage of R,**
- iv) **no longer includes a tag on the base upon cleavage of Y,** and
- v) **is capable of forming hydrogen bonds with thymine or a thymine nucleotide analogue.**

¹ An identical Paper is being entered into e

Illumina's Metzker-Based Arguments Fail

Relevant to:

IPR2018-00291, 318, 322, 385: Grounds 1-2

IPR2018-00797: Grounds 1-3

The Board Requested Additional Briefing Regarding Metzker's Teachings: Metzker Teaches Inefficient 3'-O-Allyl-dATP Incorporation

Trials@uspto.gov
Tel: 571-272-7822

Paper 20
Entered: September 18, 2018

UNITED STATES PATENT AND TRADEMARK OFFICE

BEFORE THE PATENT TRIAL AND APPEAL BOARD

ILLUMINA, INC.,
Petitioner,

v.

THE TRUSTEES OF COLUMBIA UNIVERSITY
IN THE CITY OF NEW YORK,
Patent Owner.

Case IPR2018-00797
Patent 9,868,985 B2

Case IPR2018-00797
Patent 9,868,985 B2

As Patent Owner notes, Tsien itself sets forth three requirements for a blocking group: accurate and effective incorporation by a polymerase, availability of mild conditions for rapid and quantitative deblocking, and ability of a polymerase to reinitiate cDNA synthesis. Prelim. Resp. 30–31; Ex. 1013, 20–21. These considerations were, therefore, known to a person of ordinary skill and would have been considered both individually and as a whole.

The above determinations are preliminary in nature and based on the record at this stage of the proceeding. The parties are encouraged to develop arguments further at trial, e.g., whether the limitations of an O-allyl blocking group noted in Metzker would have discouraged a person of ordinary skill from using such a group for DNA polymerization reactions.^{8,9} On this

Before JAMES A. WORTH, M.
BRIAN D. RANGE, *Administrative Patent*

RANGE, *Administrative Patent*

Institution
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record at this stage of the proceeding. The parties are encouraged to develop arguments further at trial, e.g., whether the limitations of an O-allyl blocking group noted in Metzker would have discouraged a person of ordinary skill from using such a group for DNA polymerization reactions.^{8,9} On this

Tsien regarding an allyl group's ability to serve as a cleavable blocking group.

⁹ Petitioner asserts that a person of ordinary skill would have recognized that Tsien's allyl group is advantageous for an additional reason. Petitioner

IPR2018-00797,
Institution Dec., 29

No Dispute that SBS Requires Efficient Incorporation

PCT WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification: C12Q 1/68, C12P 19/34, G01H 21/00, 21/04, G01N 35/08 A1 (11) International Publication Number: WO 91/06678 (43) International Publication Date: 16 May 1991 (16.05.91)

(21) International Application Number: (22) International Filing Date: 26 October 1989 (30) Priority Data: 427,321 26 October 1989 (71) Applicant: SRI INTERNATIONAL, Inc., 3200 Avenue, Menlo Park, CA 94025 (71)(72) Applicant and Inventor: TSIEH, P. 8535 Nottingham Place, La Jolla, CA 92037 (72) Inventors: ROSS, Pepi; 745 Contra Costa Ave., CA 94707 (US); FAHNESTOCK, Gamble Court, Hayward, CA 94542; Allan, J.; 909 North California Ave., 94303 (US).

(54) Title: DNA SEQUENCING

(57) Abstract
The present invention relates to an improved method for sequencing DNA without the use of a gel electrophoresis step. The invention involves the immobilization or entrapment within a chamber of a blocked nucleotide by means of a coupling reaction. The invention also relates to a method of determining the sequence of a DNA strand by the addition of a blocked nucleotide having an innocuous marker so that a rapid accurate determination of a DNA sequence is possible.

Blocking Groups and Methods for Incorporation

The coupling reaction generally employs 3'-hydroxyl-blocked dNTPs to prevent inadvertent extra additions.

The criteria for the successful use of 3'-blocking groups include:

- (1) the ability of a polymerase enzyme to accurately and efficiently incorporate the dNTPs carrying the 3'-blocking groups into the cDNA chain,
- (2) the availability of mild conditions for rapid and quantitative deblocking, and

Ex. 1013, Tsien, 20

Illumina Ex. 1013
IPR Petition - USP 9,718,852

35

Metzker 1994 Taught Termination*

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Nucleic Acids Research, 1994, Vol. 22, No. 20 4259-4267

Termination of DNA synthesis by novel 3'-modified-deoxyribonucleoside 5'-triphosphates

Michael L. Metzker*, Ramesh Raghavachari^{1,†}, Stephen Richards, Swanee E. Jacutin¹, Andrew Civitello, Kevin Burgess¹ and Richard A. Gibbs
Department of Molecular and Human Genetics, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030 and ¹Department of Chemistry, Texas A & M University, College Station, TX 77843, USA

Received June 3, 1994; Revised and Accepted August 1, 1994

ABSTRACT

Eight 3'-modified-dNTPs were synthesized in two different DNA template assays. From this enzymatic sort, dNTPs were shown to terminate DNA synthesis by a number of polymerases as alternative terminators in Sanger (2-Nitrobenzyl)-dATP is a UV sensitive was shown to be incorporated by DNA polymerases. Base specific efficient photolytic removal of the was demonstrated. Following synthesis was reinitiated by the in nucleotides into DNA. The identified terminator and the demonstration start DNA synthesis are initial steps of a novel sequencing strategy.

INTRODUCTION

2'-Deoxyribonucleoside-5'-triphosphates their 3'-hydroxyl position can act as directed DNA synthesis (1-12). The useful as DNA sequencing tools antimetabolites, and as antiviral agents compounds have been used for analytical (2). Overall, however, the number of characterized is small, and there is combinations of terminators and polymerases. Among the most familiar terminators the 2', 3'-dideoxynucleoside-5'-triphosphates are the basis for Sanger DNA sequencing oligonucleotide-primed DNA or RNA cally extended in a 5' → 3' direction in of dNTPs and ddNTPs to generate a pool are terminated at specific base positions different lengths are resolved by denaturation.

*To whom correspondence should be addressed.

† Present address: LI-COR Inc., Biotechnology

Table 2. Activity matrix of RP-HPLC purified 3'-protecting dNTPs challenged against commercially available polymerases

3'-modified-dATP (except compound [8])	AMV-RT	M-MuLV-RT	Klenow fragment	Sequenase®	Bst DNA polymerase	AmpliTaq® DNA polymerase	Vent(r(exo-))® DNA polymerase	rTth DNA polymerase
[1] O-methyl	Termination	Termination*	-	-	-	-	Inhibition	Inhibition*
[2] O-acyl	-	-	-	-	-	-	Inhibition	-
[3] O-allyl	-	-	-	-	-	-	Termination*	-
[4] O-tetrahydropyran	-	-	-	-	-	-	-	-
[5] O-(4-nitrobenzoyl)	-	-	-	-	-	-	-	-
[6] O-(2-aminobenzoyl)	-	-	-	-	-	-	-	-
[7] O-(2-nitrobenzyl)	-	-	-	Inhibition	Termination	Termination*	Termination*	-
[8] 3'-O-methyl-dTTP	-	Inhibition	-	Inhibition	Termination	Termination	Termination	Termination

All compounds were assayed at a final concentration of 250 μ M according to the conditions specified in Table 1. '-' means no activity was detected, 'Termination' means that the termination bands mimic ddNTP termination bands, and 'Inhibition' means the rate of DNA synthesis is reduced in a nonspecific manner. '*' means the activity was incomplete at a final concentration of 250 μ M.

Termination* Results From Inefficient Incorporation

Published online 11 January 2011

Nucleic Acids Research, 2011, Vol. 39, No. 6 e39
doi:10.1093/nar/gkq1293

Improved nucleotide selectivity and termination of 3'-OH unblocked reversible terminators by molecular tuning of 2-nitrobenzyl alkylated HOMedU triphosphates

Vladislav A. Litosh¹, Weidong Wu¹, Brian P. Stupi¹, Jinchun Wang^{1,2}, Sidney E. Morris^{1,2}, Megan N. Hersh¹ and Michael L. Metzker^{1,2,3,*}

¹LaserGen, Inc., Houston, TX 77054, ²Human Genome Sequencing Center and ³Department of Molecular & Human Genetics, Baylor College of Medicine, Houston, TX 77030, USA

Received September 30, 2010; Revised November 10, 2010; Accepted December 1, 2010

e39 Nucleic Acids Research, 2011, Vol. 39, No. 6

Page 10 of 13

ABSTRACT
We describe a novel 3'-O-allyl-terminated nucleotide with the potential to improve sequencing efficiency and read-lengths in next-generation sequencing (NGS) technologies. This

N⁶-alkylated thymidine analog would interfere with Watson-Crick base pairing, adversely affecting nucleotide selectivity. We have confirmed this hypothesis by synthesizing and characterizing N⁶-(2-nitrobenzyl)-TTP (T1). Of the eight DNA polymerases tested, only Vent(exo⁻) and Terminator II polymerase incorporated T1 with efficiency

which is analogous to the modified Vent(exo⁻) A48SL polymerase. Gardner and Jack (49) suggested that the modified Vent(exo⁻) A48SL polymerase is more tolerant of base-modified nucleotides, but with the trade-off of lower nucleotide selectivity. Here, Terminator II and Vent(exo⁻) polymerase incorporation of T1 with efficiency

these nucleotides (10). We note that 3'-O-allyl-dATP is not efficiently incorporated by Vent(exo⁻) polymerase (46). Ju and colleagues have reported, however, that

Bzyl-2-nitrobenzyl-3'-O-allyl-dATP (T1) was identified as an efficient terminator, sequencing faster in a cyclic reversible termination (CRT) assay using a homopolymer repeat template bases without during photochemical cleavage. Our results validate our earlier 3'-OH unblocked reverse

terminator. Sequence (44), a 100 polymerase containing the same 1667T amino acid variant (45) as TaqFS polymerase, using the non-repetitive template sequence 5'-TCTGATACATG. A single nucleotide addition strategy (16) was used to demonstrate DNA sequencing feasibility, whereby, the polymerase is paused only until addition of the next complementary nucleotide. In their companion paper, the authors stated that the 3'-OH requires blocking with a chemical moiety, such as an 3'-O-allyl group, to effect the termination property of these nucleotides (10). We note that 3'-O-allyl-dATP is not efficiently incorporated by Vent(exo⁻) polymerase (46). Ju and colleagues have reported, however, that their dysubstituted 3'-O-allyl-dATPs were incorporated with Terminator II polymerase, which contains the Y49V amino acid variant in addition to the A48SL variant found in the Terminator polymerase. The 409 amino acid residue of Terminator acts as a steric gate for incorporation of ribonucleotides (NTPs) (47-49). In contrast, the 3'-OH unblocked 5-(5'-2-oxo-butyl)-2-nitrobenzyl-3'-O-allyl-dATP (T1) is an efficient terminator with both Vent(exo⁻) and Terminator polymerases, but is not incorporated with TaqFS polymerase. Proximity of the 2-nitrobenzyl group to the nucleobase and size of the alkyl group attached to its 5-methylene carbon are important structural features that confer the unique properties of termination and selectivity upon 3'-OH unblocked 5-(5'-2-oxo-butyl)-2-nitrobenzyl-3'-O-allyl-dATP (T1).

Vent(exo⁻) and Terminator are highly homologous polymerases, showing ~77% identity and ~89% similarity by protein sequence alignment. It is, therefore, not surprising that both perform with similar efficiencies in incorporation and termination assays. Our nucleotide selectivity assays, however, revealed significantly lower rates for TTP and HOMedU-TTP using Terminator polymerase compared with Vent(exo⁻) polymerase. Terminator polymerase contains the A48SL variant, which is analogous to the modified Vent(exo⁻) A48SL polymerase. Gardner and Jack (49) suggested that the modified Vent(exo⁻) A48SL polymerase is more tolerant of base-modified nucleotides, but with the trade-off of lower nucleotide selectivity. Here, Terminator II and Vent(exo⁻) polymerase incorporation of T1 with efficiency

*To whom correspondence should be addressed. Email: metzker@bcm.tmc.edu

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Published online 18 September 2007

Nucleic Acids Research, 2007, Vol. 35, No. 19 6339-6349
doi:10.1093/nar/gkm189

Termination of DNA synthesis by N⁶-alkylated, not 3'-O-alkylated, photocleavable 2'-deoxyadenosine triphosphates

Weidong Wu¹, Brian P. Stupi¹, Vladislav A. Litosh¹, Dena Mansouri¹, Demetra Farley¹, Sidney Morris^{1,2}, Sherry Metzker¹ and Michael L. Metzker^{1,2,3,*}

¹LaserGen, Inc., Houston, TX 77054, ²Department of Molecular & Human Genetics and ³Human Genome Sequencing Center, Baylor College of Medicine, Houston, TX 77030, USA

Received July 25, 2007; Revised August 17, 2007; Accepted August 20, 2007

ABSTRACT
The Human Genome Project has facilitated

species, yet the current expensive, labor-intensive, and error-prone process of sequencing remains on mass. Of the various methods, the most efficient is the use of the Illumina sequencing-by-synthesis (SBS) technology. The SBS technology is a novel paradigm of a photocleavable 2'-deoxyadenosine triphosphate (dATP), which, upon use in DNA synthesis, the 2'-nitrobenzyl-dATP remains attached to the DNA strand, preventing further incorporation of dATP. This property is desirable for several commercial available DNA polymerases while maintaining good discrimination against mismatch incorporation. Upon removal of the 2-nitrobenzyl group with light, the natural nucleotide is restored with molecular scanning. A five-base experiment, illustrating the exquisite, stepwise addition through homopolymer repeat, demonstrates the applicability of this technology for high-throughput sequencing.

6348 Nucleic Acids Research, 2007, Vol. 35, No. 19

efficiency (C₉₈) of 92.7% on a solid support. The estimated signal 'S', calculated from the equation S = (C₉₈)ⁿ (15), is 68.4% for the five-base read. At cycle five, the signal for the correct +1 product is 40% (i.e. 1.5 × 10⁻¹), the difference of which we attribute to photobleaching of the dye-primer with increasing exposure to the UV light (data not shown). Signal loss is also due to dephasing, observed as n-1 (incomplete dephasing) and n+1 products (natural nucleotide carryover from the previous cycle). While ongoing efforts are focused on reducing dephasing products, >80% of the total signal is derived from the correct +1 product, with base-calling easily performed from the primary data in Figure 5A and B.

DISCUSSION

We have discovered that the attachment of a small, photocleavable 2-nitrobenzyl group to the 3'-position of 2-deoxyadenosine results in this triphosphate acting as a RT 'without blocking the 3'-end'. The novel RT, N⁶-(2-nitrobenzyl)-dATP (T1), provides favorable enzymatic properties with a variety of wild-type and mutant DNA polymerases. This is unlike the situation for 3'-O-allyl-dATP, which typically acts as a poor substrate for DNA polymerases. For example, screening 3'-O-allyl-dATP with eight different DNA polymerases revealed initial activity at high micromolar concentrations with only Vent(exo⁻) DNA polymerase (8). The

differing from that of the traditional 7-deaza position of BigDye terminators (34) and 3'-O-allyl terminators (7,10). Upon chemical deprotection of the 3'-O-allyl terminators, a residual propargyl amino group remains on the nucleobase, resulting in an accumulating molecular scar with subsequent CRT cycles. The primary advantage of N⁶-alkylation is that, upon directed photocleavage with 365 nm UV light, the modified nucleotide is transformed back into its natural state without molecular scarring (Figure 2D). The enhanced enzymatic properties and the N⁶-cleavage site, transforming the efficiently incorporated RT back into natural DNA, are anticipated to improve the cycle efficiency and read-length of the CRT method.

These observations suggest that the N⁶-(2-nitrobenzyl)-dATP (T1) represents an ideal candidate as a RT for CRT sequencing, illustrated by superior, single base addition through a homopolymer repeat. Of the polymerases tested here, however, not all exhibited this property, with the Family B polymerases revealing incorporation of a second modified nucleotide, albeit at varying levels. Recently, we have discovered that substitution of the 2-nitrobenzyl group can also 'tune' the termination properties of these polymerases to give exclusively single-base products (unpublished data). To exploit the application of 3'-unblocked RTs in CRT sequencing, efforts are underway to create fluorescently labeled analogs of compound T1, and extrapolate this nucleotide model to the remaining nucleobases for production of a novel, four-

DNA polymerases. This is unlike the situation for 3'-modified nucleotides, which typically act as poor substrates for DNA polymerases. For example, screening 3'-O-allyl-dATP with eight different DNA polymerases revealed limited activity at high micromolar concentrations with only Vent(exo⁻) DNA polymerase (8). The

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Ex. 2025, Metzker (2011), 10

Ex. 1017, Metzker (2007), 6348

Columbia's Use of Metzker's Later Papers Is Proper

Yeda Research v. Mylan Pharmaceuticals Inc., 906 F.3d 1031 (2018)

128 U.S.P.Q.2d 1314

906 F.3d 1031
United States Court of Appeals, Federal Circuit.

YEDA RESEARCH and
Development Co., Ltd., Appellant
v.
MYLAN PHARMACEUTICALS INC.,
Amneal Pharmaceuticals LLC, Appellees

2017-1594

2017-1595

2017-1596

Decided: October 12, 2018

Synopsis

Background: Assignee of three patents for a treatment for relapsing-remitting multiple sclerosis sought review of Patent Trial and Appeal Board's (PTAB) final written decisions on inter partes review finding the claims in patents unpatentable as obvious.

Holdings: The Court of Appeals, Reyna, Circuit Judge, held that:

assignee was given proper notice and opportunity to respond to evidence of study of less frequent dosing of treatment;

expert's reliance on study that was published after priority date of patents was permissible;

PTAB's error in relying on study that was published after priority date of patents was harmless;

substantial evidence supported PTAB's finding that regimen of 40 milligram glatiramer acetate (GA) injected three times per week was obvious;

substantial evidence supported PTAB's finding that a person of ordinary skill in the art had reason to switch from a regimen of 20 milligrams GA injected once per day to 40 milligrams GA injected three times per week;

substantial evidenced supported PTAB's finding that claim requiring that claimed method reduce the frequency of reactions was obvious; and

PTAB did not err in finding that claims that identified the specific days that injections was obvious.

Affirmed.

*1034 Appeals from the
Trademark Office, Patent
Nos. IPR2015-00643, IPR2
IPR2015-01976, IPR2015-01

Attorneys and Law Firms

William M. Jay, Goodwin Procter
argued for appellant. Also
James, II; Elizabeth Holland
Wiesen, Boston, MA; John C.
LLP, Washington, DC; Les
NY.

David Lee Anstaett, Perkins
argued for appellees. Appel
Inc. also represented by St
Swanson, Brandon Michael
Dan L. Bagatell, Hanover
McCullough, Seattle, WA.

Anthony James Fitzpatrick,
MA, for appellee Amneal
represented by Vincent Capu
Patrick Gallagher, Boca Ra

Before Reyna, Bryson, and Stoll, Circuit Judges.

Opinion

Reyna, Circuit Judge.

*1035 In this consolidated appeal, Appellant Yeda Research & Development Co., Ltd. challenges the Patent Trial and Appeal Board's final written decisions finding the claims of U.S. Patent Nos. 8,232,250, 8,399,413, and 8,969,302 unpatentable as obvious in three *inter partes* review proceedings. We affirm the Board's decisions.¹

the challenge”). The Board has recognized that non-prior art evidence of what was known “cannot be applied, independently, as teachings separately combinable” with other prior art, but “can be relied on for their proper supporting roles, e.g., indicating the level of ordinary skill in the art, what certain terms would mean to one with ordinary skill in the art, and how one with ordinary skill in the art would have under-stood a prior art disclosure.” *Dominion Dealer Sols., LLC v. AutoAlert,*

Yeda Res. v. Mylan Pharm.,
906 F.3d 1031, 1041 (Fed. Cir. 2018)
(internal citations omitted)

Illumina's Contention That SBS Requirements Changed Over Time Is Belied by the Evidence

Stephen Menchen, Ph.D.
Deposition

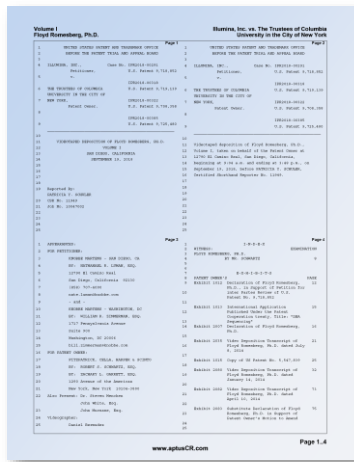
- Q. You are aware that standards for what's efficient incorporation of DNA analogs changes over time as technology improves?
- A. Yeah, I -- I understand what you are asking. I can't think of a specific example of that happening.

Ex. 1113, Menchen Dep., 369:21-370:6

initiation of chain growth. Moreover, this technique is plagued by any inefficiencies of incorporation and deprotection. Because incorporation and 3'-OH regeneration are not completely efficient, a pool of initially identical extending strands can rapidly become asynchronous and sequences cannot be resolved beyond a few limited initial additions.

Ex. 2013, Stemple (2000), 2-3

Metzker Did Not Teach That the 3'-O-Allyl-dATP Had Useful Properties as an Alternative Terminator



Floyd Romesberg, Ph.D.

Deposition

Q. Metzker does not say that the 3'-O-allyl dNTP was shown to terminate DNA synthesis and may be used as an alternative terminator in Sanger sequencing, right?

A. He does not say that.

Ex. 2126, Romesberg Dep., 106:10-107:2

Metzker Abandoned the 3'-O-Allyl-dATP

© 1994 Oxford University Press
Nucleic Acids Research, 1994, Vol. 22, No. 20, 4263-4265

Termination of DNA synthesis by novel 3'-modified-deoxyribonucleoside 5'-triphosphates

Michael L Metzker*, Ramesh Raghavachari*, Stephen Richards, Swamee E Jacutin*, Andrew Ciolek, Kevin Burgess* and Richard A Gibbs
Department of Molecular and Human Genetics, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030 and *Department of Chemistry, Texas A & M University, College Station, TX 77843, USA

Received June 3, 1994; Revised and Accepted July 25, 1994

ABSTRACT
Eight 3'-modified-dNTPs were synthesized and tested in two different DNA template assays for incorporation activity. From this screen, three 3'-O-allyl-dNTPs were shown to terminate DNA synthesis mediated by a number of polymerases and may be used as alternative terminators in Sanger sequencing. 3'-O-(2-nitrobenzyl)-dATP is a UV sensitive nucleoside and was shown to be incorporated by several thermostable DNA polymerases. Base specific termination and efficient photolytic removal of the 3'-protecting group was demonstrated. Following incorporation, DNA synthesis was monitored by the incorporation of natural nucleotides into DNA. The identification of this stable terminator and the demonstration of a new cycle stopper DNA synthesis are initial steps in the development of a novel sequencing strategy.

INTRODUCTION
2'-Deoxythymidine-5'-triphosphate (dTTP) modified at the 3'-hydroxyl position can act as terminators of sequence-dependent DNA synthesis (1-12). These nucleoside analogs are useful as DNA sequencing units, mechanistic probes, mutagenesis, and as antiviral agents. Consequently, such compounds have been used in chemical and biological studies (13). Overall, however, the number of compounds that are well characterized in vitro and in vivo is limited. In the past, the use of terminators and polymers has been limited by the lack of a 3'-O-allyl-dNTP. The 3'-O-allyl-dNTPs are incorporated by a number of polymerases and are incorporated into DNA. The identification of this stable terminator and the demonstration of a new cycle stopper DNA synthesis are initial steps in the development of a novel sequencing strategy.

Among the most useful terminators of DNA synthesis are the 2', 3'-dideoxynucleoside-5'-triphosphates (ddNTPs) that are the basis for Sanger DNA sequencing (13). In this method, oligonucleotide-primed DNA or RNA synthesis are enzymatically extended to 3' - 5' direction in the presence of a mixture of ddNTPs and dNTPs to generate a population of nucleic acids that are terminated at specific base positions. DNA fragments of different lengths are resolved by denaturing polyacrylamide gel electrophoresis and detected either by radioactivity or by the use of fluorescently labeled nucleotides. Despite the limitations of gel electrophoresis for sequencing large amounts, this method has been the favored approach for many years (13,14).

Improvements to the Sanger protocols are being sought to increase the efficiency of large scale sequencing of whole genomes (14). We and others (15-18) have independently conceived a radically different, gel-free alternative to the Sanger protocol for DNA sequencing. This method, called Address Sequencing Scheme (BACS), is based on the sequential incorporation of fluorescently labeled nucleotides along the growing DNA strand. BACS requires repetitive cycles of incorporation of each nucleotide in the sequence to identify the incorporated nucleotide. In contrast to Sanger sequencing, BACS has the advantage that nucleotides would not require gel electrophoresis and there is no need for a sequencing primer. The complete scheme demands a multiplex that are separated by polymerase, specific distinct for each base, while during the polymerization and subsequent efficient and rapid detection in solution. These stringent requirements are formidable for the design and synthesis of the required nucleotides and the development of the sequencing process itself.

The incorporation of nucleotides is mediated by a variety of polymerases. The majority of 3'-modified nucleotides have been reported to be incorporated by DNA polymerase β . Consequently, eight 3'-modified-dNTPs were synthesized and tested for their ability to terminate DNA synthesis by a variety of polymerases. The majority of 3'-modified nucleotides have been reported to be incorporated by DNA polymerase β . Some combinations of terminators were identified using two different polymerase systems. One of these compounds, 3'-O-(2-nitrobenzyl)-dATP (1), was used to demonstrate one complete cycle of termination, degradation, and reinitiation of DNA synthesis.

*To whom correspondence should be addressed.
*Present address: LACOR Inc., Biotechnology Division, Lincoln, NE 68503, USA

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for incorporation by a variety of polymerases. Three compounds, 3'-O-methyl-dATP [1], 3'-O-methyl-dTTP [8], and 3'-O-(2-nitrobenzyl)-dATP [7], proved to be interesting DNA synthesis terminators. In Sanger sequencing, the 3'-O-methyl analogs

an alteration in enzymatic fidelity is discussed below. Compounds [1], [8], and [7] showed specific termination and were further evaluated with respect to their concentration dependent effects.

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2. METZKER, MICHAEL L. and RAGHAVACHARI, RAMESH. 1994. Nucleic Acids Research, 22(20):4263-4265.

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169. METZKER, MICHAEL L. and RAGHAVACHARI, RAMESH. 1994. Nucleic Acids Research, 22(20):4263-4265.

170. METZKER, MICHAEL L. and RAGHAVACHARI, RAMESH. 1994. Nucleic Acids Research, 22(20):4263-4265.

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Columbia's Patent "Admission" Says Nothing More Than Metzker Itself Says about the 3'-O-Allyl-dATP

Columbia Patent

sequencing projects. Consequently, several groups have investigated such a system with an aim to construct an ultra high-throughput DNA sequencing procedure (Cheeseman 1994, Metzker et al. 1994). Thus far, no complete success of using such a system to unambiguously sequence DNA has been reported. The pyrosequencing approach that employs

polymerases (Axelrod et al. 1978). 3'-O-allyl-dATP was also shown to be incorporated by Ventr(exo-) DNA polymerase in the growing strand of DNA (Metzker et al. 1994).

Ex. 1075, '985 Patent,
2:27-32, 3:28-30*

Metzker 1994

incorporation. First, the protecting groups containing ether linkages at the 3'-position, compounds [1], [3], [4], [7], and [8] were incorporated by some of the polymerases. With the

Ex. 1016, Metzker (1994), at 4265

Illumina's Solexa Patent Application Credits Columbia With Discovering $-\text{CH}_2-\text{CH}=\text{CH}_2$ Capping Group

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(71) Applicant (as filed):	SEE BELOW LIMITED (GB/GB), Chesham Research Park, Little Chesham, Nr. Suffolk, Wakenay, UK (GB)
(72) Inventors and	
Inventors/Applicants (for US only):	MILLTON, John (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); WU, Xiaohu (CN/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); SMITH, Mark (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); BRENNAN, Joseph (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); BARNES, Colin (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); LI, Xiaohu (CN/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB)
(73) Assignors and	
Assignors/Applicants (for US only):	MILLTON, John (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); WU, Xiaohu (CN/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); SMITH, Mark (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); BRENNAN, Joseph (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); BARNES, Colin (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); LI, Xiaohu (CN/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB)
(74) Attorneys and	
Attorneys/Applicants (for US only):	MILLTON, John (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); WU, Xiaohu (CN/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); SMITH, Mark (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); BRENNAN, Joseph (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); BARNES, Colin (GB/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB); LI, Xiaohu (CN/GB); Sokoła, Linda (GB/GB); Sokoła, Robert, Pk, Little Chesham, Nr. Suffolk, Wakenay, Essex CB10 1XJ (GB)
(54) Title: MODIFIED NUCLEOTIDES	
(57) Abstract: The invention provides modified nucleotides or nucleoside mixtures that show or describe their sugar moiety having a removable 3' blocking group when attached to a group of the structure O^2 where O^2 is any of C^1 , C^2 , C^3 , C^4 , C^5 , C^6 , C^7 , C^8 , C^9 , C^{10} , C^{11} , C^{12} , C^{13} , C^{14} , C^{15} , C^{16} , C^{17} , C^{18} , C^{19} , C^{20} , C^{21} , C^{22} , C^{23} , C^{24} , C^{25} , C^{26} , C^{27} , C^{28} , C^{29} , C^{30} , C^{31} , C^{32} , C^{33} , C^{34} , C^{35} , C^{36} , C^{37} , C^{38} , C^{39} , C^{40} , C^{41} , C^{42} , C^{43} , C^{44} , C^{45} , C^{46} , C^{47} , C^{48} , C^{49} , C^{50} , C^{51} , C^{52} , C^{53} , C^{54} , C^{55} , C^{56} , C^{57} , C^{58} , C^{59} , C^{60} , C^{61} , C^{62} , C^{63} , C^{64} , C^{65} , C^{66} , C^{67} , C^{68} , C^{69} , C^{70} , C^{71} , C^{72} , C^{73} , C^{74} , C^{75} , C^{76} , C^{77} , C^{78} , C^{79} , C^{80} , C^{81} , C^{82} , C^{83} , C^{84} , C^{85} , C^{86} , C^{87} , C^{88} , C^{89} , C^{90} , C^{91} , C^{92} , C^{93} , C^{94} , C^{95} , C^{96} , C^{97} , C^{98} , C^{99} , C^{100} , C^{101} , C^{102} , C^{103} , C^{104} , C^{105} , C^{106} , C^{107} , C^{108} , C^{109} , C^{110} , C^{111} , C^{112} , C^{113} , C^{114} , C^{115} , C^{116} , C^{117} , C^{118} , C^{119} , C^{120} , C^{121} , C^{122} , C^{123} , C^{124} , C^{125} , C^{126} , C^{127} , C^{128} , C^{129} , C^{130} , C^{131} , C^{132} , C^{133} , C^{134} , C^{135} , C^{136} , C^{137} , C^{138} , C^{139} , C^{140} , C^{141} , C^{142} , C^{143} , C^{144} , C^{145} , C^{146} , C^{147} , C^{148} , C^{149} , C^{150} , C^{151} , C^{152} , C^{153} , C^{154} , C^{155} , C^{156} , C^{157} , C^{158} , C^{159} , C^{160} , C^{161} , C^{162} , C^{163} , C^{164} , C^{165} , C^{166} , C^{167} , C^{168} , C^{169} , C^{170} , C^{171} , C^{172} , C^{173} , C^{174} , C^{175} , C^{176} , C^{177} , C^{178} , C^{179} , C^{180} , C^{181} , C^{182} , C^{183} , C^{184} , C^{185} , C^{186} , C^{187} , C^{188} , C^{189} , C^{190} , C^{191} , C^{192} , C^{193} , C^{194} , C^{195} , C^{196} , C^{197} , C^{198} , C^{199} , C^{200} , C^{201} , C^{202} , C^{203} , C^{204} , C^{205} , C^{206} , C^{207} , C^{208} , C^{209} , C^{210} , C^{211} , C^{212} , C^{213} , C^{214} , C^{215} , C^{216} , C^{217} , C^{218} , C^{219} , C^{220} , C^{221} , C^{222} , C^{223} , C^{224} , C^{225} , C^{226} , C^{227} , C^{228} , C^{229} , C^{230} , C^{231} , C^{232} , C^{233} , C^{234} , C^{235} , C^{236} , C^{237} , C^{238} , C^{239} , C^{240} , C^{241} , C^{242} , C^{243} , C^{244} , C^{245} , C^{246} , C^{247	

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WO02/29003 (The Trustees of Columbia University in the City of New York) describes a sequencing method which may include the use of an allyl protecting group to cap the 3'-OH group on a growing strand of DNA in a polymerase reaction. The allyl group is introduced according to the procedure of Metzker (*infra*) and is said

compatible, chemistry.

Metzker et al., (Nucleic Acids Research, 22(20): 4259-4267, 1994) discloses the synthesis and use of eight 3'-modified 2-deoxyribonucleoside 5'-triphosphates (3'-modified dNTPs) and testing in two DNA template assays for incorporation activity. The 3'-modified dNTPs included 3'-allyl deoxyriboadenosine 5'-triphosphate (3'-allyl dATP). However, the 3'-allyl blocked compound was not used to demonstrate a complete cycle of termination, deprotection and reinitiation of DNA synthesis: the only test results presented were those which showed the ability of this compound to terminate DNA synthesis in a single termination assay, out of eight such assays conducted, each conducted with a different DNA polymerase.

WO02/29003 (The Trustees of Columbia University in the City of New York) describes a sequencing method which may include the use of an allyl protecting group to cap the 3'-OH group on a growing strand of DNA in a polymerase reaction. The allyl group is introduced according to the procedure of Metzker (*infra*) and is said

Ex. 1125, 3

Dr. Romesberg in 2005 Credited Columbia With Pioneering the Allyl-Capping Group

Principal Investigator/Program Director (Last, First, Middle): Romesberg, Floyd E.

A. Specific Aims

This R21 submission is in response to PA-03-058, 'Exploratory/Developmental Bioengineering Research Grants.' We propose the directed evolution of new DNA polymerases that will facilitate several emerging sequencing technologies. After successful completion of the proposed research we will seek additional funding to develop the most promising applications.

There is great incentive for developing low cost technologies for sequencing genomes. During the past two decades, along with computational advances, the parallelization, automation, and refinement of the Sanger method has reduced sequencing costs from tens of dollars per sequenced base to hundredths of a cent per sequenced base. However, the cost of sequencing a mammalian-sized genome is still tens of millions of dollars, and it is unlikely that further advances in instrumentation alone will achieve the 10,000-fold reduction in cost set as a goal by the NIH.

A variety of new sequencing methods currently in development have the potential to make genome sequencing commonplace. Most of these new methods rely on polymerase mediated DNA synthesis with modified deoxynucleotide triphosphate (dNTP) substrates. As a result, these DNA sequencing methods are all limited by the ability of the available polymerases to recognize modified dNTPs. Despite obvious limitations with the polymerases, and in contrast to the great effort devoted towards improvements in automation and new technique development, little effort has been directed toward using the modern methods of directed evolution to evolve enzymes that are better suited to sequencing.

We have developed an activity-based selection system to evolve polymerases with novel activities (Figure 1). As described in detail below, the selection system involves the co-display on phage of DNA polymerase libraries and an oligonucleotide substrate that primes synthesis with an attached template. If a displayed polymerase mutant recognizes the attached substrate and the provided dNTPs, then it will extend the primer, and only then covalently attach a biotin-dUTP. Biotinylated phage particles may be selectively recovered using a streptavidin solid support. We have successfully used the system to evolve several polymerases that catalyze unnatural reactions with efficiencies that rival those of natural polymerases with their native

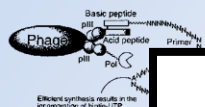


Figure 1. Phage particle with attached substrate. X represents a natural substrate. Only two phage proteins are designed so that they incorporate DNA synthesis with natural or modified substrates [1,2]. We are now interested in evolving polymerases that improve different sequencing methods. The proposal focuses on evolving the *Thermus aquaticus* polymerase I fragment of *E. coli* DNA Polymerase I as a backup in the event that unforeseen problems arise with Sf. The specific aims of this proposal are:

Specific Aim 1, Year 1: Evolve polymerases that efficiently synthesize DNA using Cy5-labeled dNTPs using novel methodologies.

Specific Aim 2, Year 2: Evolve polymerases that efficiently synthesize DNA using 3'-O-allyl protecting groups to continue synthesis after allyl group removal.

Specific Aim 3, Year 2: Collaborate to evaluate evolved polymerases in actual sequencing applications.

The overall goal of the proposed research is the directed evolution of DNA polymerases to enable revolutionary sequencing methodologies. To facilitate the last specific aim, we have established collaborations with Prof. Steve Quake (Stanford), Prof. Jingyue Ju (Columbia), and Dr. Phillip Ordoxian, director of the Scripps sequencing facility.

B. Background and Significance

Currently, Sanger dideoxy sequencing and capillary electrophoresis is the most well-established and popular technique employed for DNA sequencing.

Principal Investigator/Program Director (Last, First, Middle): Romesberg, Floyd E.

The Ju group has pioneered the development of novel dNTPs that not only bear photo-detachable nucleobase-linked fluorophores (see above), but that also bear 3'-OH protecting groups. First proposed

Columbia has reported a set of four dyes, one linked to each dNTP by a photolabile group that was removed by Thermo Sequenase. Additionally, the approach was also demonstrated for the addition of dyes to

Specifically, the Ju group has developed 3'-O-allyl protecting groups that may be catalytically removed with palladium [11]. However, like all 3'-O-

concentrations and extended incubation periods to drive extension of the 'deprotected' primer termini.

The Ju group has pioneered the development of novel dNTPs that not only bear photo-detachable nucleobase-linked fluorophores (see above), but that also bear 3'-OH protecting groups. First proposed by Metzger in 1994 [10], reversible modification of the 3'-OH allows for strict control over the synthesis beyond the modified nucleotide since the required nucleophile is not available until deprotection. When optimized, 3' modification should allow all four labeled dNTPs to be present in a given reaction, as opposed to flushing the system with solutions containing only one labeled dNTP at a time. In addition, these protecting groups would prevent multiple incorporations of the same dNTP (modified or unmodified) at homopolymeric runs. Specifically, the Ju group has developed 3'-O-allyl protecting groups that may be catalytically removed with palladium [11]. However, like all 3'-O-modified dNTPs examined, as well as the nucleobase-modified dNTPs discussed above, these 3'-protected dNTPs are very inefficient substrates

Function.

Many 'Type I' (or 'A family') polymerases have played an important role in sequencing efforts, including both Taq (from which Sf is derived) and Kf [12]. Careful consideration of the structure and substrate repertoire of these polymerases is expected to facilitate their directed evolution.

(B3.1) Type I DNA Polymerases Structure and Mechanism. In addition to being the polymerases of choice for sequencing, Type I DNA polymerases are an attractive starting point for directed evolution experiments due to their small size, thorough characterization [13], and structural and functional homology [14,15].

Crystal structure data for each of these type I polymerases shows a structure resembling a right-hand with 'finger,' 'palm,' and 'thumb' subdomains forming a deep substrate binding cleft (Figure 2). The duplex DNA approaches the polymerase domain from the 3'-5' exonuclease side of the cleft and binds with the primer terminus in the polymerase active site, packed against the N-

PHS 398/290 (Rev. 09/04)

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Continuation Format Page

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Continuation Format Page

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Ex. 2118, Romesberg NIH Grant, 29

Illumina's Belated Contamination, High Concentration, and Vent Polymerase Variant Theories Fail

Relevant to:

IPR2018-00291, 318, 322, 385: Grounds 1-2

IPR2018-00797: Grounds 1-3

Contamination Did Not Cause Metzker's Termination*: Metzker's Assays Differentiate Incorporation and Contamination

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Termination of DNA synthesis by novel 3'-modified-deoxyribonucleoside 5'-triphosphates

Michael L. Metzker*, Ramesh Raghavachari¹, Stephen Richards, Swanee E. Jacutin¹, Andrew Oviello, Kevin Burgess¹ and Richard A. Gibbs
Department of Molecular and Human Genetics, Baylor College of Medicine, One Baylor Plaza, Houston, TX 77030 and ¹Department of Chemistry, Texas A & M University, College Station, TX 77843, USA

Received June 3, 1994; Rev.

ABSTRACT

Eight 3'-modified-dNTPs in two different DNA templates

(4-nitrobenzoyl)-dATP [6]. The yield was reduced in that case to 57% due probably to a rearrangement of the 4-nitrobenzoyl group from the 3'-position to the 5'-position (data not shown).

Nucleic Acids Research, 1994, Vol. 22, No. 20 4263

could not be identified. In each of the Oligo-template gels, the reactions contained all the required nucleotides except the natural nucleotide corresponding to the analog tested. The samples

[1] by AMV-RT. In addition to termination, some readthrough was also observed due to the presence of contaminating dATP. All RP-HPLC purified 3'-modified-dATPs (compounds [1]–[7]) showed approximately 1% dATP contamination, and these trace levels could not be removed by subsequent RP-HPLC.

Among the most familiar is the 2', 3'-dideoxynucleosides are the basis for Sanger DNA oligonucleotide-primed DNA cally extended to a 5'–3' din of dNTPs and dNTPs to gene are terminated at specific but different lengths are resolved)

remaining natural nucleotides, so that other aspects of the enzyme performance could be identified. We found that all the polymerases misincorporated other dNTPs in the absence of the complement dNTP, and this nucleotide readthrough was concentration dependent. Thus, minimum dNTP concentrations that gave efficient incorporation, but no apparent misincorporation were first defined in this assay. These dNTP concentrations were then used to determine the minimum dNTP concentration that yielded complete termination. *Pfu* (exo⁺) DNA polymerase was excluded from the Oligo-template assay since a dNTP concentration that yielded complete termination for this enzyme

that 3'-O-methyl-dATP [1] is approximately 200 to 250-fold less efficiently incorporated by AMV-RT than dATP (compare lanes 6 and 8; 5 mM and 1 mM, respectively). In Figure 3B, the Oligo-template gel also shows the incorporation of 3'-O-methyl-dATP [1] by AMV-RT. In addition to termination, some readthrough was also observed due to the presence of contaminating dATP. All RP-HPLC purified 3'-modified-dATPs (compounds [1]–[7]) showed approximately 1% dATP contamination, and these trace levels could not be removed by subsequent RP-HPLC. 3'-O-Methyl-dATP [1] was also incorporated by M-MuLV-RT and inhibited DNA synthesis by rTth and Ventre (exo⁺)

*To whom correspondence should be

[†] Present address: LICOR Inc., Bo

Table 2. Activity assays of RP-HPLC purified 3'-terminating dNTPs challenged against commercially available polymerases

3'-modified-dATP (Group compound [X])	AMV-RT	M-MuLV-RT	Klenow fragment	Longhorn [†]	RNase H ⁺ polymerase	AmpliTaq [†] DNA polymerase	Thermus thermophilus DNA polymerase	Therminator [†] DNA polymerase
[2] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[3] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[4] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[5] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[6] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[7] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[8] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[9] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[10] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[11] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[12] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[13] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[14] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[15] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[16] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[17] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[18] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[19] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[20] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[21] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[22] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
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[29] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
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[99] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination
[100] O-methyl	Termination	Termination	Termination	Termination	Termination	Termination	Termination	Termination

All compounds were assayed at a final concentration of 250 µM according to the conditions specified in Table 1. "–" means no activity was observed. "Termination" means that the termination bands (dNTP termination bands) and "Inhibition" means the rate of DNA synthesis is reduced in a non-specific manner. * means the activity was incorporated at a final concentration of 250 µM.

Short Technical Reports

SHORT TECHNICAL REPORTS

Manuscript published in the Short Technical Reports section are shorter and less comprehensive in scope than full Research Reports.

dTTP from RP-HPLC purified 3'-O-methyl-dTTP and 3'-O-methyl-dATP, respectively. These data highlight the importance of natural nucleotide contamination when incorporating synthetic incorporation data and provide an alternative strategy for the detection of dNTP contamination.

ciently high that the DNA polymerase selectively incorporates the preferred natural dTTP over the 3'-O-methyl-dTTP analog. Incorporation of the natural dTTP in the desired termination lane, referred to here as natural-terminated, is the key to the assay.

INTRODUCTION

Characterization of dideoxynucleoside (dNTP) analogs as 1' systems has been demanding of DN general. DNA pol- stered selectively dNTP analogs this fully useful for the trival compounds, potentially have bio- logical applications, large can be used to identify mutants, and to study chemical synthesis. However, the study dNTPs has been pro- ing because the same chemical synthesis compounds in the same chemical, which is one the chemical of the subsequent use is typically involving removal of the co from 3'-O-methyl- Previously, we feasibility of a stop- ing strategy called quencing scheme (thesis and expansion different 3'-O-methyl- dNTPs [1].

lution before incorporation by human immunodeficiency virus type 1 (HIV-1) RT (data not shown) and contaminating dTTP from a 3'-O-methyl-dTTP solution before incorporation by *Pol* DNA polymerase (data not shown). Thus, these data show that dNTP contamination of RP-HPLC 3'-O-methyl-dNTPs can be effectively removed by the enzymatic stop-up assay. Casad et al. (1) have provided evidence for the hydrolysis of 3'-O-methyl linkages by a mechanism of catalytic cleavage of DNA polymerase. Both 3'-O-methyl and 3'-O-methyl-dNTPs protecting groups described in the current study are attached to the sugar ring by ether linkages. The oligo-template assay used here and elsewhere (3) has been designed to differentiate between the incorporation of the natural dNTP (read-through) and the 3'-O-methyl-dNTP (true termination) (Figures 2 and 3). Using this assay, we have observed that the majority of RP-HPLC purified 3'-O-methyl-dNTPs, maintain initial, but significant levels of dNTP contamination. Previously, we evaluated 7 different 3'-O-methyl-dNTPs, 3 of which contained ether linkages that

incorporation data. Here, we have demonstrated that enzymatic stop-up is a viable strategy that uses conventional reagents for the purification of novel RP-HPLC-purified 3'-O-methyl-dNTP analogs. The development of enzymatic stop-up also addresses the important issue of nucleotide read-through in important DNA sequencing strategies such as IAS.

REFERENCES

1. Casad, R., B. Carlson and R.E. Sartin, 1993. Catalytic cleavage of DNA polymerase. *Proc. Natl. Acad. Sci. USA* 90:1005-1009.
2. Casad, R. and R.E. Sartin, 1993. DNA polymerase. *Biochemical reactions with nucleic acids*. *Gene* 141:1-14.
3. Metzker, M.L., K. Raghavachari, S. Richards, R.E. Jacobs, A. Gibbs, K. Burgess and R.A. Gibbs, 1994. Termination of DNA synthesis by novel 3'-O-methyl-dNTPs. *Nucleic Acids Res.* 22:4259-4267.

This work was supported in part by Grant 1 R01 HG014249 and 1 R01 AG12144 and grants in R.E. from the Texas Advanced Technology Program and National Insti-

ether linkages. The oligo-template assay used here and elsewhere (3) has been designed to differentiate between the incorporation of the natural dNTP (read-through) and the 3'-O-methyl-dNTP (true termination) (Figures 2 and 3). Using this assay, we have observed

Ex. 1016, Metzker (1994), 4263

Ex. 2131, Metzker (1998), 817

High Nucleotide Concentrations Are Incompatible With SBS

Filed on behalf of:

Patent Owner Illumina Cambridge Ltd.
By: Brenton R. Babcock
William R. Zimmermann
Jonathan E. Bachand
KNOBBE, MARTENS
2040 Main Street, 14th
Irvine, CA 92614
Tel.: (949) 760-0404
Fax: (949) 760-9502
Email: BoxIllum

IPR2013-00266

IPR2013-00266

UNITED STATES

BEFORE THE

INTELLECTUAL

ILLUMINA CAMBRIDGE LTD.

Patent Owner

Case IPR2013-00266 (LMG)
Patent 8,158,346

SECOND DECLARATION

progress).” Ex. 1028 at 3. A person of ordinary skill in the art seeking to develop sequencing-by-synthesis methods would recognize the need to use a nucleotide with a 3’-protecting group that could be incorporated by a polymerase with high fidelity at moderate to low concentrations. Therefore, a person of skill in the art

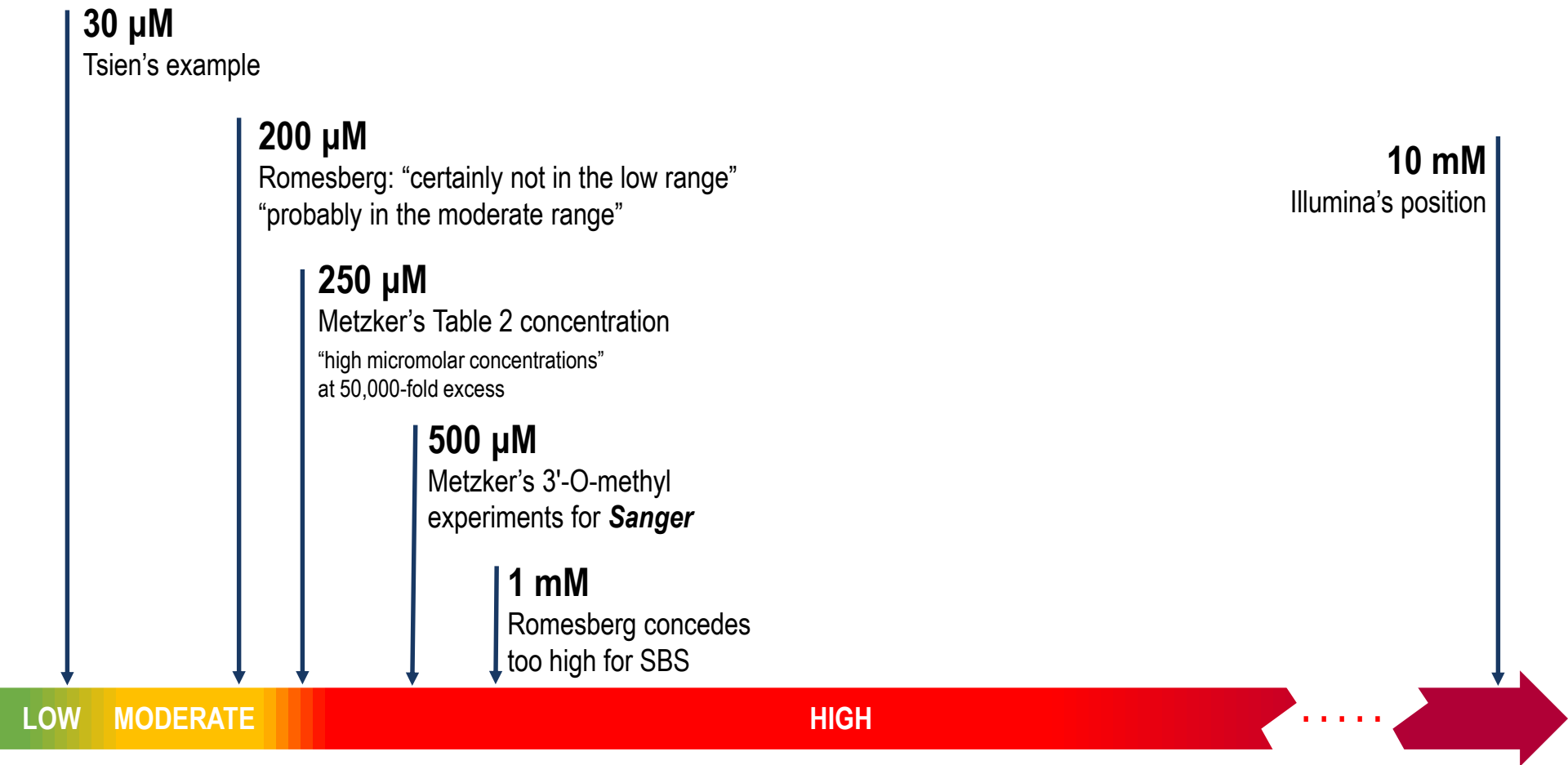
Natl. Acad. Sci. USA, 87:4946-4950 at 4947 (1990) (Ex. 2055); Bebenek et al., J. Biol. Chem., 267:3589-3596 at 3591 (1992) (Ex. 2056). Thus, it is not surprising that Canard taught that “these modified nt are not very efficient chain terminators, a result awaiting precise determination of their K_m , V_{max} and k_{cat} (work in

Ex. 2081,
Romesberg, ¶17

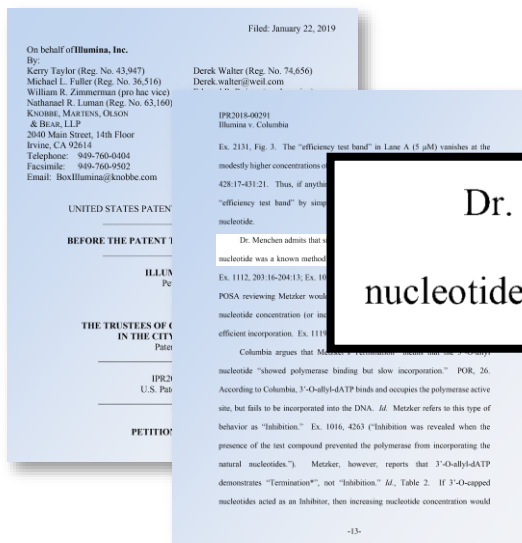
were 1 mM and 2 mM. *Id.* at Fig. 3A. However, polymerases lose the ability to accurately replicate a DNA strand at high nucleotide concentrations because the selectivity of polymerases is hindered at high nucleotide concentrations. *See, e.g.,*

Ex. 2081,
Romesberg, ¶17

High Nucleotide Concentrations Are Incompatible With SBS



Dr. Menchen Did Not Testify That Increasing Concentration Increases Incorporation



Dr. Menchen admits that simply increasing the concentration of 3'-O-capped nucleotide was a known method for achieving efficient polymerase incorporation.

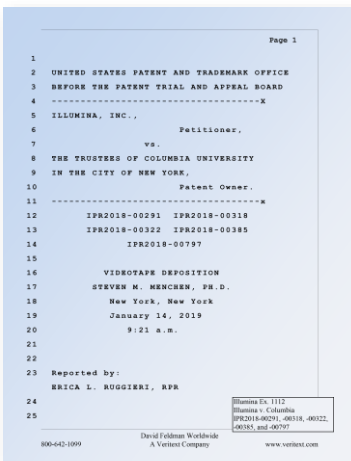
IPR2018-00291, Reply, 13

Stephen Menchen, Ph.D. Deposition

Q. Based on the, your expertise in the Metzker paper, is it your belief that increasing the concentration increases efficiency until you are basically a hundred percent efficient?

A. No. I -- I don't think you can assume that.

Ex. 1112, Menchen Dep., 206:23-207:6



Metzker 1994 Does Not Teach That Increasing Concentration Solves Problem of Inefficient Incorporation

On behalf of Illumina, Inc. By: Kerry Taylor (Reg. No. 43,947) Michael L. Fuller (Reg. No. 38,516) William R. Zimmerman (pro hac vice) Nathaniel R. Luman (Reg. No. 63,160) KNOWLE, MATHESON, OLSON & BEAR, LLP 2040 Main Street, 14th Floor Irvine, CA 92614 Telephone: 949-760-0404 Facsimile: 949-760-9502 Email: Box@illumina@knobbe.com	Derek Walter (Reg. No. 74,656) Derek.walter@icwll.com (IPR)2018-00291 Illumina v. Columbia determine read-length and throughput Ex. 1017, 6348. Similarly, Drs. Ran Ex. 2118, 27. The 2006 statements the NIH's call to dramatically re- Furthermore, post-filing statements positive indicator of the allyl group's methods. <i>Id.</i> 521. 3. A POSA would have Columbia alleges that a POSA because Metzker discloses 250 μM, pM. POR. 30. Tsien's concentra- "especially with derivatized dNTP excesses (over stoichiometry) of v 5592-93. Columbia asserts that "Termal O-allyl-dATP indicates inefficient Metzker's tending to increase the so incorporation efficiency. Ex. 10 increasing the concentration"). Metzker demonstrates that increasing the concentration of a 3'-O-modified-dNTP to 500 μM with Ventrex® polymerase
--	--

O-allyl-dATP indicates inefficient incorporation. POR, 21. Columbia ignores Metzker's teaching to increase the concentration of 3'-O-modified-dNTP to enhance incorporation efficiency. Ex. 1016, 4266 ("problems could be resolved by increasing the concentration"). Metzker demonstrates that increasing the

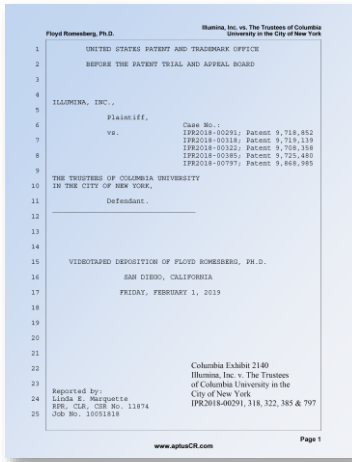
IPR2018-00291, Reply, 16

[illegible]

Several unexpected results were observed in the use of the 3'-*O*-methyl-dNTPs as terminators in DNA sequencing. First, the 3'-*O*-methyl-dTTP [8] reduced enzyme fidelity. Also, we have not identified a polymerase that will incorporate both 3'-*O*-methyl-dATP [1] and 3'-*O*-methyl-dTTP [8]. These problems could be resolved by increasing the concentration of dATP in the former case to titrate away artifact termination bands and by testing more polymerases in the latter case. This issue, however,

Ex. 1016, Metzker (1994), 4266

Vent Variants: Illumina Provides No Evidence That a POSA Would Expect Vent Variants to Efficiently Incorporate 3'-O-Allyl-dATP



Floyd Romesberg, Ph.D. Deposition

Q. And there's no experimental data in Gardner that demonstrates efficient incorporation of a 3'-O-propenyl modified nucleotide with mutated vent (exo-) polymerase, correct?

A. No. What I was suggesting this as is that a POSA would have known that there were mutants of vent that were more tolerant to the 3' position. He would not have known the[y] accepted 3'-O-allyl. My only point in the dec, to be clear, was that a POSA would have known to go look. He would -- he would not have known. He would have known to look and try.

Ex. 2140, Romesberg Dep., 130:12-22

Illumina’s Theories Are Belied by Real-World Evidence



Illumina's Cleavage Condition Arguments Fail

Relevant to:

IPR2018-00291, 318, 322, 385: Grounds 1-2

IPR2018-00797: Grounds 1-3

The Board Requested Briefing Regarding Allyl Ether Cleavage Conditions

Trials@uspto.gov
Tel: 571-272-7822

Paper 20
Entered: September 18, 2018

UNITED STATES PATENT AND TRADEMARK OFFICE

BEFORE THE PATENT TRIAL AND APPEAL BOARD

ILLUMINA, INC.,
Petitioner,

v.

THE TRUSTEES OF COL
IN THE CITY OF
Patent O

Case IPR20
Patent 9,86

Before JAMES A. WORTH, MICHEL
BRIAN D. RANGE, *Administrative Pa*

RANGE, *Administrative Patent Judge*.

DECISION
Institution of *Inter Partes* Review
35 U.S.C. § 314

12:27–29, 24:5–25:3. We invite further briefing on this issue at trial, including whether it was appreciated in the art that sufficiently mild conditions were known for cleaving allyl groups when working with nucleotide analogues. We also invite briefing on (1) whether or not the

IPR2018-00797, Institution Decision, 25-26

No Dispute SBS (BASS) Requires SBS-Compatible Cleavage Conditions

PCT WORLD INTELLECTUAL PROPERTY ORGANIZATION International Bureau	
INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)	
(51) International Patent Classification: C12Q 1/68, C12P 19/34 /C12H 21/00, 21/04, C01N 35/08	(11) International Publication Number: WO 91/06678 (43) International Publication Date: 16 May 1991 (16.05.91)
(31) International Application Number: PCT/US90/06178 (32) International Filing Date: 26 October 1990 (26.10.90)	(74) Agents: CHEN, John, Y. SRI International, 333 Ravenswood Avenue, Menlo Park, CA 94025-3493 (US) et al.
(30) Priority data: 427,321 26 October 1989 (26.10.89) US	(81) Designated States: AT (European patent), BE (European patent), CA, CH (European patent), DE (European patent), DK (European patent), ES (European patent), FR (European patent), GB (European patent), GR (European patent), IT (European patent), JP, LI (European patent), NL (European patent), SE (European patent)
(71) Applicants: SRI INTERNATIONAL (US/US), 333 Ravenswood Avenue, Menlo Park, CA 94025-3493 (US)	Published With international search report Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.
(71)(72) Applicant and inventor: TS'EN, Roger, Y. (US/US), 8335 Pennington Place, La Jolla, CA 92037-2312 (US)	
(72) Inventors: BOSS, Paul; 7451 Contra Costa Avenue, Berkeley, CA 94707 (US); FAHNESTOCK, Margaret; 2728 Gamble Court, Hayward, CA 94542 (US); JOHNSTON, Allan, J.; 909 North California Avenue, Palo Alto, CA 94303 (US)	

Blocking Groups and Methods for Incorporation

The coupling reaction generally employs 3'-hydroxyl-blocked dNTPs to prevent inadvertent extra additions.

The criteria for the successful use of 3'-blocking groups include:

- (1) the ability of a polymerase enzyme to accurately and efficiently incorporate the dNTPs carrying the 3'-blocking groups into the cDNA chain,
- (2) the availability of mild conditions for rapid and quantitative deblocking, and

Ex. 1013, Tsien, 20

schemes may lead to significantly improved throughput. A major challenge in development of BASS, however, is that the 3'-blocking group must be removed in an aqueous medium using reagents/conditions that do not denature or modify double strand DNA. To address this issue, the focus of

NUCLEOSIDES & NUCLEOTIDES, 18(2), 197-201 (1999)

SYNTHESIS OF FLUORESCENT, PHOTOLABILE 3'-O-PROTECTED NUCLEOSIDE TRIPHOSPHATES FOR THE BASE ADDITION SEQUENCING SCHEME

Mike B. Welch and Kevin Burgess*

Department of Chemistry, Texas A & M University, PO Box 300012, College Station, TX 77842-3012

ABSTRACT: The dansylated nucleoside triphosphates **1a** and **1b** were prepared as a prelude to investigating sequencing of DNA via a scheme that does not involve electrophoresis.

Throughput of sequence data is critical in The Human Genome Project and similar ventures. Gel electrophoresis is a "bottle-neck" in high throughput sequencing of DNA hence it is desirable to develop methods that avoid this technique. Several groups have been interested in an approach that we call, "The Base Addition Sequencing Scheme", BASS.¹⁻⁴ This involves modified nucleotides **1** that are: (i) incorporated by DNA replicating enzymes; (ii) spectroscopically distinct (at λ^{*}) so that the parent base (A, T, G, and C) can be differentiated; and, (iii) labile at the 3'-position such that the 3'-hydroxyl terminus of a polynucleotide can be regenerated. This scheme may be slower and sequence less bases per experiment than existing methods but, unlike methods involving gels, there is potential for multiple experiments to be run in parallel. Such combinatorial sequencing schemes may lead to significantly improved throughput. A major challenge in development of BASS, however, is that the 3'-blocking group must be

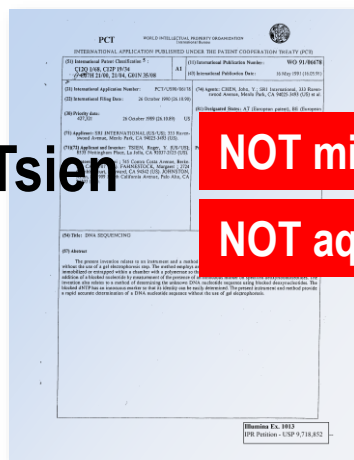
Columbia Exhibit 2041

197 Illumina, Inc. v. The Trustees of
University in the City of
www.dakor.com

Ex. 2041, Welch, 197-98

Illumina's Petition-Cited References Do Not Teach SBS-Compatible Cleavage Conditions

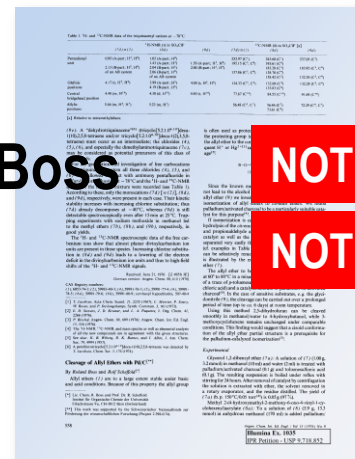
Tsien



NOT mild

NOT aqueous

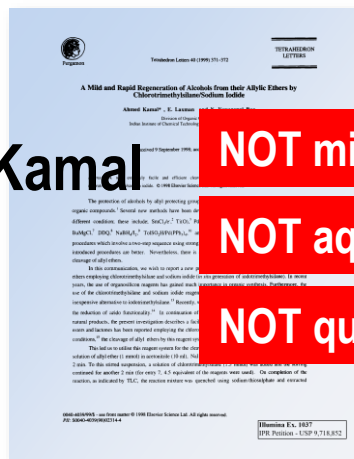
Boss



NOT mild

NOT rapid

Kamal

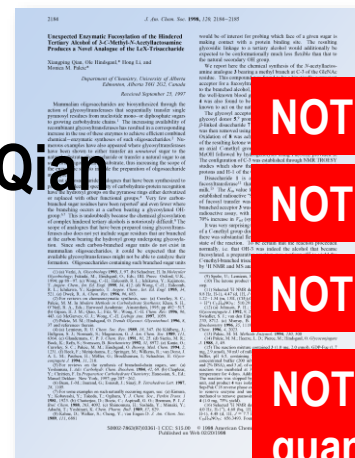


NOT mild

NOT aqueous

NOT quantitative

Qian



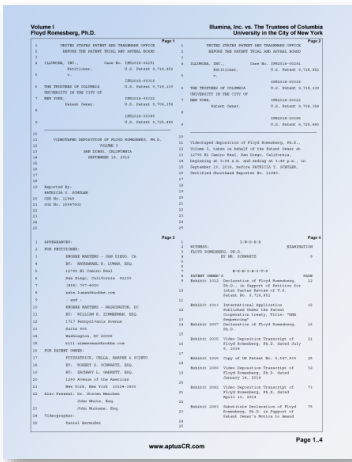
NOT mild

NOT aqueous

NOT rapid

NOT reliably quantitative

Tsien Does Not Teach SBS-Compatible Cleavage Conditions



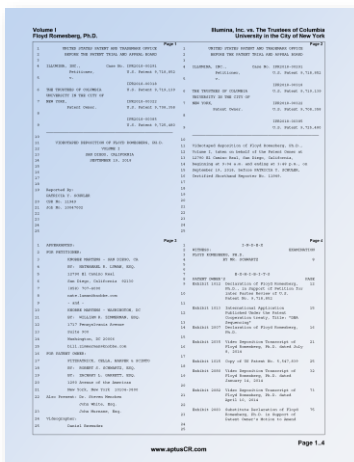
Floyd Romesberg, Ph.D.
Deposition

Q. So a person reading Tsien along with Gigg and Warren would not use Gigg's condition for cleavage of allyl ethers for SBS because there would be no expectation of success using those cleavage conditions, correct?

A. I think that is fair.

Ex. 2126, Romesberg Dep., 232:10-15

Kamal Does Not Teach SBS-Compatible Cleavage Conditions



Floyd Romesberg, Ph.D. Deposition

Q. But [Kamal's] reaction takes place in acetonitrile with **no water present**?

A. Yes. And in that case it would **denature the DNA**. And you would dehydrate the DNA and cause it to denature.

Q. And Kamal's allyl deprotection method **uses chlorotrimethylsilane**, correct?

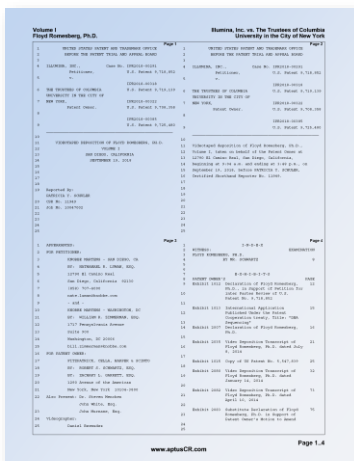
A. It does.

Q. And **chlorotrimethylsilane is not compatible with water**, correct?

A. No, it is not.

Ex. 2126, Romesberg Dep., 234:23-235:8

Boss Does Not Teach SBS-Compatible Cleavage Conditions



Floyd Romesberg, Ph.D. Deposition

Q. So the concentration of perchloric acid that was used to deprotect the allyl in the Boss deprotection reaction **would denature DNA**, correct?

A. The Kit reference suggests that it would.

Q. And that deprotection reaction in Boss **would not be considered a mild condition** that would not denature DNA, correct?

A. That is correct.

* * *

Q. So none of Boss's reaction conditions satisfied Tsien's requirement for **rapid deprotection**, correct?

A. I would agree with that.

Ex. 2126, Romesberg Dep., 190:7-14, 203:17-20

Qian Does Not Teach SBS-Compatible Cleavage Conditions

Volume 1 Floyd Romesberg, Ph.D.		Exhibit, Inc. vs. The Trustees of Columbia University in the City of New York	
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99	99	99	99
100	100	100	100

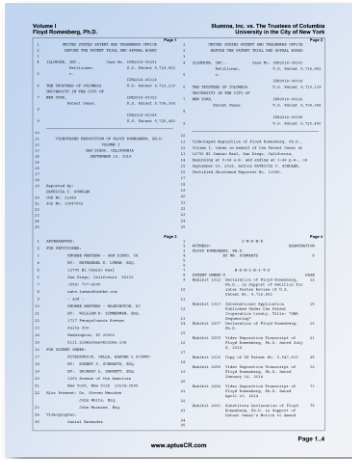
Floyd Romesberg, Ph.D. Deposition

Q. But this reference that you are relying on [Qian] to say that there was quantitative cleavage of the allyl protecting group **uses conditions that are not themselves compatible with SBS**, correct?

A. That is correct.

Ex. 2126, Romesberg Dep., 163:23-164:2

Dr. Romesberg Concedes if Tsien's Cleavage Conditions Are Not Met, It Will Not Work for Tsien's SBS Methods



Floyd Romesberg, Ph.D.
Deposition

Q. So according to Tsien, if any of these criteria are not met, Tsien says it wouldn't be expected to work for his SBS method, correct?

A. Yes.

Ex. 2126, Romesberg Dep., 45:1-4

Illumina's New Qian Argument

Tsien

NOT mild

NOT aqueous

Boss

NOT mild

NOT rapid

Kamal

NOT mild

NOT aqueous

NOT quantitative

Qian

NOT mild

NOT aqueous

**Illumina belatedly argues
Qian could be modified to be aqueous
using 3 Genet references, which are
not for allyl ether cleavage**

Illumina Previously Argued Against Allowing Newly Proposed Cleavage Conditions In Reply Briefing

Trials@uspto.gov
571-272-7822

Paper No. 84
February 2, 2015

illumina

1 RECORD OF ORAL HEARING
2 UNITED STATES PATENT AND TRADEMARK OFFICE
3 -----
4 BEFORE THE PATENT TRIAL AND APPEAL BOARD
5 -----

6 INTELLIGENT BIO-SYSTEMS, INC.

7 Petitioner

8 v.

9 ILLUMINA CAMBRIDGE LIMITED

10 Patent Owner
11 -----

12 Oral Hearing Held: Friday, October 10, 2014

13 Case IPR2013-00517

14 Patent 7,566,537

15
16
17
18
19 BEFORE: LORA M. GREEN, FRANCISCO C. PRATS, and
20 SCOTT E. KAMHOLZ, Administrative Patent Judges.

21 The above-entitled matter came on for hearing on
22 Friday, October 10, 2014, at 10:01 a.m., Hearing Room A, at
23 the U.S. Patent and Trademark Office, 600 Dulany Street,
24 Alexandria, Virginia.

25 Columbia Exhibit 2032
Illumina, Inc. v. The Trustees of
University in the City of New York
IPR2018-00291

Now, I would like to spend some time on the issue of optimization, because you heard here today that one of skill in the art could easily change Zavgorodny's removal conditions and optimize.

This is an argument that came out on reply for the first time. It is an improper new prima facie case. If you

* * *

And so they are limited to the conditions in the petition. The petition didn't say one word about optimization. It didn't say one word about changing the conditions. This is a completely new argument on reply.

Ex. 2032, Illumina, Oral Hearing Tr., 46:5-10, 20-23

Intelligent Bio-Systems, Inc. v. Illumina Cambridge Ltd.,
821 F.3d 1359, 1369 (Fed. Cir. 2016) (Federal Circuit agreeing)

Illumina Previously Argued That Modifying Cleavage Conditions Is Difficult

Trials@uspto.gov
571-272-7822

Paper No. 84
February 2, 2015

1 RECORD OF ORAL HEARING
2 UNITED STATES PATENT AND TRADEMARK OFFICE
3 -----
4 BEFORE THE PATENT TRIAL AND APPEAL BOARD
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6
7 INTELLIGENT BIO-SYSTEMS, INC.

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9 v.

10 ILLUMINA CAMBRIDGE LIMITED

11 Patent Owner

12 -----
13 Oral Hearing Held: Friday, October 10, 2014

14
15 Case IPR2013-00517

16 Patent 7,566,537

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18
19 BEFORE: LORA M. GREEN, FRANCISCO C. P.
20 SCOTT E. KAMHOLZ, Administrative Patent Judges.

21
22 The above-entitled matter came on for hearing on
23 Friday, October 10, 2014, at 10:01 a.m., Hearing Room A
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25 Alexandria, Virginia.

Columbia Exhibit 2032
Illumina, Inc. v. The Trustees of Columbia
University in the City of New York
IPR2018-00291

illumina

Moreover, IBS didn't give you any test data.

There is no test results that says, oh, here are the tests that show you can very easily optimize Zavgorodny's removal conditions. They didn't present them because they know how difficult that is. This just isn't what happens in the real world.

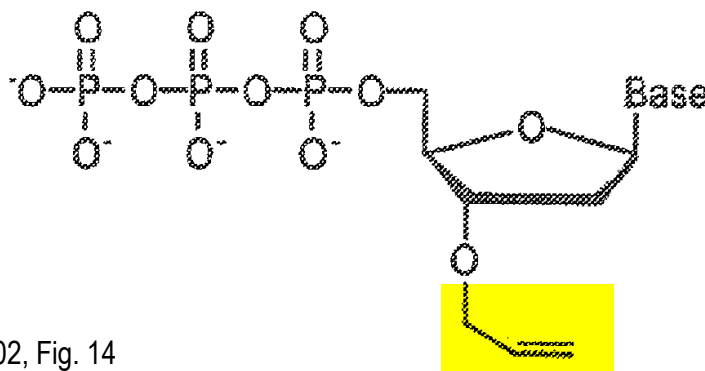
Ex. 2032, Illumina, Oral Hearing Tr., 48:10-15

Illumina's New References Relate to Alloc, Which Is a Carbonate Protecting Group, Not an Ether

Columbia's Patent



Columbia Capping Group: An Allyl *Ether*

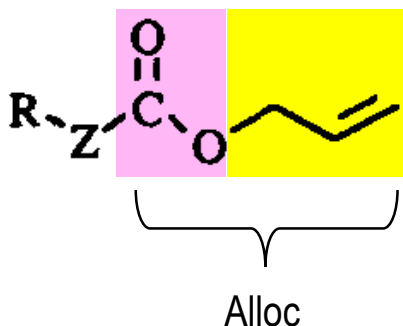


Ex. 1002, Fig. 14

Lemaire-Audoire, et al.



Illumina's Belated References: Allyl **Oxy Carbonyl** ("Alloc")



Ex. 1115, 248

Illumina Concedes That the Prior Art Taught That Ethers Are Hard to Cleave

a hydroxyl group upon deprotection. We reasoned that **ether bonds would be hard to cleave** under mild conditions compatible with DNA chemical stability, whereas

Ex. 2031, Canard, 4

Filed: September 2, 2014
On behalf of: Patent Owner Illumina Cambridge Ltd.
By:
Lead Counsel:
Robert A. Lawler
REINHART BROOKER VAN DUREN & C.
Tel.: 608-278-2217
Email: rlawler@reinhardlaw.com
Backup Counsel:
Bretton R. Babcock
William R. Zimmerman (pro hac vice)
Jonathan E. Buckard
Sheila N. Swanson
KIMBLE, MARTIN, OLSON & BEAR, LLP
Tel.: (949) 760-4004
Email: Bob.Illumina@kimbobee.com
Derek C. Worley (pro hac vice)

7. In Ex. 2154 at 81:8-13, Dr. Metzker testified that it was not known that **ether bonds were hard to cleave** under DNA compatible conditions. But the art did teach this. Ex. 2043, p. 4; Ex. 1002, 3:9-14.

Ex. 2033, Illumina, 3

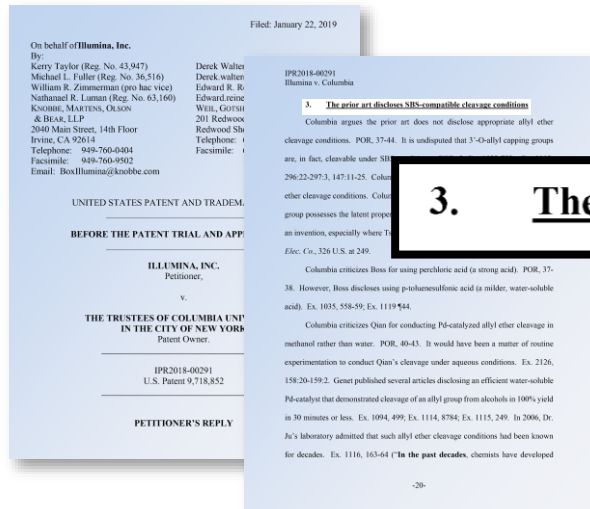
illumina

Canard's '94 paper, Exhibit 2043, expressly states: We reasoned that **ether bonds would be hard to cleave** under mild conditions compatible with DNA chemical stability. So even in '94 there was a hint that the conditions needed to cleave an ether weren't going to work in DNA synthesis.

Ex. 2032, Illumina, Oral Hearing Tr., 37:7-12

Illumina failed to provide evidence that alloc references teach allyl ether cleavage under SBS-compatible conditions

Illumina's Current Position That SBS-Compatible Allyl Ether Cleavage Conditions Were Known Prior to 2000 Contradicts Its Previous Position



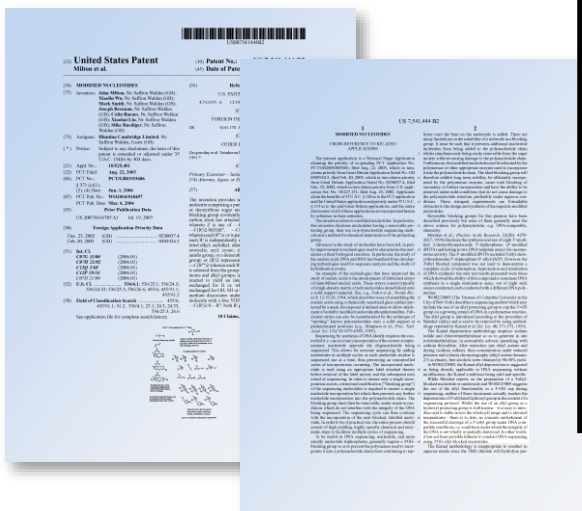
Illumina's Current Position

3. The prior art discloses SBS-compatible cleavage conditions

IPR2018-00291,
Reply, 20

Illumina's Previous Position

sequencing protocol. Whilst the use of an allyl group as a hydroxyl protecting group is well known—it is easy to introduce and is stable across the whole pH range and to elevated temperatures—there is to date, no concrete embodiment of the successful cleavage of a 3'-allyl group under DNA compatible conditions, i.e. conditions under which the integrity of the DNA is not wholly or partially destroyed. **In other words, it has not been possible hitherto to conduct DNA sequencing using 3'OH allyl-blocked nucleotides.**



Ex. 2015, '444 Illumina Patent, 2:57-65

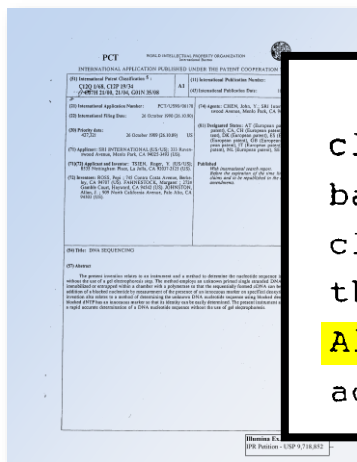
Illumina's Tsien-Based Arguments Fail

Relevant to:

IPR2018-00291, 318, 322, 385: Ground 1

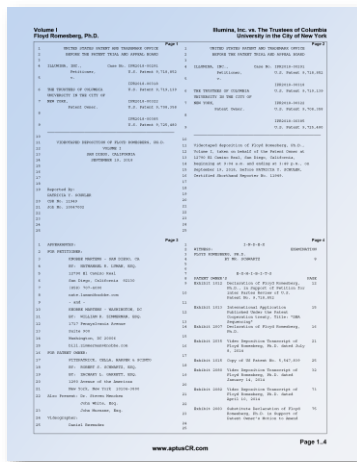
IPR2018-00797: Grounds 1-2

Tsien Does Not Describe the $-\text{CH}_2-\text{CH}=\text{CH}_2$ Capping Group



A wide variety of hydroxyl blocking groups are cleaved selectively using chemical procedures other than base hydrolysis. 2,4-Dinitrobenzenesulfonyl groups are cleaved rapidly by treatment with nucleophiles such as thiophenol and thiosulfate (Letsinger et al., 1964). Allyl ethers are cleaved by treatment with Hg(II) in acetone/water (Gigg and Warren, 1968).

Ex. 1013, Tsien, 24



Floyd Romesberg, Ph.D. Deposition

Q. And throughout Gigg and Warren there are disclosures of allyl ethers that are both unsubstituted and substituted, correct?

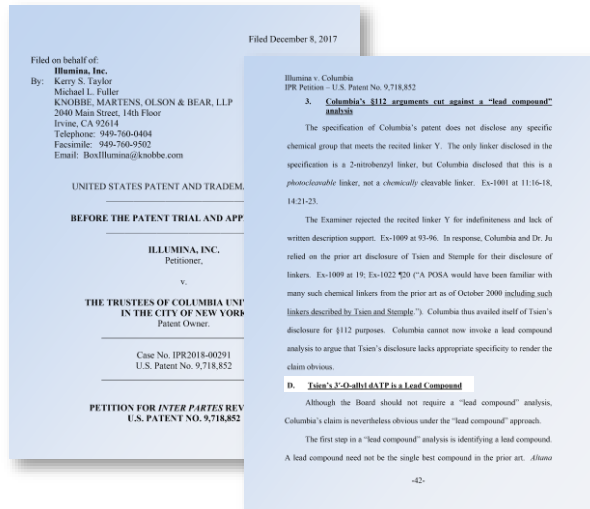
A. Apparently, yes.

Q. And throughout Gigg and Warren there are also disclosures of allyl ethers other than a three carbon allyl ether, correct?

A. That is correct. Obviously, from what you just pointed out. At least.

Ex. 2126, Romesberg Dep., 214:5-13

Illumina's Current Position That Tsien Describes the $-\text{CH}_2-\text{CH}=\text{CH}_2$ Capping Group Contradicts Its Previous Position



Illumina's Current Position

D. Tsien's 3'-O-allyl dATP is a Lead Compound

IPR2018-00291,
Petition, 42

Illumina's Previous Position

used at the remote position, Tsien does not describe using an allyl ether at the 3' position, as claimed.

Ex. 2065, 86

Tsien does not teach or suggest a nucleoside 5' triphosphate having an allyl removable blocking moiety protecting the 3' position linked to the 3' carbon via an ether linkage. Ward is

Ex. 2065, 88

Illumina's Motivation to Select $-\text{CH}_2-\text{CH}=\text{CH}_2$ Capping Group Is Based Solely on Tsien

Filed December 8, 2017

Filed on behalf of:

Illumina, Inc.

By: Kerry S. Taylor
Michael L. Fuller
KNOBBE, MARTENS, OLSON & BEAR, LLP
2040 Main Street, 14th Floor
Irvine, CA 92614
Telephone: 949-760-0404
Facsimile: 949-760-9502
Email: BoxIllumina@knobbe.com

UNITED STATES PATENT AND TRADEMARK OFFICE

BEFORE THE PATENT TRIAL AND APPEAL BOARD

ILLUMINA, INC.
Petitioner,

v.

**THE TRUSTEES OF COLUMBIA UNIVERSITY
IN THE CITY OF NEW YORK**
Patent Owner.

Case No. IPR2018-00291
U.S. Patent No. 9,718,852

PETITION FOR
U.S. PATENT

Illumina v. Columbia

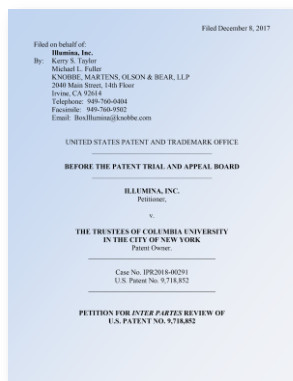
IPR Petition – U.S. Patent No. 9,718,852

The Board and Federal Circuit determined a POSA reading Tsien would clearly envisage a nucleotide having a fluorescent label attached via a cleavable linker and a 3'-OH capping group. Ex-1005 at 10; Ex-1008 at 31. Tsien discloses a nucleotide analogue having a fluorescent label attached via a cleavable linker to a base and a 3'-OH capping group. Tsien discloses that a 3'-O-allyl is an advantageous capping group because "deblocking occurs only when the specific deblocking reagent is present and premature deblocking during incorporation is minimized." Ex-1013 at 24:29-25:3. Tsien's disclosure of the desirability of the allyl protecting group would have motivated a POSA to select and use the 3'-O-allyl capping group on the nucleotide analogues disclosed by Tsien. Indeed, combining favorable disclosures within Tsien "does not require a leap of inventiveness." *Boston Sci. Scimed, Inc. v. Cordis Corp.*, 554 F.3d 982, 991 (Fed. Cir. 2009); Ex-1012 ¶¶94-95. Thus, based on Tsien's disclosure, a POSA would have been motivated to use Tsien's 3'-O-allyl group in Tsien's sequencing methods.

Cir. 2009); Ex-1012 ¶¶94-95. Thus, based on Tsien's disclosure, a POSA would have been motivated to use Tsien's 3'-O-allyl group in Tsien's sequencing methods.

IPR2018-00291, Petition, 34

The “Non-Base Hydrolysis” Advantage Alleged in the Petition Does Not Exist for Allyl Ethers



Petition

Tsien discloses that the allyl group is advantageous because it is “cleaved selectively using chemical procedures other than base hydrolysis ... [which has] some advantages over groups cleavable by base hydrolysis—deblocking occurs

IPR2018-00291, Petition, 21-22 (citing Gigg and Warren)

Institution Decision

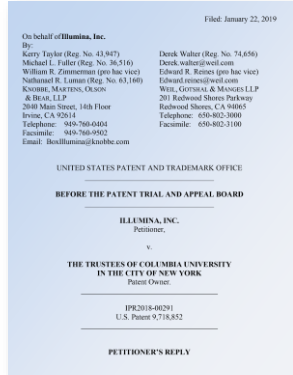
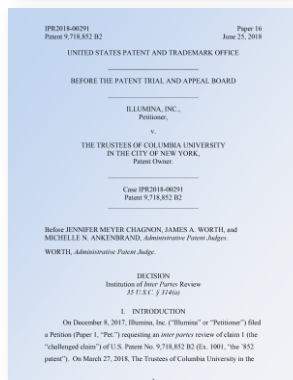
(using “well-established synthetic procedures”). Based on the evidence at this stage of the proceeding, we are persuaded that Petitioner has made a sufficient showing that a person of ordinary skill would have combined the teachings of Tsien to use the allyl group that Tsien discloses to be an advantageous blocking group, to block the 3'-oxygen, as elsewhere disclosed in Tsien's method. *See* Ex. 1013, 12:27–29, 24:5–25:3. We invite further

IPR2018-00291, Institution Dec., 13-14

Reply

cleavage. Tsien cites to the pioneering 1968 Gigg article, which discloses that allyl ether is cleavable by base hydrolysis. Ex. 1013, 24:29-30; Ex. 1046, 1905-06. Tsien

IPR2018-00291, Reply, 19



Illumina's Belated Argument That Tsien Additionally Teaches Non-Base Hydrolysis Is Unsupported

PCT WORLD INTELLECTUAL PROPERTY ORGANIZATION International Bureau	
INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)	
(51) International Patent Classification ⁵ : C12Q 1/68, C12P 19/34 C12N 21/00, 21/04, G01N 35/08	A1 (11) International Publication Number: WO 91/06678 (43) International Publication Date: 16 May 1991 (16.05.91)
(21) International Application Number: PCT/US90/06178 (22) International Filing Date: 26 October 1990 (26.10.90)	(74) Agents: CHEN, John, Y., SRI International, 333 Ravenswood Avenue, Menlo Park, CA 94025-3493 (US) et al. (81) Designated States: AT (European patent), BE (European
(30) Priority data: 427,321 26 October 1989	A w cleaved select base hydrolysis cleaved rapidly thiophenol and Allyl ethers a acetone/water Tetrahydrothio conditions us Cruse et al.,
(71) Applicant: SRI INTERNATIONAL INC. 3333 Wood Avenue, Menlo Park, CA 94025	
(71)(72) Applicant and inventor: TSIEN, R. 8555 Nottingham Place, La Jolla, CA 92037 (71)(72) Inventors: ROSS, Pepi, 745 Contra Rey, CA 94707 (US); FARNSTOCK, Gamble Court, Hayward, CA 94542; Altan, J., 909 North California Avenue, 94303 (US).	
(54) Title: DNA SEQUENCING	
(57) Abstract The present invention relates to an improved method for determining the sequence of a DNA without the use of a gel electrophoresis step, immobilized or entrapped within a chamber addition of a blocked nucleotide by measuring fluorescence. The invention also relates to a method of determining the sequence of a DNA without the use of a blocked dNTP as an in situ marker so as to provide a rapid accurate determination of a DNA sequence.	

Illumina Ex. 1013
IPR Petition - USP 9,718,852

WO 91/06678

PCT/US90/06178

-24-

A wide variety of hydroxyl blocking groups are cleaved selectively using chemical procedures other than base hydrolysis. 2,4-Dinitrobenzenesulfonyl groups are cleaved rapidly by treatment with nucleophiles such as thiophenol and thiosulfate (Letsinger et al., 1964). Allyl ethers are cleaved by treatment with Hg(II) in acetone/water (Gigg and Warren, 1968). Tetrahydrothiofuranyl ethers are removed under neutral conditions using Ag(I) or Hg(II) (Cohen and Steele, 1966; Cruse et al., 1978). These protecting groups, which are

Allyl ethers are cleaved by treatment with Hg(II) in
30 acetone/water (Gigg and Warren, 1968).
Tetrahydrothiofuranly ethers are removed under neutral
conditions using Ag(I) or Hg(II) (Cohen and Steele, 1966;
Cruse et al., 1978). These protecting groups, which are
35 stable to the conditions used in the synthesis of dNTP
analogues and in the sequence incorporation steps, have
some advantages over groups cleavable by base hydrolysis.

Ex. 1013,
Tsien, 24

Illumina's Dower-Based Arguments Fail

Relevant to:

IPR2018-00291, 318, 322, 385: Ground 2

IPR2018-00797: Ground 3

Dower's Method Is SBS

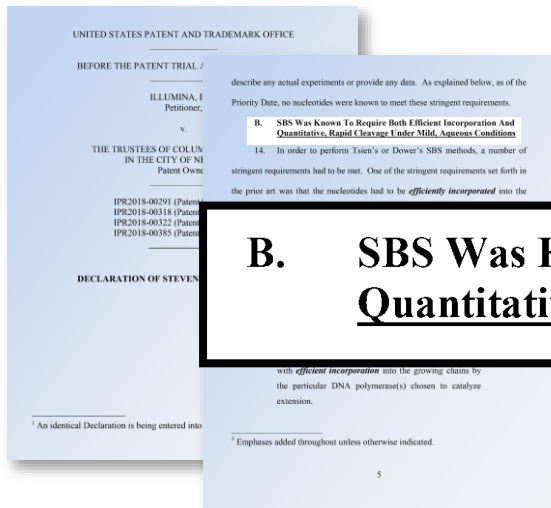
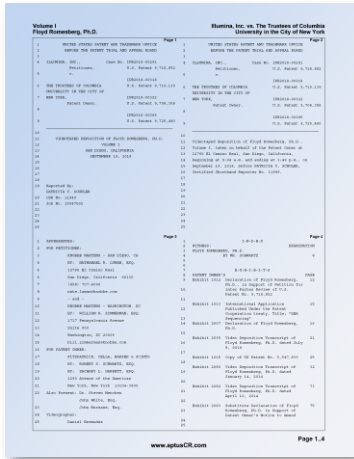
Floyd Romesberg, Ph.D.

Deposition

Q. And the Dower patent relates to SBS, correct?

A. That is correct.

Ex. 2126, Romesberg Dep., 25:15-17



B. SBS Was Known To Require Both Efficient Incorporation And Quantitative, Rapid Cleavage Under Mild, Aqueous Conditions

Ex. 2116, Menchen Decl., 5

Dower's SBS Method Requires Efficient Incorporation

United States Patent [19]
Dower et al.

Patent Number: 5,547,839
Date of Patent: Aug. 20, 1996

[54] SEQUENCING OF SURFACE IMMOBILIZED
POLYMERS UTILIZING
MICROFLUORESCENCE DETECTION

[75] Inventors: William J. Dower, Menlo Park;
Stephen P. A. Fodor, Palo Alto, both
of Calif.

[73] Assignee: Affymax Technologies N.V., Curacao,
Netherlands Antilles

[21] Appl. No.: 626,730

[22] Filed: Dec. 6, 1990

Related U.S. Application Data

[63] Continuation-in-part of Ser. No. 492,462, Mar. 7, 1990, Pat.
No. 5,142,854, which is a continuation-in-part of Ser. No.
362,901, Jan. 7, 1989, abandoned.

[51] Int. Cl.⁶ C12Q 1/68

[52] U.S. Cl. 435/6; 536/24.33; 536/24.3

[58] Field of Search 435/6, 91, 810,
435/973, 975, 912; 436/227, 530, 531,
56, 94, 800, 808; 536/27, 24.33; 935/77

References Cited

U.S. PATENT DOCUMENTS

4,542,102 9/1985 Danagupta et al. 435/6
4,582,789 4/1986 Sheldon et al. 435/6
4,656,127 4/1987 Musny 435/6
4,689,405 9/1987 Frank et al. 536/27
4,713,226 12/1987 Danagupta et al. 435/6
4,855,225 8/1989 Fung et al. 435/6
4,889,818 12/1989 Gelfand et al. 435/194
4,962,037 10/1990 Jett et al. 435/6
4,965,188 10/1990 Mullis et al. 435/6
5,002,867 3/1991 Macveizer 435/6
5,028,840 6/1991 Danagupta et al. 536/27
5,075,216 12/1991 Imus et al. 435/6
5,126,239 6/1992 Livak et al. 435/6
5,143,854 9/1992 Perng et al. 436/118

FOREIGN PATENT DOCUMENTS

392546 10/1990 European Pat. Off.
223654 1/1991 United Kingdom

OTHER PUBLICATIONS

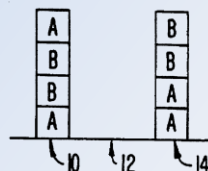
Ikehara et al., "The Synthesis of Polynucleotides," *Advances in Carbohydrate Chemistry and Biochemistry* (1979) 36:135-213.
Taugita et al., "Sensitization of Edman Amino Acid Derivatives Using the Fluorescent Reagent, 4-Aminofluorescein," *J. Biochem.* (1989) 106:60-65.
Ross et al., "Interspersed Crosslinks due to 4,5,8-Trimethyl-pyranol and Near Ultraviolet Light in Specific Sequences of Animal DNA," *J. Mol. Biol.* (1988) 201:339-351.
Bairdaga, 27 Sep. 1991, *Science* 253:1489.
Fodor et al., 15 Feb. 1991, *Science* 251:767-773.
Dower and Fodor, 1991, *Ann. Rep. Med. Chem.* 26:271-280.
Little, 16 Aug. 1990, *Nature* 346:611-612.
Craig et al., 1990, *Nucl. Acids, Res.* 18:2653-2660.
Chatterjee et al., 1990, *J. Am. Chem. Soc.* 112:6397-6399.
Pfeiffer et al., 10 Nov. 1989, *Science* 246:810-812.
Evans et al., Jul. 1989, *Proc. Natl. Acad. Sci. USA* 86:5030-4.
Saiki and Gelfand, 1989, *Amplifications* 1:4-6.
(List continued on next page.)

Primary Examiner—W. Gary Jones
Assistant Examiner—Scott Hostetman
Attorney, Agent, or Firm—Townsend & Townsend & Crew
LLP

ABSTRACT

Means for simultaneous parallel sequence analysis of a large number of biological polymer macromolecules. Apparatus and methods may use fluorescent labels in repetitive chemistry to determine terminal monomers on solid phase immobilized polymers. Reagents which specifically recognize terminal monomers are used to label polymers at defined positions on a solid substrate.

7 Claims, 10 Drawing Sheets



Illumina Ex. 1015
IPR Petition - USP 9,718,852

25

nents appropriate to the particular DNA polymerase selected, are placed in the incubation chamber in direct contact with the surface.

Step 2: Fluorescent chain terminators (analog of dATP, dCTP, dGTP, and TP, each labeled with fluorophore preferably emitting at a distinguishable wavelength) are added to the reaction at a sufficient concentration and under suitable reaction conditions (time, temperature, pH, ionic species, etc., See Sambrook et al. (1989) *Molecular Cloning*, vols. 1-3, and Prober et al.) to cause essentially all of the chains on the surface to be extended by one base and thereby terminated. Detection of the specific label thereby incorporated into each chain identifies the last base added at each positional address in the matrix.

Step 3: The chain termination should be reversible by some means, such as treatment with light, heat, pH, certain other chemical or biological (enzymatic) reagents, or some combination of these. Typically the chain termination results from a blocking moiety which is labile to mild treatment. By one of these means, the blocked 3'OH of the terminating base must be made available for chain extension in the next round of polymerization.

Step 4: There are several suitable labeled, terminator structures as follows:

(a) The fluorophore itself functions as the chain terminator by placement on the 3' hydroxyl through a linkage that

26

followed by scanning detection. After all four fluorophores are added, reversal of the termination is performed, allowing for the addition of the next base analog. Then, each scanning step determines whether the immediately preceding labeled nucleotide had been incorporated at each distinct position.

The structures of the fluorescently labeled and reversible terminator base analogs are selected to be compatible with efficient incorporation into the growing chains by the particular DNA polymerase(s) chosen to catalyze extension. For example, where two different chain terminators are used, they may be utilized by two different polymerases that are both present during the chain extension step.

Step 5: An optional step is the permanent capping of chain extension failures with high concentrations of dideoxynucleoside triphosphates. This step serves to reduce the background of fluorescence caused by addition of an incorrect base because of inefficient chain extension (termination) at an earlier step.

Step 6: After scanning to determine fluorescence, the fluorophore is removed or deactivated. Deactivation of the fluorophore can be achieved by a photodestruction event. The chain elongation block is reversed (usually by removing a blocking group to expose the 3'OH) by suitable methods that depend on the particular base analogs chosen; and the substrate is washed in preparation for the next round of polymerization.

The structures of the fluorescently labeled and reversible terminator base analogs are selected to be compatible with efficient incorporation into the growing chains by the particular DNA polymerase(s) chosen to catalyze extension.

Ex. 1015,
Dower, 26:6-9

nucleotide triphosphate analogs as described by Prober et al. (1987) *Science* 238:336-341. A second, unlabeled and reversible, set of terminators is also required. Examples of these compounds are deoxynucleotide triphosphates with small blocking groups such as acetyl, BOC, NBOC and NBOC on the 3'OH. These groups are easily and efficiently removed under conditions of high or low pH, exposure to light or heat, etc. After each round of base addition and detection, the fluorophores are deactivated by exposure to light under suitable conditions (these chains have their 3' labeling moiety destroyed and remain terminated, taking part in no further reactions). The unlabeled, reversible terminators are unblocked at the 3'OH by the appropriate treatment to allow chain extension in subsequent rounds of elongation. The proportion of chains labeled in each round can be controlled by the concentration ratio of fluorescent to non-fluorescent terminators, and the reaction can be driven to completion with high concentrations of the unlabeled terminators.

(d) A single dye strategy is used where all the base analog terminators carry the same fluorophore and each is added one at a time: A, C, G, T. The addition of each base is

The DNA will usually be attached to the substrate by the 3' or 5' terminus depending on the scheme of labeling and cleavage. Because there are well-known 5'-labeling methods, see, e.g., Gait (1984) *Oligonucleotide Synthesis: A Practical Approach*, IRL Press, Oxford, this discussion will assume the 3' end is attached to the substrate with the 5' end free.

Step 1: All the 5'-end bases are labeled with 5'-specific chemistry, e.g., 5' amino linkage to FITC, Nelson et al. (1989) *Nucl. Acid Res.* 17:7179-7186, which is hereby incorporated herein by reference.

Step 2: Scan the matrix to obtain the background level.

Step 3: Optional: Cap all of the labeling failures, e.g., polymers whose ends were not labeled.

Step 4: The terminal A's are removed with end-base, A-specific reagents (such a reagent may be chemical or biological). One example is a 5'-fluorescein-dAMP-specific exonuclease made as a catalytic antibody (see the description above for a scheme of producing this reagent).

Step 5: Scan the matrix to detect those chains that had terminated in A (these will be reduced in fluorescence compared to the fluorescently labeled background).

Illumina's Belated Position That Dower Does Not Require Efficient Incorporation Is Belied by Its Petition

Illumina's Current Position

Dower does not recite the incorporation efficiency criteria of Tsien,

IPR2018-00291, Reply, 25

Illumina's Previous Position

Dower also discloses that the label and 3'-OH blocking group "are selected to be compatible with efficient incorporation into the growing chains by the particular DNA polymerase(s) chosen to catalyze extension." *Id.* at 26:6-9. Dower's

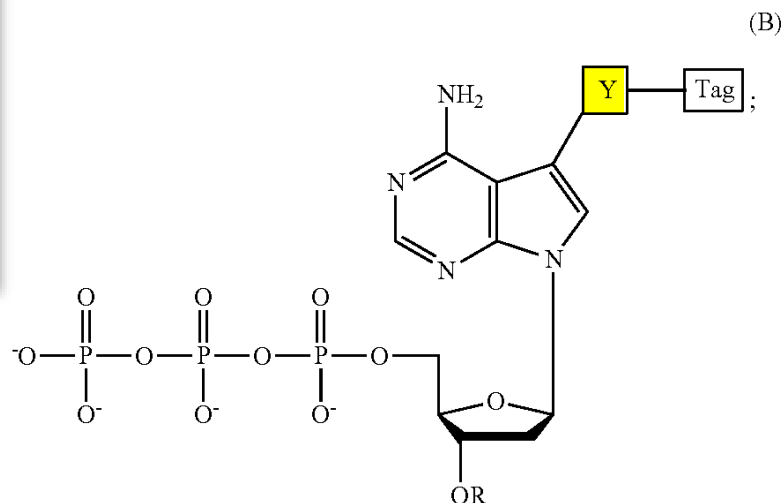
IPR2018-00291, Petition, 65

Dower/Prober Does Not Describe the Claimed Cleavable Linker “Y”

Claim:

Ex. 1075, claim 1

wherein **Y** represents a chemically cleavable, chemical linker which (a) does not interfere with recognition of the analogue as a substrate by a DNA polymerase and (b) is stable during a DNA polymerase reaction;



Dower:

No Y

Ex. 2116, Menchen Decl., ¶111

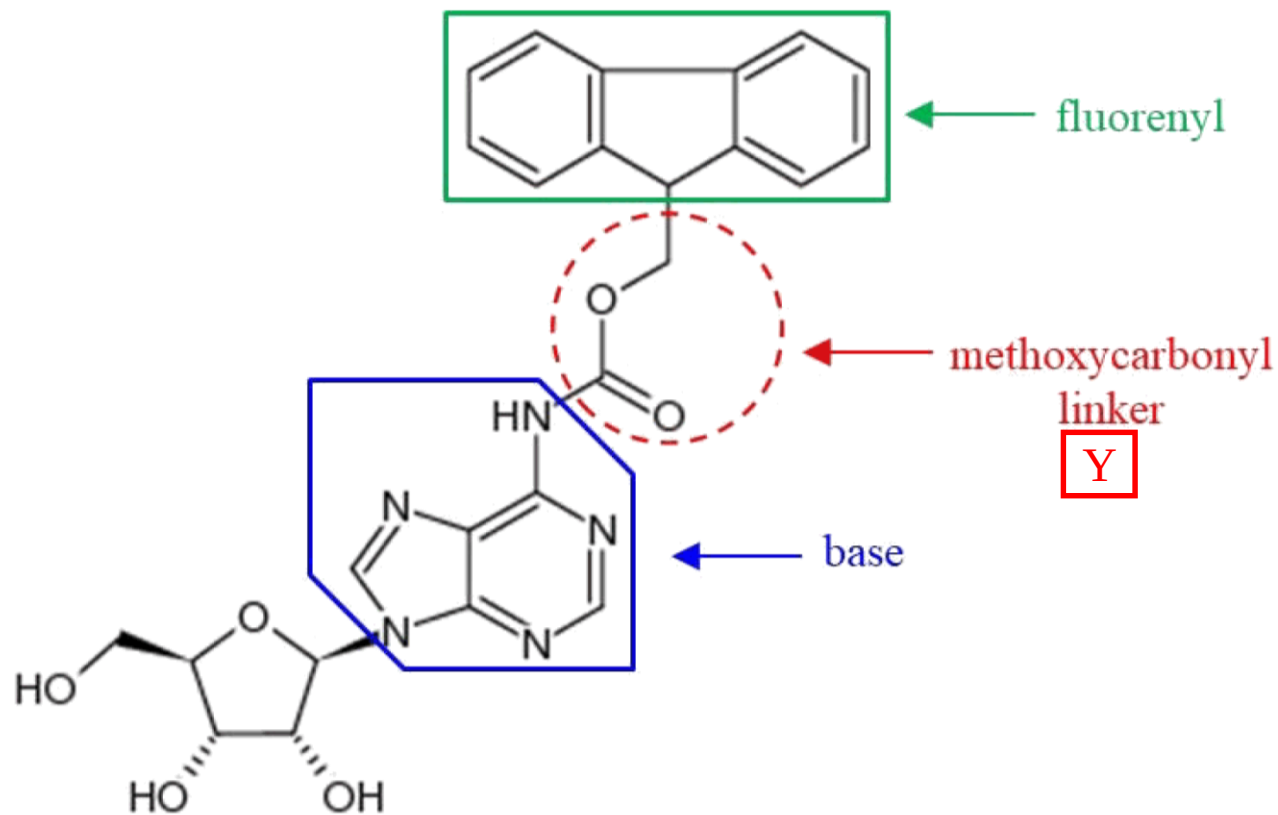
Prober:

No cleavable Y

Ex. 2113, Romesberg Dep., 124:11-15

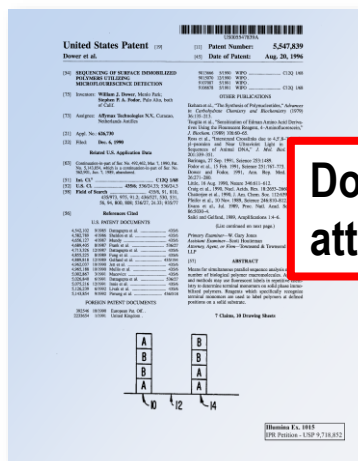
Ex. 2116, Menchen Decl., ¶111

Illumina Belatedly Alleges Dower's FMOC Has Built-In Cleavable Linker

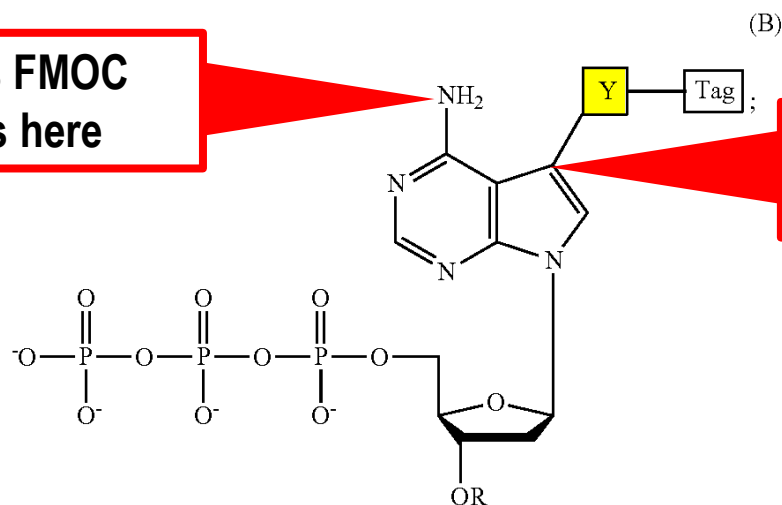


IPR2018-00291, Reply, 26

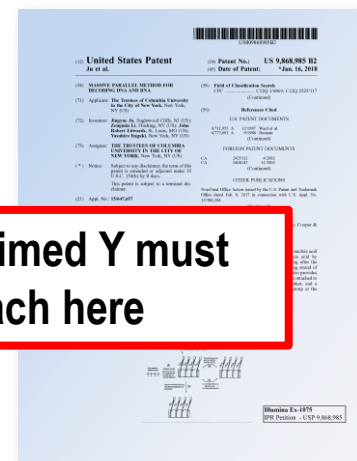
Dower's FMOC Attaches to Different Site Than Claimed Y



Dower's FMOC attaches here



**Claimed Y must
attach here**

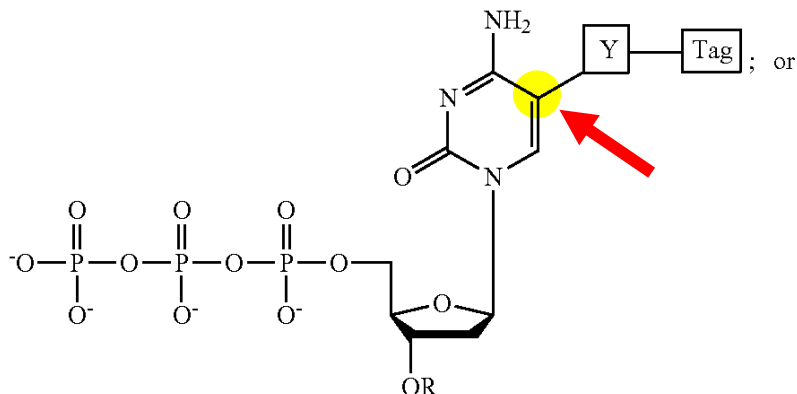


Ex. 1075, claim 1

Dower's FMOC Attaches to Different Site Than Claimed Y

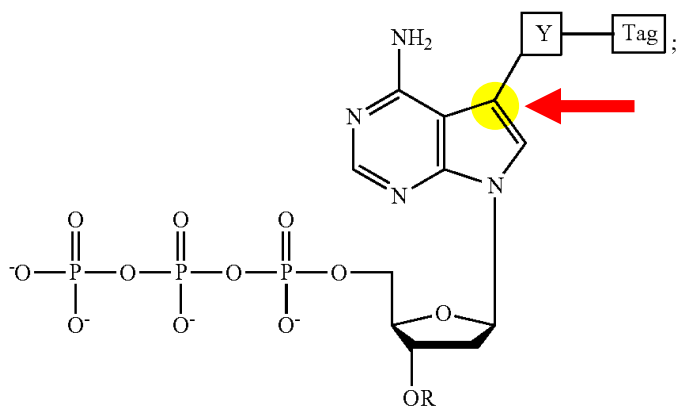
Claimed linker "Y" on 5-pyrimidine carbon

(C)



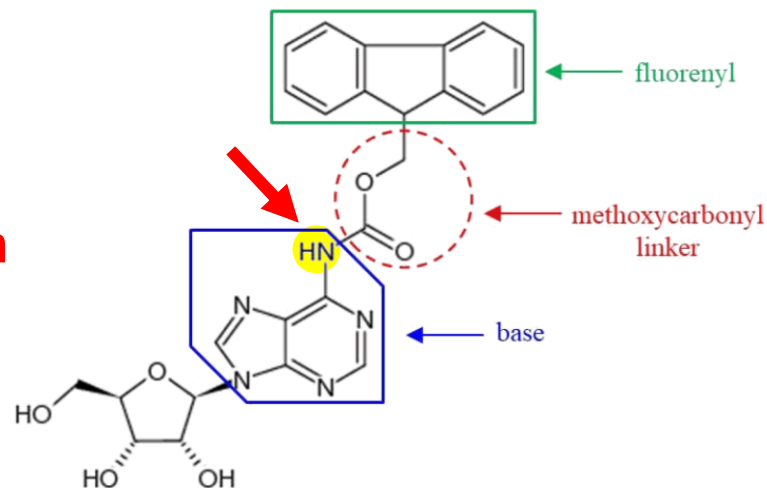
Claimed linker "Y" on 7-deazapurine carbon

(B)



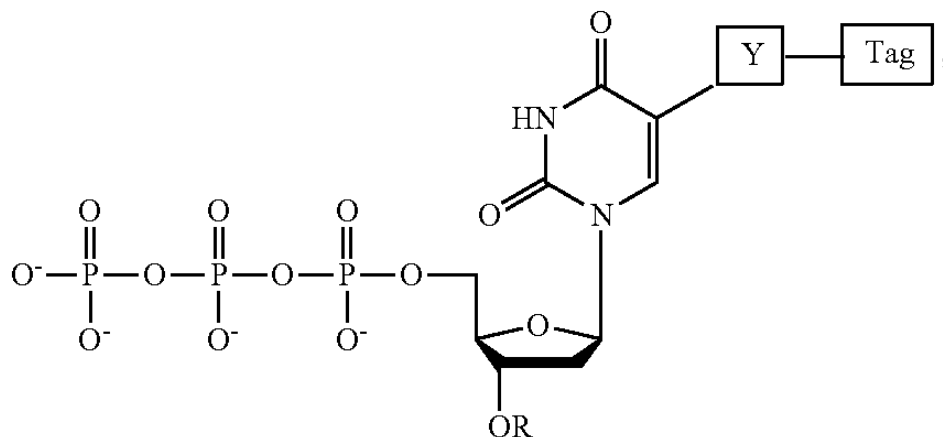
Ex. 1075, '985 Patent, claim 1

Dower FMOC on exocyclic amine

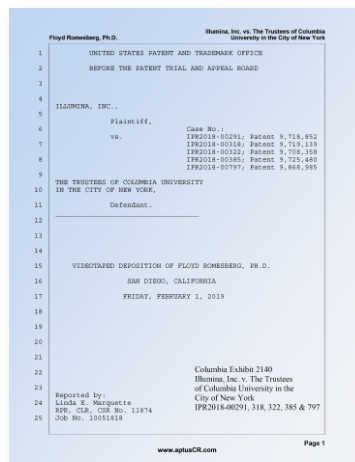


IPR2018-00291, Reply, 26

Dower's FMOC Cannot Attach to Thymine Because It Lacks an Exocyclic Amine



Ex. 1003, '139 Patent, claim 1



Floyd Romesberg, Ph.D.

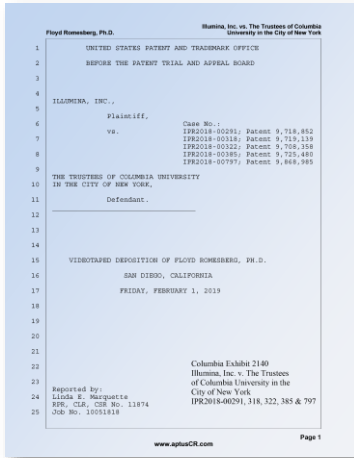
Deposition

Q. But Dower doesn't teach attaching FMOC to the three bases that he teaches attaching it to through any kind of additional linker, correct?

A. Dower gives an example where he uses FMOC to attach directly to the three of the four that you could directly attach to. **T is not.**

Ex. 2140, Romesberg Dep., 197:16-21

ILLUMINA Presents No Evidence That Chemistry Existed to Attach FMOC to Claimed Position



Floyd Romesberg, Ph.D. Deposition

Q. You haven't cited any references in any declarations in this IPR -- these IPRs with conditions where you could attach FMOC to the 5 position, correct?

A. No. You asked me if I thought you could. And I'm simply speculating.

Q. Okay. But you haven't cited any.

A. No, nor do I know of any.

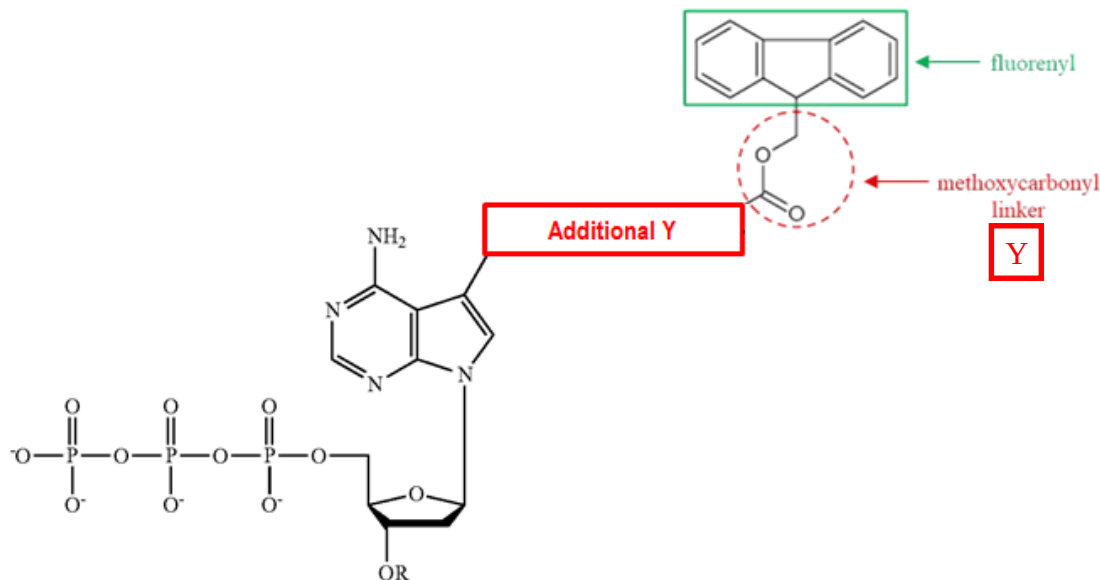
Q. Okay. And the same for the diaza position, attaching it to the diaza position of a 7 —

...

A. I did not.

Ex. 2140, Romesberg Dep., 201:8-23

Illumina's Most Recent Position Requires Adding Another Linker

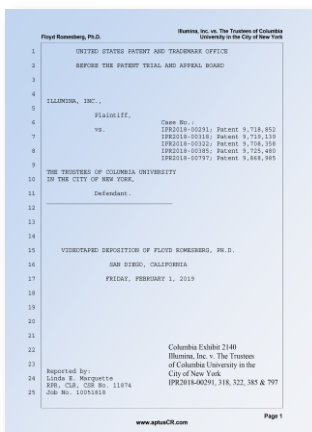


Floyd Romesberg, Ph.D.

Deposition

Q. But Dower doesn't teach attaching Fmoc to the three bases that he teaches attaching it to through any kind of additional linker, correct?

A. ...He didn't attach the Fmoc directly, but it would be an exocyclic amine that you would exactly be able to do that if you chose. **Or you would be able to choose an additional linker between them. And that's what Seitz did.**



Ex. 2140, Romesberg Dep., 197:16-198:8

Columbia Did Not Admit That Kamal Teaches SBS-Compatible Cleavage Conditions

Relevant to:

IPR2018-00291, 318, 322, 385: Grounds 1-2

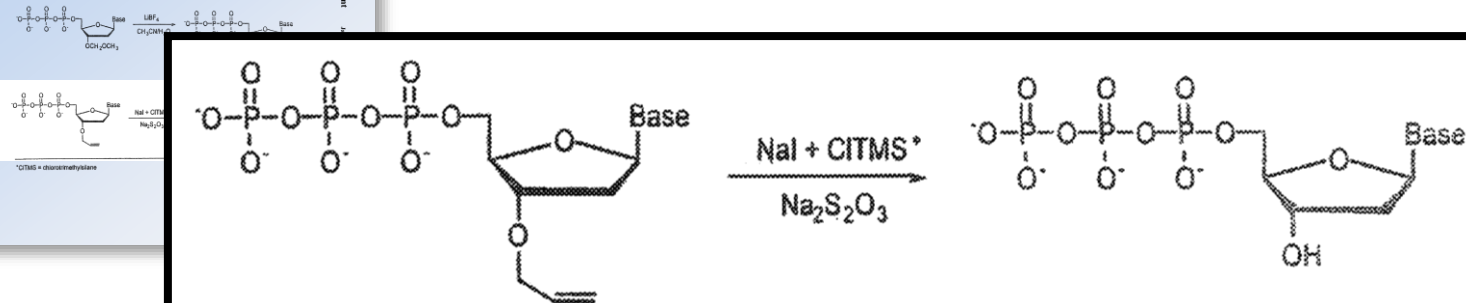
IPR2018-00797: Grounds 1-3

Columbia's Statements Concerning Allyl Protecting Groups Are Not Admissions of Prior Art Teachings

These groups can be removed chemically with high yield **as shown in FIG. 14** (Ireland, et al, 1986; Kamal et al. 1999).

Ex. 1075, '985 Patent, 26:22-23*

FIGURE 14



Ex. 1075, '985 Patent, Fig. 14

FIGURE 14 is Not in Kamal

* References incorporated in their entirety, Ex. 1075, 1:35-38

Columbia Does Not Cite Kamal for Mild and Specific Conditions

US009868985B2

(12) **United States Patent**
Ju et al.

(10) **Patent No.:** US 9,868,985 B2
(45) **Date of Patent:** *Jan. 16, 2018

(54) **MASSIVE PARALLEL METHOD FOR DECODING DNA AND RNA**

(71) **Applicant:** The Trustees of Columbia University in the City of New York, New York, NY (US)

(72) **Inventors:** Jingyue Ju, Englewood Cliffs, NJ (US); Zengmin Li, Flushing, NY (US); John Robert Edwards, St. Louis, MO (US); Yasuhiro Itagaki, New York, NY (US)

(73) **Assignee:** THE TRUSTEES OF COLUMBIA UNIVERSITY IN THE CITY OF NEW YORK, New York, NY (US)

(*) **Notice:** Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days. This patent is subject to a terminal disclaimer.

(21) **Appl. No.:** 15/647,657

(22) **Filed:** Jul. 12, 2017

(65) **Prior Publication Data**
US 2017/0313737 A1 Nov. 2, 2017

Related U.S. Application Data

(60) Continuation of application No. 15/167,917, filed on May 27, 2016, now Pat. No. 9,725,480, which is a (Continued)

(51) **Int. Cl.**
C12Q 1/68 (2006.01)
C07H 19/14 (2006.01)
(Continued)

(52) **U.S. Cl.**
CPC **C12Q 1/6869** (2013.01); **C07H 19/14** (2013.01); **C07H 21/00** (2013.01); **C12Q 1/68** (2013.01);
(Continued)

(58) **Field of Classification Search**
CPC **C12Q 1/6869**; **C12Q 2525/117**
(Continued)

(56) **References Cited**
U.S. PATENT DOCUMENTS
4,711,955 A 12/1987 Ward et al.
4,772,691 A 9/1988 Herman
(Continued)
FOREIGN PATENT DOCUMENTS
CA 2425112 4/2002
CA 2408143 11/2002
(Continued)
OTHER PUBLICATIONS
Non-Final Office Action issued by the U.S. Patent and Trademark Office dated Feb. 8, 2017 in connection with U.S. Appl. No. 15/380,284.
(Continued)

(74) **Attorney, Agent, or Firm:** Dunham LLP

(57) **ABSTRACT**
This invention provides methods for detecting the identity of a nucleotide analog is incorporated into a polymerase reaction. The invention also provides nucleotide analogs which comprise unique labels attached to the nucleotide analog through a cleavable linker, and a cleavable chemical group to cap the —OH group at the 3'-position of the deoxyribose.

2 Claims, 28 Drawing Sheets

The chemical cleavage of the MOM and allyl groups is fairly mild and specific, so as not to degrade the DNA template moiety. For example, the cleavage of the allyl group takes 3

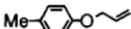
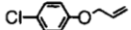
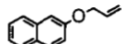
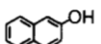
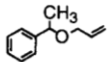
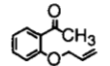
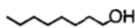
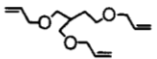
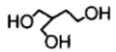
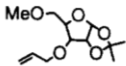
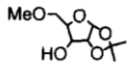
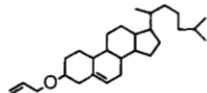
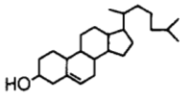
Ex. 1075, '985 Patent, 26:24-26

Illumina Ex-1075
IPR Petition - USP 9,868,985

Columbia Admitted Only That Kamal Taught 93% Cleavage of Non-Nucleotide Protected Alcohols Under Kamal's Conditions

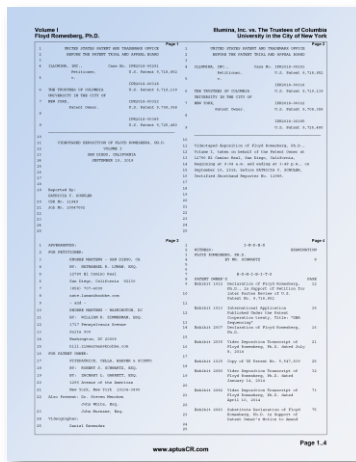
moiety. For example, the cleavage of the allyl group takes 3 minutes with more than 93% yield (Kamal et al. 1999),

Ex. 1075, '985 Patent, 26:26-27

Entry	Substrate	Time(min)	Product	Yield(%)
1		2		98
2		2		98
3		2		98
4		3		96
5		5		90
6		3		95
7		3		95 ¹⁶
8		2		93
9		3		93

Ex. 1037, Kamal, 372

Kamal's 93% Cleavage Is Not Useful For SBS



Floyd Romesberg, Ph.D. Deposition

A. Let me answer the question that I think you're getting at is that I think that we would need -- you would need for Tsien's sequencing by synthesis method, I think the answer is applicable to that, that you would need something greater than 95 percent for cleavage of the 3' hydrox group, if that's the question that you're asking.

Ex. 2126, Romesberg Dep., 70:12-18

Illumina Improperly Relies on Hindsight

Otsuka Pharmaceutical Co., Ltd. v. Sandoz, Inc., 678 F.3d 1280 (2012)
 102 U.S.P.Q.2d 1729

KeyCite Yellow Flag - Negative Treatment
 Distinguished by EX PARTE ROBERT EUGENE AND BING LI,
 Patent Tr. & App. Bd., May 11, 2018

678 F.3d 1280

United States Court of Appeals,
 Federal Circuit.

OTSUKA PHARMACEUTICAL
 CO., LTD., Plaintiff-Appellee,
 v.

SANDOZ, INC., Sun Pharmaceutical
 Industries, Ltd., Synthron BV, Synthron
 Holdings BV, Synthron Laboratories, Inc., and
 Synthron Pharmaceuticals, Inc., Defendants,
 and

Apotex Inc. and Apotex Corp.,
 Defendants-Appellants,

and
 Teva Pharmaceuticals USA, Inc.,
 Barr Laboratories, Inc., and Barr
 Pharmaceuticals, Inc., Defendants-Appellants.

Nos. 2011-1126, 2011-1127.

May 7, 2012.

Rehearing En Banc Denied Aug. 7, 2012.

Synopsis

Background: Patentee filed action against generic drug
 manufacturers alleging infringement of patent on active
 ingredient in drug approved by Food and Drug
 Administration (FDA) for treatment of schizophrenia,
 bipolar disorder, irritability associated with autistic
 disorder in pediatric patients, and as add-on treatment for
 depression. Manufacturers counterclaimed alleging that
 claims were invalid for obviousness and obviousness-type
 double patenting. The United States District Court for
 the District of New Jersey, Mary L. Cooper, J., 2010 WL
 4596324, sustained validity of asserted claims of patent
 after bench trial. Defendants appealed.

Holdings: The Court of Appeals, Lourie, Circuit Judge,
 held that:

carbostyrils could not be considered prior art;

patent was not obvious based on prior art patent that
 disclosed unsubstituted butoxy compound and “laundry
 list” of potential central nervous system controlling effects
 from carbostyryl;

patent was not obvious based on prior art declaration
 that disclosed three sets of test data comparing carbostyryl
 derivatives on mice;

manufacturers did not prove by clear and convincing
 evidence that patent was obvious based on prior art
 foreign patent application;

patent was not obvious based on patentee’s own prior art
 development compound; and

person of ordinary skill would not have been motivated to
 pursue 2,3-dichloro substitution on phenyl ring as would
 have been required to convert unsubstituted butoxy to
 carbostyryl.

The inventor's own path itself never leads to a conclusion
 of obviousness; that is hindsight. What matters is the

Of counsel on the brief were Robert E. Macentron and
 John D. Murnane, Fitzpatrick, Cella, Harper & Scinto, of
 New York, New York.

Steven E. Feldman, Husch Blackwell LLP, of Chicago,
 IL, argued for Defendants-Appellants Apotex Inc., et al.
 With him on the brief were Daniel R. Cherry and Sherry
 L. Rollo.

Elizabeth J. Holland, Kenyon & Kenyon LLP,
 of New York, NY, for defendants-appellants Teva
 Pharmaceuticals USA, Inc., et al. With her on the brief
 was Maria Luisa Palmese.

Before LOURIE, MOORE, and REYNA, Circuit Judges.

Opinion

LOURIE, Circuit Judge.

Otsuka Pharm. Co. v. Sandoz, Inc.,
 678 F.3d 1280, 1296 (Fed. Cir. 2012)

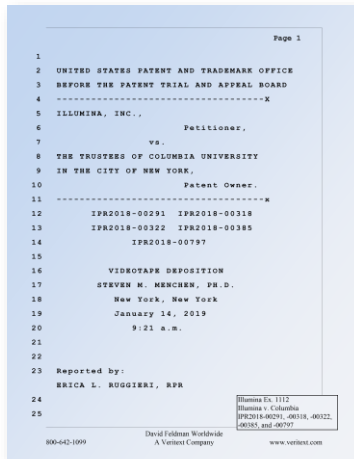
Dr. Menchen's Sanger Sequencing Patents Are Irrelevant

Relevant to:

IPR2018-00291, 318, 322, 385: Grounds 1-2

IPR2018-00797: Grounds 1-3

Dr. Menchen Did Not Include “Allyloxy” in His Sanger Patents Because of Metzker 1994



Stephen Menchen, Ph.D.
Deposition

Q. What would a person of ordinary skill in the art in 2000 gather from Metzker 1994 in terms of the incorporation efficiency of allyl groups, do you know?

A. They would look at Metzker and see that when the allyl group's on dA that it's inefficiently incorporated with Vent.

Q. How come you still identified allyl groups as a suitable modifier for 3' position in your patents?

A. Because that's what Metzker calls it. But he definitely refers to the structure of propenyl in that paper. **He calls it allyl in the paper.**

Ex. 1112, Menchen Dep., 188:20-189:13

1 UNITED STATES PATENT AND TRADEMARK OFFICE

2 REOPEN THE PATENT TRIAL AND APPEAL BOARD

3

4 ILLUMINA, INC.,

5 Plaintiff,

6 vs. Case No.:

7 IP20018-00291; Patent 5,718,852

8 IP20018-00318; Patent 5,735,119

9 IP20018-00322; Patent 5,708,168

10 IP20018-00360; Patent 5,725,400

11 IP20018-00797; Patent 5,668,995

12 THE TRUSTEES OF COLUMBIA UNIVERSITY

13 IN THE CITY OF NEW YORK,

14 Defendant.

15

16 VIGORATED DEPOSITION OF FLOYD ROMERBERG, FR.D.

17 SAN DIEGO, CALIFORNIA

18 FRIDAY, FEBRUARY 1, 2019

19

20

21

22 Columbia Exhibit 7140

23 Illumina, Inc. v. The Trustees

24 of Columbia University in the

25 City of New York

26 IP20018-00291, 318, 322, 365 & 797

27

28

29

30

31

32

Reported by:
Lidia E. Margueta
P.R. Clk. CMB No. 11874
O&B No. 10051818

www.justCt.com

Page 1

Deposition

Q. Okay. But for SBS the goal would be to have very high termination, **almost complete termination** of the growing DNA strands to prevent out-of-phase sequencing, correct?

Ex. 2140, Romesberg Dep., 149:1-17

SBS Methods Require Multiple Cycles

Relevant to:

IPR2018-00291, 318, 322, 385: Grounds 1-2

IPR2018-00797: Grounds 1-3

The Board Invited Briefing On Whether Columbia's Method Patent Requires Multiple Cycles of SBS: It Does

Trials@uspto.gov
Tel: 571-272-7822

Paper 20
Entered: September 18, 2018

UNITED STATES PATENT AND TRADEMARK OFFICE

BEFORE THE PATENT TRIAL AND APPEAL BOARD

ILLUMINA, INC.,
Petitioner,

v.

THE TRUSTEES OF COLUMBIA UNIVERSITY
IN THE CITY OF NEW YORK,
Patent Owner.

Case IP
Patent 9

Before JAMES A. WORTH, MICHAEL
BRIAN D. RANGE, *Administrative Patent Judges*

RANGE, *Administrative Patent Judges*

DEPARTMENT OF COMMERCE
Institution of 7
35 U.S.C. § 314

Case IPR2018-00797
Patent 9,868,985 B2

including whether it was appreciated in the art that sufficiently mild conditions were known for cleaving allyl groups when working with nucleotide analogues. We also invite briefing on (1) whether or not the parties take the position that either challenged claim necessarily requires sequencing, for example, of whole genomes as Dr. Menchen discusses and (2) whether or not the claims otherwise require that recited steps be repeated. Ex. 2052 ¶¶ 17–21; *see also, e.g.*, Prelim. Resp. 30–32.

Allegedly inconsistent positions

Patent Owner also argues that Kamal (Ex. 1037) establishes that cleavage was 93% and argues that such a cleavage rate (as a percent) is

nucleotide analogues. We also invite briefing on (1) whether or not the parties take the position that either challenged claim necessarily requires sequencing, for example, of whole genomes as Dr. Menchen discusses and (2) whether or not the claims otherwise require that recited steps be repeated.

As explained above, Petitioner sufficiently establishes sufficiently quantitative cleavage. To the extent that Petitioner has previously argued to the Board that over 97% cleavage is required, the full quotation from Exhibit 2029 is: “The experts for both parties agreed that SBS requires that the 3'-protecting group of the nucleotides be removed with greater than 90%

IPR2018-00797,
Institution Dec., 26

Illumina Previously Conceded That SBS Should Be Able to Sequence 20 Bases

Case: 15-1123 Document: 27 Page: 1 Filed: 03/10/2015

No. 2015-1123

IN THE
United States Court of Appeals
FOR THE FEDERAL

ILLUMINA CAMB

v.

INTELLIGENT BIO-S

APPEAL FROM THE UNITED STATES PATENT TRIAL AND APPEAL BOARD

BRIEF OF PATENT OWNERS
ILLUMINA CAMB

BRENTON R. BABCOCK
KNOBBE, MARTENS, OLSON
& BEAR, LLP
2040 Main Street, 14th Fl.
Irvine, CA 92614
(949) 760-0404

KERRY S. TAYLOR, PH.D.
NATHANIEL R. LUMAN, PH.D.
KNOBBE, MARTENS, OLSON
& BEAR, LLP
12790 El Camino Real
San Diego, CA 92130
(858) 707-4000

March 10, 2015

WILLIAM J. KNOBBE
& BEAR, LLP
1717 Pennsylvania Avenue N.W.
Suite 900
Washington, DC 20006
(202) 640-6400

Attorneys for Appellant

Columbia Exhibit 2029
Illumina, Inc. v. The Trustees of Columbia
University in the City of New York
IPR2018-00291

Case: 15-1123 Document: 27 Page: 16 Filed: 03/10/2015

determined in a stepwise fashion. JA66 at 2-65 JA67 at 3-11. A generic

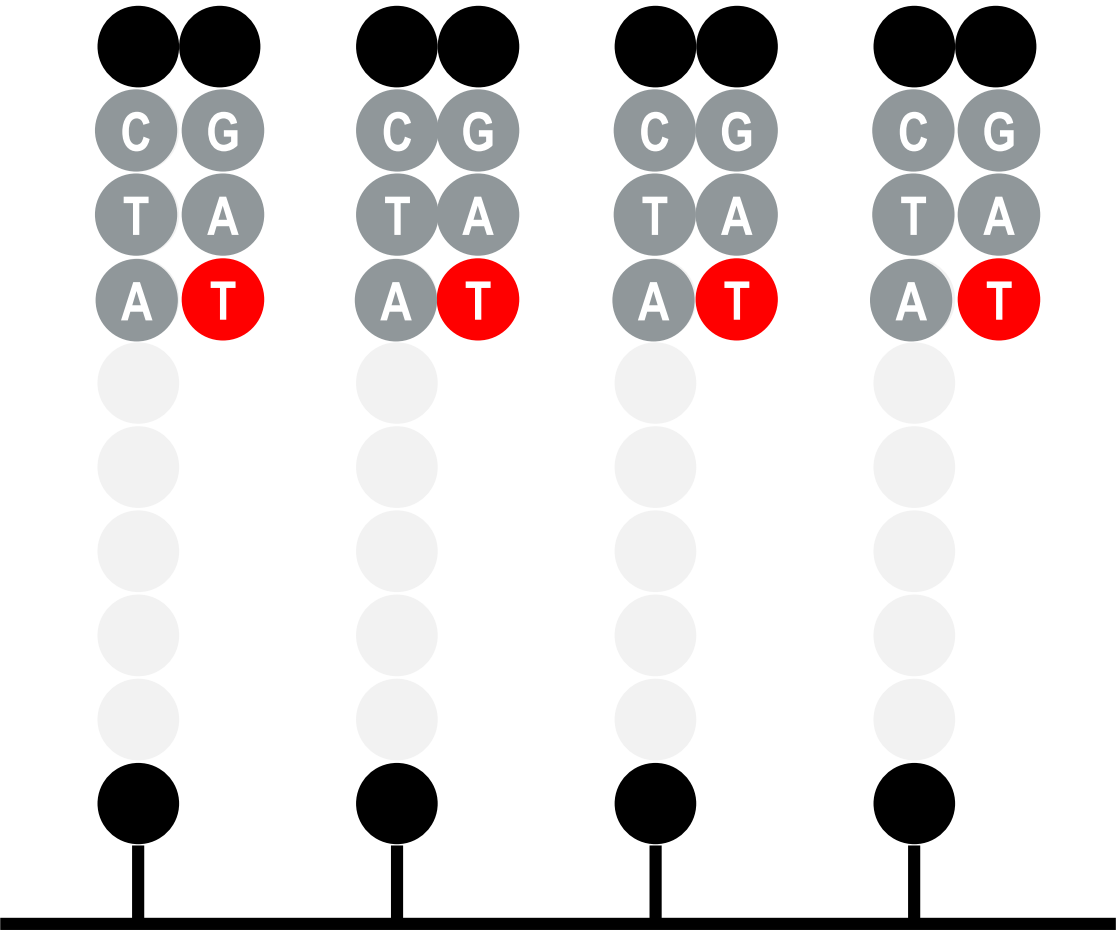
repeated for each and every nucleotide in the template DNA strand. Due to the complexity of DNA sequencing methods, an SBS process should be able to determine the sequence of at least 20 consecutive nucleotides in the template DNA strand to be effective. JA4983 at 11.9-12; JA2866 at 1.21-JA2867 at 1.25;

Ideally, this process of incorporation, identification, and deblocking is repeated for each and every nucleotide in the template DNA strand. Due to the complexity of DNA sequencing methods, an SBS process should be able to determine the sequence of at least 20 consecutive nucleotides in the template DNA strand to be effective. JA4983 at 11.9-12; JA2866 at 1.21-JA2867 at 1.25; JA2899 at 1.14-JA2900 at 1.6. The prior art taught SBS methods that achieved much greater sequence lengths. For example, WO 91/06678 ("Tsien") describes in detail SBS methods that allow sequencing of "25 to 300, or more" consecutive nucleotides in the template DNA strand. JA1501 at 11.33-36. Since

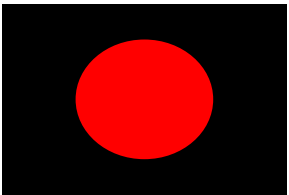
6

Ex. 2029, 6

Efficient Incorporation

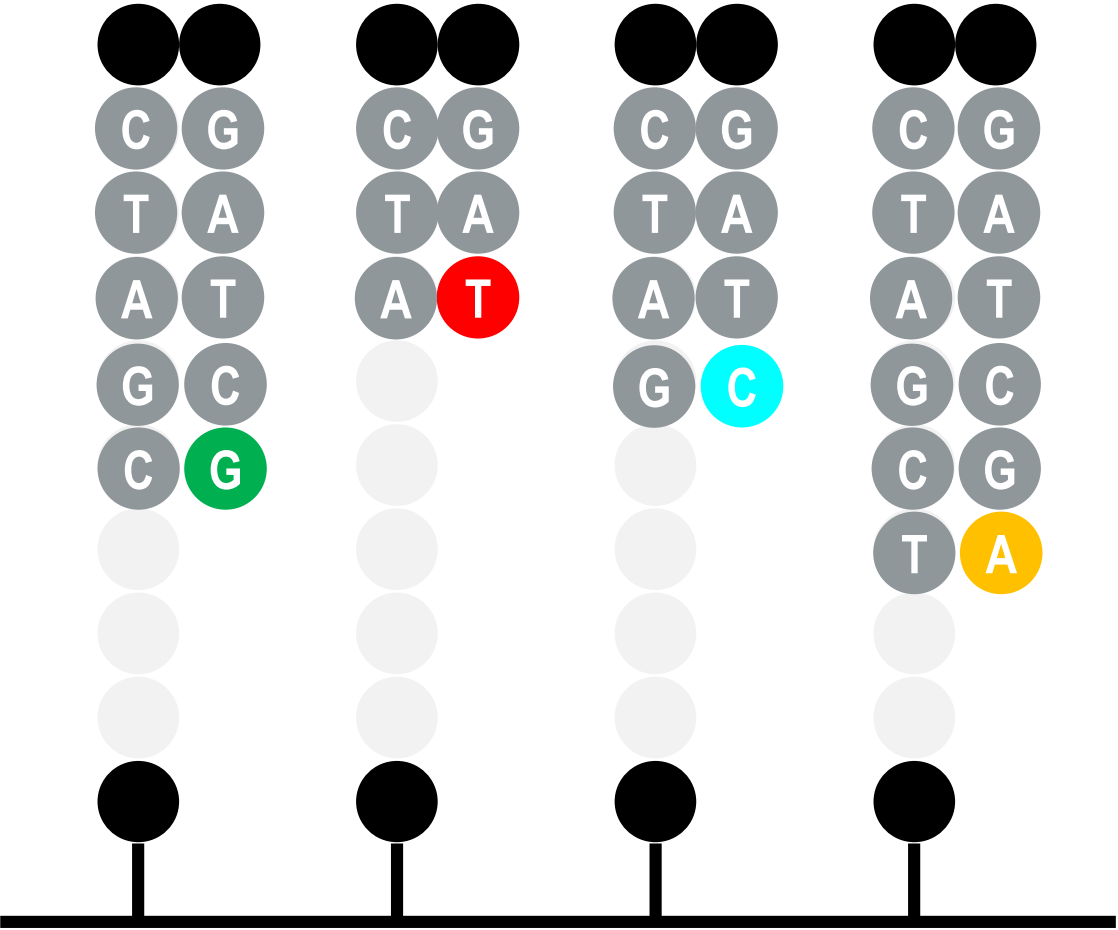


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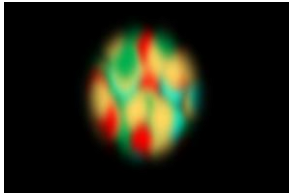


Incorporated Base = **T**

Out of Phase



Label Detection:



Incorporated Base = ?

ADDITIONAL SLIDES

A POSA Would Consult Welch 1999, Which Is Directed to SBS (BASS) and Does Not Teach Using Small Capping Groups (max 3.7Å)

FULL PAPER

Syntheses of Nucleosides Designed for Combinatorial DNA Sequencing

Mike B. Welch,^[a] Carlos I. Martinez,^[b] Alex J. Zhang,^[a] Song Jin,^[a] Richard Gibbs,^[b] and Kevin Burgess^{*,[a]}

Abstract: Nucleoside triphosphates **1** with 3'-O-blocking groups that are both photolabile and fluorescent were required to investigate the viability of a combinatorial fashion (see Figure 1). Four compounds were prepared to realize this goal. Two of the danyl-functional benzyl ether gr two, **18a** and **18b** carbonic group tion of these anal

by using five different DNA replicating enzymes, but the analogues were not incorporated. These results were surprising in view of the fact that previous studies had shown that 3'-O-(2'-nitrobenzyl)adenosine triphosphate **II** was incorporated by *E. coli* DNA polymerase crystal structure as a model demonstrates that analogues **14a**, **14i**, **18a** and **18i** are too large to fit into the enzyme active site, whereas accommodation of the unsubstituted 2-nitrobenzyl compound **II** is much less demanding. We conclude that both the nucleo-



T7 Polymerase/Primer-Template/ 3'-O-modified dNTP

Complete DNA + costly and time-consuming large. Most of the cu technological im Gilbert^[3] scheme systems for proc cleotides on com chines⁵⁻⁷ couple electrophoresis is trophoresis is an nevertheless they factors to accepti tional sequencing require gel elect hecamer strings wherein the appo cleotide library¹⁰ remains to be prc

[a] Prof. Dr. K. Burgess, Texas A & M University, Chemistry Department, P.O. Box 300012, Department of M Baylor College of Medicine, TX 77030

Chem. Eur. J. 1999, 5, 1

FULL PAPER

K. Burgess et al.

Conclusions

3'-O-Blocked nucleoside triphosphates, wherein the 3'-protecting group is both photolabile and fluorescent, are synthetically accessible. However, they tend to be too big to fit into the active site of DNA polymerases as evidenced by the data from the activity screens and the molecular-simulation experiments. It seems clear that modifications to the polymerase enzyme as well as to the nucleoside triphosphate are required if BASS is to be developed into a viable sequencing scheme. There are indications that modern methods for combinatorial mutagenesis of proteins, gene shuffling,^[48] have the potential to overcome this obstacle, but those methods must be refined further before this goal can be realized.

Experimental Section

General procedures: High-field NMR spectra were recorded on a Unity-300 spectrometer (H at 300 MHz, ¹³C at 75 MHz). ¹H chemical shifts are reported in δ relative to CDCl₃ (7.26 ppm) and ¹³C chemical shifts are reported in ppm relative to CDCl₃ (77.0 ppm) unless otherwise specified. Multiplicities in ¹H NMR are reported as (br) broad, (s) singlet, (dd) doublet, (t) triplet, (q) quartet, (m) multiplet, and (m) multiplet.

J = 10.5, 1.5 Hz, 1H), 4.76 (brs, 1H), 4.55–4.50 (m, 3H), 4.30 (t, *J* = 6.3 Hz, 2H), 4.00–3.80 (m, 3H), 3.16 (q, *J* = 6.6 Hz, 2H), 2.35 (ddd, *J* = 13.2, 5.7, 2.4 Hz, 1H), 2.22–2.12 (m, 1H), 1.75 (p, *J* = 7.2 Hz, 2H), 1.63 (s, 3H), 1.59–1.38 (m, 4H), 1.06 (s, 9H); ¹³C NMR (CDCl₃, 75 MHz): δ = 160.3, 160.2, 156.3, 150.7, 148.8, 136.9, 135.5, 125.2, 134.1, 133.8, 132.8, 132.2, 130.5, 130.2, 130.1, 128.4, 128.0, 127.9, 125.9, 117.6, 110.3, 87.0, 85.4, 72.0, 65.5, 64.0, 41.6, 41.1, 40.8, 29.6, 28.2, 27.0, 23.1, 19.3, 12.8; HRMS (positive-ion FAB, nitrobenzyl alcohol (NBA)): calcd for C₂₄H₂₆N₂O₈SiNa 451.3299, found 451.3327.

3'-O-(4'-N-Allyloxycarbonyl)-5'-amino-1-yl-oxycarbonyl)-2'-nitrophenylmethyl)thymidine (4): Compound **3** (700 mg, 0.84 mmol), THF (8 mL), and tetrabutylammonium fluoride (TBAF) (1.26 mL, 1.26 mmol) were stirred at 25 °C for 15 min. The reaction was concentrated and purified directly by flash chromatography with a gradient of 100% EtOAc, then 1 to 5% MeOH in EtOAc as the eluent to give **4** as a foam (589 mg, 88% yield). *R_f* = 0.11 (75% EtOAc/hexanes); IR (neat): ν = 3479, 2947, 1720, 1642 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): δ = 8.54 (d, *J* = 1.8 Hz, 1H), 8.06 (dd, *J* = 8.1, 1.5 Hz, 1H), 7.60 (s, 1H), 7.24 (d, *J* = 8.1 Hz, 1H), 6.17 (t, *J* = 6.6 Hz, 1H), 5.89–5.76 (m, 1H), 5.44 (s, 2H), 5.21 (d, *J* = 1.4 Hz, 1H), 5.12 (d, *J* = 10.5 Hz, 1H), 4.97 (br, 1H), 4.47–4.41 (m, 3H), 4.28 (t, *J* = 6.3 Hz, 2H), 4.00 (t, *J* = 6.6 Hz, 2H), 3.89 (br, 1H), 3.81–3.69 (m, 2H), 3.12 (q, *J* = 6.6 Hz, 2H), 3.95 (br, 1H), 2.25–2.19 (m, 2H), 1.99 (s, 3H), 1.74 (p, *J* = 7.5 Hz, 2H), 1.59–1.35 (m, 4H); ¹³C NMR (CDCl₃, 75 MHz): δ = 164.3, 163.2, 156.4, 150.7, 148.7, 136.8, 135.5, 133.9, 132.8, 130.5, 128.4, 125.8, 117.5, 110.1, 87.0, 86.5, 71.1, 65.6, 65.4, 62.1, 41.5, 40.7, 40.3, 29.5, 28.2, 23.0, 13.1; HRMS (positive-ion FAB, NBA): calcd for C₂₄H₂₆N₂O₈Na 463.2122, found 463.2122.

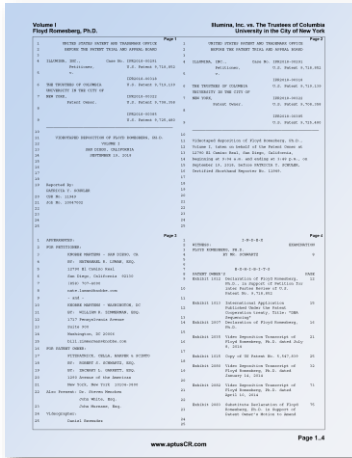
3'-O-(4'-N-Allyloxycarbonyl)-5'-amino-1-yl-oxycarbonyl)-2'-nitrophenylmethyl)thymidine (4): Compound **3** (700 mg, 0.84 mmol), THF (8 mL), and tetrabutylammonium fluoride (TBAF) (1.26 mL, 1.26 mmol) were stirred at 25 °C for 15 min. The reaction was concentrated and purified directly by flash chromatography with a gradient of 100% EtOAc, then 1 to 5% MeOH in EtOAc as the eluent to give **4** as a foam (589 mg, 88% yield). *R_f* = 0.11 (75% EtOAc/hexanes); IR (neat): ν = 3479, 2947, 1720, 1642 cm⁻¹; ¹H NMR (CDCl₃, 300 MHz): δ = 8.54 (d, *J* = 1.8 Hz, 1H), 8.06 (dd, *J* = 8.1, 1.5 Hz, 1H), 7.60 (s, 1H), 7.24 (d, *J* = 8.1 Hz, 1H), 6.17 (t, *J* = 6.6 Hz, 1H), 5.89–5.76 (m, 1H), 5.44 (s, 2H), 5.21 (d, *J* = 1.4 Hz, 1H), 5.12 (d, *J* = 10.5 Hz, 1H), 4.97 (br, 1H), 4.47–4.41 (m, 3H), 4.28 (t, *J* = 6.3 Hz, 2H), 4.00 (t, *J* = 6.6 Hz, 2H), 3.89 (br, 1H), 3.81–3.69 (m, 2H), 3.12 (q, *J* = 6.6 Hz, 2H), 3.95 (br, 1H), 2.25–2.19 (m, 2H), 1.99 (s, 3H), 1.74 (p, *J* = 7.5 Hz, 2H), 1.59–1.35 (m, 4H); ¹³C NMR (CDCl₃, 75 MHz): δ = 164.3, 163.2, 156.4, 150.7, 148.7, 136.8, 135.5, 133.9, 132.8, 130.5, 128.4, 125.8, 117.5, 110.1, 87.0, 86.5, 71.1, 65.6, 65.4, 62.1, 41.5, 40.7, 40.3, 29.5, 28.2, 23.0, 13.1; HRMS (positive-ion FAB, NBA): calcd for C₂₄H₂₆N₂O₈Na 463.2122, found 463.2122.

Conclusions

3'-O-Blocked nucleoside triphosphates, wherein the 3'-protecting group is both photolabile and fluorescent, are synthetically accessible. However, they tend to be too big to fit into the active site of DNA polymerases as evidenced by the data from the activity screens and the molecular-simulation experiments. It seems clear that modifications to the polymerase enzyme as well as to the nucleoside triphosphate are required if BASS is to be developed into a viable sequencing scheme. There are indications that modern methods for combinatorial mutagenesis of proteins, gene shuffling,^[48] have the potential to overcome this obstacle, but those methods must be refined further before this goal can be realized.

Ex. 1023, 956

No Expectation That Small Capping Group Met Tsien's Requirements for SBS



Floyd Romesberg, Ph.D. Deposition

Q. ...It is your opinion that a person of ordinary skill in the art **would not have had an expectation** that a small and removable protecting group would also possess all of the listed properties of Tsien's protecting group?

A. Yes.

Ex. 2126, Romesberg Dep., 82:21-83:2

No Expectation a 3'-O-Allyl Thymine, Cytosine or Guanine Nucleotide Would be Incorporated

	AMV-RT	M-MuLV-RT	<i>Bst</i>	AmpliTaq	Vent(exo-)	<i>rTth</i>
3'-O-methyl-dATP						
3'-O-methyl-dTTP						

A Compound and All of Its Properties Are Inseparable

Application of Papesch, 50 C.C.P.A. 1084 (1963)
315 F.2d 381, 137 U.S.P.Q. 43

KeyCite Yellow Flag - Negative Treatment
Distinguished by Scheda International Limited v. Actavis Laboratories
FL, Inc., D.N.J., November 21, 2016

315 F.2d 381

United States Court of Customs and Patent Appeals.

Application of VIKTOR PAPESCH.

Patent Appeal No. 6882.

March 20, 1963.

Synopsis

From a decision of the Patent Office Board of Appeals, Serial No. 836,870 rejecting the applicant's claims for chemical compounds, the applicant appealed. The United States Court of Customs and Patent Appeals, Rich, Judge, held that the Board of Appeals erred in failing to consider a biological or pharmaceutical property of the compounds on the ground that to chemists the structure of the compounds would be so obvious as to be beyond doubt and that showing of such properties is to be used only to resolve doubt.

Reversed.

Attorneys and Law Firms

*382 *1085 Helmut A. Wegner, Chicago, Ill., for appellant.

Clarence W. Moore, Washington, D.C. (Raymond E. Martin, Washington, D.C., of counsel), for Commissioner of Patents.

Before WORLEY, Chief Judge, and RICH, MARTIN, SMITH and ALMOND, judges.

Opinion

RICH, Judge.

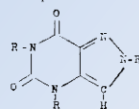
*1086 This appeal is from the decision of the Patent Office Board of Appeals affirming the rejection of claims 1-3, the only claims presented in appellant's application Ser. No. 836,870, filed August 31, 1959, for "2,4,6-TRIALKYLPIRAZOLO (4,3-d)-4,5,6,7-TETRAHYDROPYRIMIDINE-5,7-DIONES."

The specification, which is brief and occupies less than three pages of the printed record, states:

'The trialkyl compounds of this invention have been found to possess unexpectedly potent anti-inflammatory activity in contrast to the related trimethyl compound. The instant compounds are also diuretic agents.'

Claim 1 reads:

'A compound of the structural formula



wherein R is a lower alkyl radical containing more than one and less than five carbon atoms.'

From the standpoint of patent law, a compound and all of its properties are inseparable; they are one and the same thing. The graphic formulae, the chemical nomenclature,

'Robins et al., J. Am. Chem. Soc., Vol. 78, pp. 2418-2422 (1956)'

The action is so brief and to the point that we quote it in full:

'Claims 1-3 are rejected as being unpatentable over Robins et al. Note Compound XVI. The ethyl and n-butyl side chains depicted in applicant's claims 2 and 3 are obvious homologs of the methyl groups shown in identical positions in the reference compound and the method of preparation is substantially the same. (In re Henze, 636 O.G. 698).'

Compound XVI of the Robins et al. article is described by its structural formula which would be identical with that of appellant's *1087 claim 1, supra, if all of the three

In re Papesch, 315 F.2d 381, 391 (C.C.P.A. 1963)