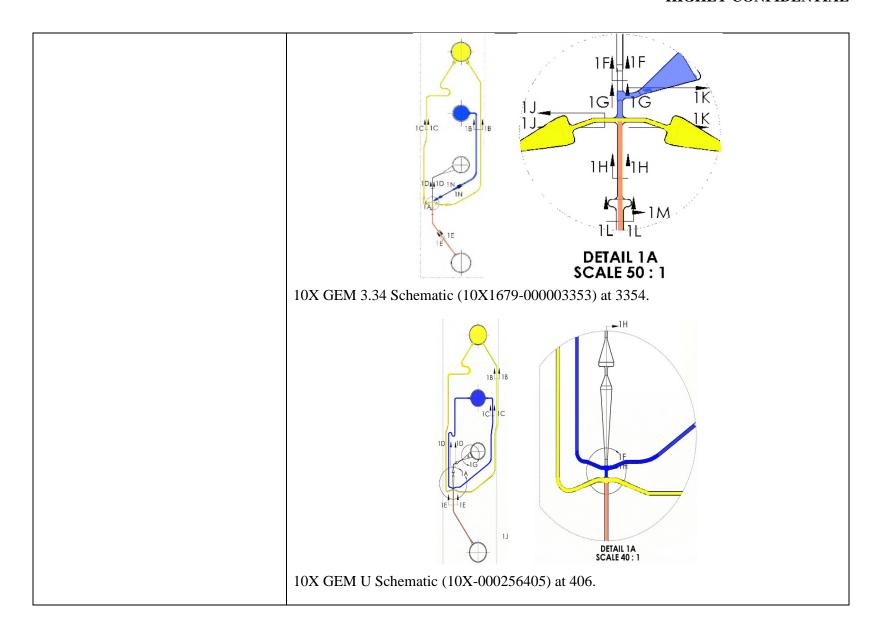
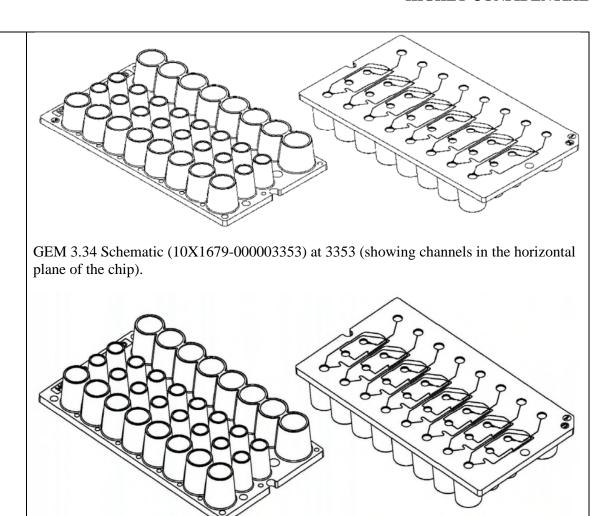
Exhibit B: Initial Infringement Claim Chart – U.S. Patent No. 9,562,837

Claim Language '837 Patent	10X's Double Junction Accused Products
Claim 1[a]. An assembly, the assembly comprising:	To the extent the preamble is limiting, the 10X Double Junction Accused Products literally infringe this claim element because they are an assembly:
	GEM 3.34 Schematic (10X1679-000003353) at 3353 (showing channels in the horizontal plane of the chip)

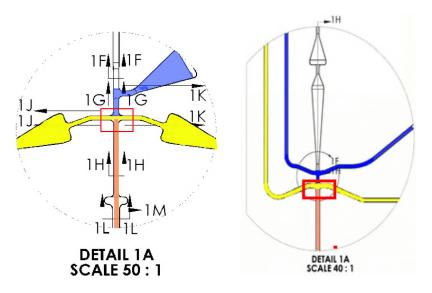
	10X GEM U Schematic (10X-000256405) at 406.
	Single Cell 3' v3 User Guide (10X1679-000002059) at 2084; Single Cell V(D)J User Guide (10X1679-000002997) at 3027; Single Cell ATAC User Guide (10X1679-000001131) at 1158; Genome User Guide v2 (10X1679-000000679) at 710.
[b] a microchannel in a horizontal plane;	The 10X Double Junction Accused Products literally infringe this element because they include a microchannel (orange) that is in the horizontal plane of the chip:



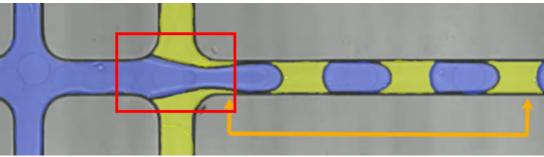


10X GEM U Schematic (10X-000256405) at 406.

[c] at least one droplet formation module comprising a sample inlet, an immiscible fluid inlet, and a junction, The 10X Double Junction Accused Products literally infringe this element because they include at least one droplet formation module comprising a sample inlet (blue), an immiscible fluid inlet (yellow), and a junction (red square):

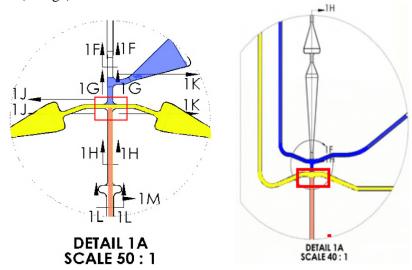


10X GEM 3.34 Schematic (10X1679-000003353) at 3354; 10X GEM U Schematic (10X-000256405) at 406.



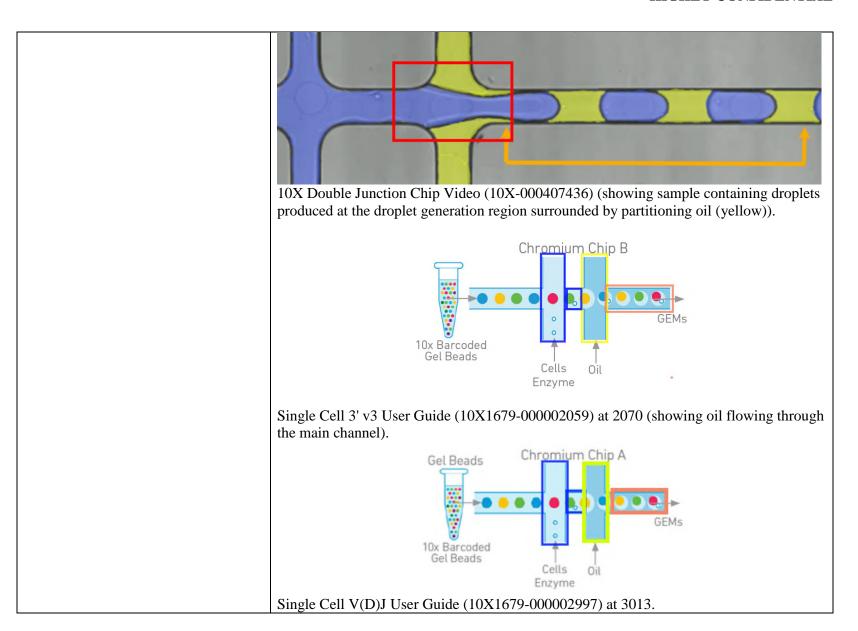
10X Double Junction Chip Video (10X-000407436).

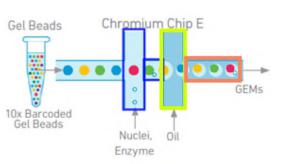
[d] wherein the junction is located between the sample inlet and the microchannel and the droplet formation module is configured to produce droplets comprising the sample surrounded by the immiscible fluid; and The 10X Double Junction Accused Products literally infringe this element because they include a junction (red square) located between the sample inlet (blue) and the microchannel (orange):



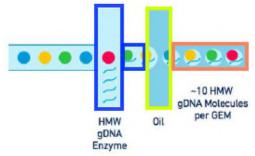
10X GEM 3.34 Schematic (10X1679-000003353) at 3354; 10X GEM U Schematic (10X-000256405) at 406.

The droplet formation module comprises a sample inlet (blue), an immiscible fluid inlet (yellow), and a junction (red square), as shown above and described in element 1(c), that is configured to produce droplets containing sample and surrounded by a partitioning oil (immiscible fluid):





Single Cell ATAC User Guide (10X1679-000001131) at 1143.

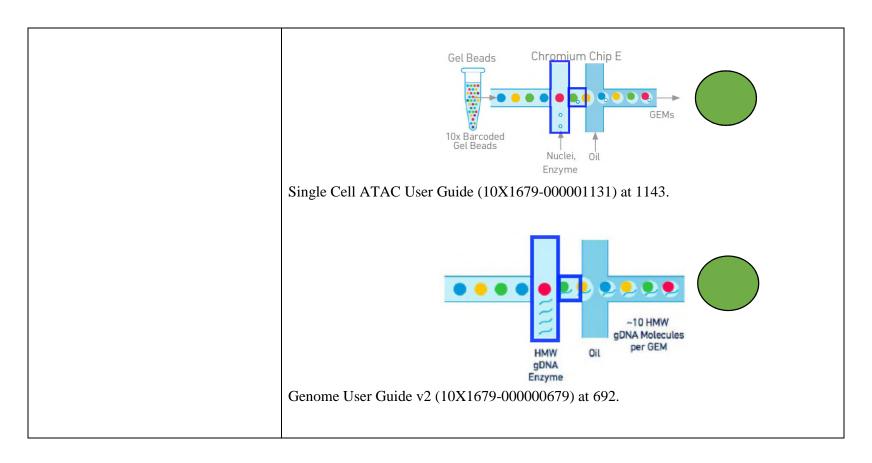


Genome User Guide v2 (10X1679-000000679) at 692.

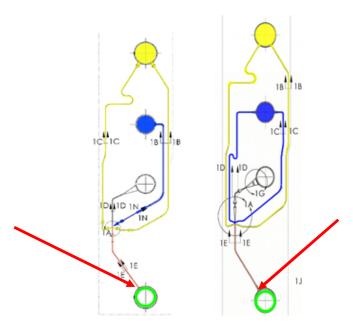
See also Single Cell 3' v3 User Guide (10X1679-000002059) at 2070 ("It is done by partitioning thousands of cells into nanoliter-scale Gel Beads-in-emulsion (GEMs)"); *id.* ("GEMs are generated by combining barcoded Single Cell 3' v3 Gel Beads, a Master Mix containing cells and Partitioning Oil."); Single Cell V(D)J User Guide (10X1679-000002997) at 3013 ("It does so by partitioning thousands of cells into nanoliter-scale Gel Beads-in-emulsion (GEMs)."); *id.* ("GEMs are generated by combining barcoded Single Cell 5' Gel Beads, a Master Mix with cells, and Partitioning Oil on Chromium Chip A.");

See also Single Cell ATAC User Guide (10X1679-000001131) at 1143 ("This is achieved by transposing nuclei in bulk solution; then using a microfluidic chip, the nuclei are partitioned into nanoliter-scale Gel Beads-in-emulsion (GEMs)."); *id.* ("GEMs are

generated by combining barcoded Gel Beads, transposed nuclei, a Master Mix, and Partitioning Oil on a Chromium Chip E."). See also Genome User Guide v2 (10X1679-000000679) at 692 ("In the microfluidic Genome Chip, a library of Genome Gel beads is combined with an optimal amount of HMW template gDNA in Master Mix and partitioning oil to create Gel Bead-in-Emulsions (GEMs)."); id. at 706 ("[S]lowly transfer 90 µl Sample Master Mix-denatured gDNA mixture into the bottom of wells in the row labeled 1."); id. at 750 ("aqueous sample volume"). The 10X Double Junction Accused Products literally infringe this element because they [e] at least one downstream separation chamber comprising a droplet include at least one downstream separation chamber (green circle). receiving chamber inlet and at least one droplet receiving outlet, Chromium Chip B **GEMs** 10x Barcoded Gel Beads Cells Oil Enzyme Single Cell 3' v3 User Guide (10X1679-000002059) at 2070 (showing droplets flowing towards a separation chamber). Chromium Chip A Gel Beads **GEMs** 10x Barcoded Gel Beads Cells Oil Enzyme Single Cell V(D)J User Guide (10X1679-000002997) at 3013.

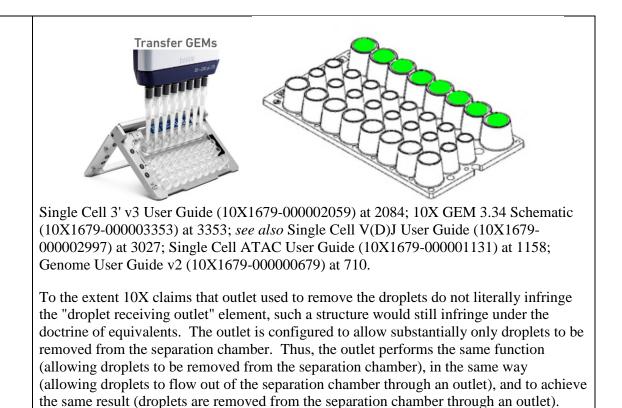


The separation chamber (green) comprising a droplet receiving chamber inlet, identified below by the red arrow as the interface between the microchannel (orange) and separation chamber (green) below. At this interface, the droplets enter the separation chamber.



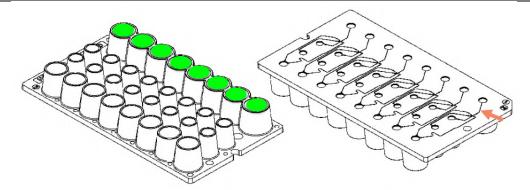
10X GEM 3.34 Schematic (10X1679-000003353) at 3354; 10X GEM U Schematic (10X-000256405) at 406.

The separation chamber also comprises a droplet receiving outlet for removing the droplets from the separation chamber following droplet generation, which is shown below as the location where the pipette is inserted to remove the "GEMs":

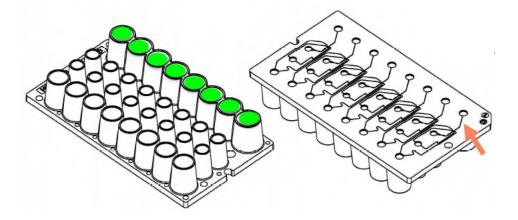


[f] wherein the separation chamber is upright to the microchannel and out of the horizontal plane;

The 10X Double Junction Accused Products literally infringe this element because the separation chamber (green) is upright to the microchannel (orange arrow), and out of the horizontal plane:

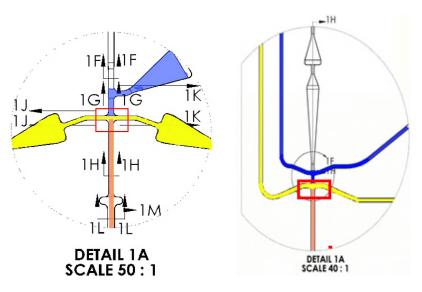


10X GEM 3.34 Schematic (10X1679-000003353) at 3353.

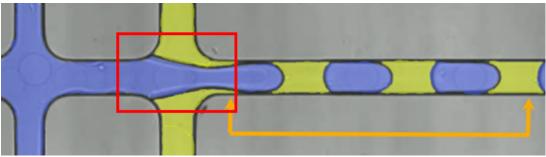


10X GEM U Schematic (10X-000256405) at 405; *see also* Single Cell 3' v3 User Guide (10X1679-000002059) at 2076, 2084 (showing separation chamber sticking out of the top surface of the chip); Single Cell V(D)J User Guide (10X1679-000002997) at 3019, 3027 (showing separation chamber sticking out of the top surface of the chip); Single Cell ATAC User Guide (10X1679-000001131) at 1147, 1158 (showing separation chamber sticking out of the top surface of the chip); Genome User Guide v2 (10X1679-000000679) at 710, 742 (showing separation chamber sticking out of the top surface of the chip).

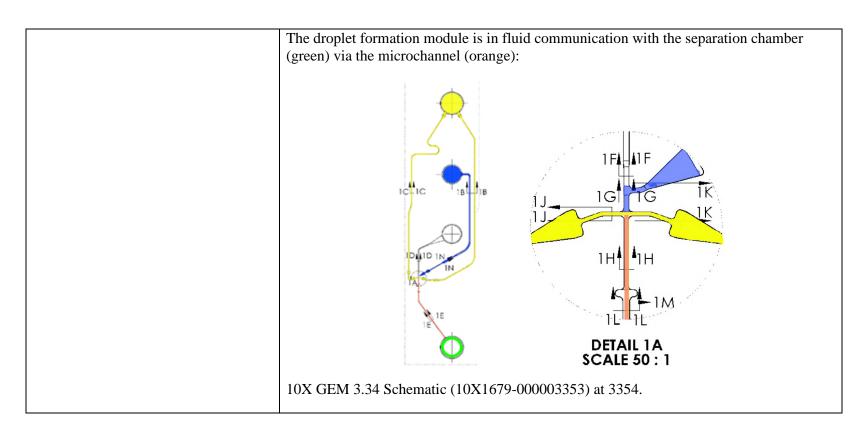
[g] wherein the droplet formation module and the separation chamber are in fluid communication with each other via the microchannel; and The 10X Double Junction Accused Products literally infringe this element because they include a droplet formation module, comprising a sample inlet (blue), an immiscible fluid inlet (yellow), and a junction (red square), as shown below and described in element 1(c):

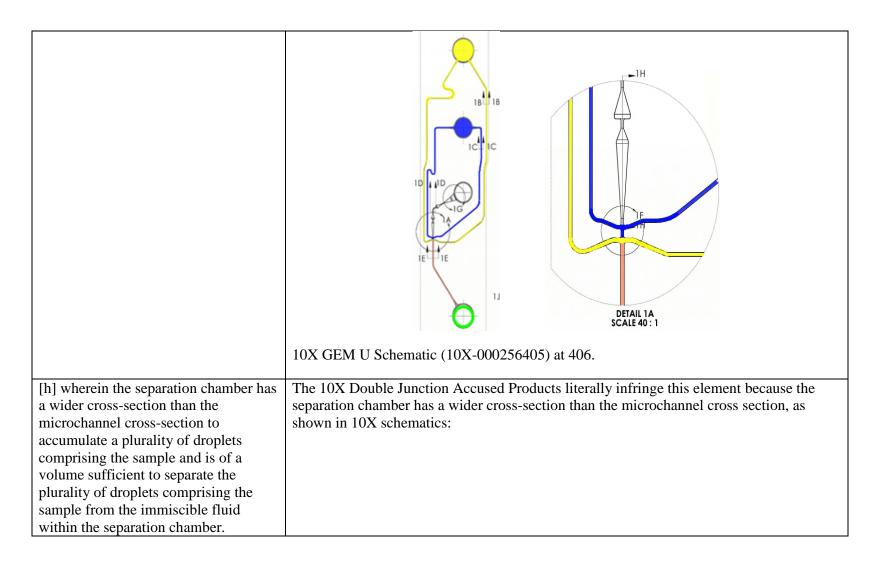


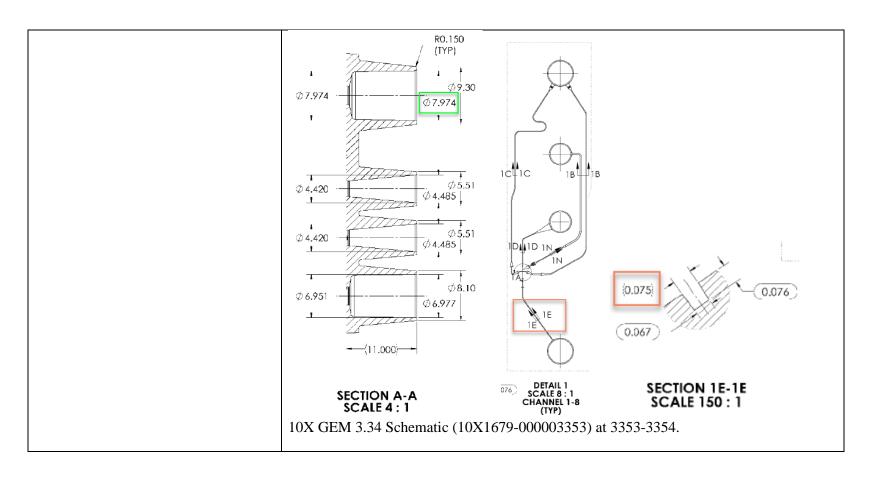
10X GEM 3.34 Schematic (10X1679-000003353) at 3354; 10X GEM U Schematic (10X-000256405) at 406.

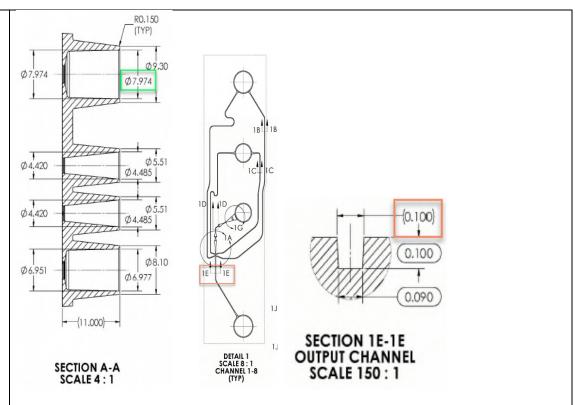


10X Double Junction Chip Video (10X-000407436).









10X GEM U Schematic (10X-000256405) at 405-406.

See also Single Cell 3' v3 User Guide (10X1679-000002059) at 2084 (showing pipettes used to "Transfer GEMs" from the separation chamber); Single Cell V(D)J User Guide (10X1679-000002997) at 3027 (showing pipettes used to "Transfer GEMs" from the separation chamber); Single Cell ATAC User Guide (10X1679-000001131) at 1158 (showing pipettes used to "Transfer GEMs" from the separation chamber); Genome User Guide v2 (10X1679-000000679) at 710 (showing pipettes used to transfer GEMs from the separation chamber).

Within the separation chamber, a plurality of droplets comprising sample are accumulated. *See* Single Cell 3' v3 User Guide (10X1679-00002059) at 2070 ("It is done by partitioning thousands of cells into nanoliter-scale Gel Beads-in-emulsion (GEMs)."); *id*. ("GEMs are generated by combining barcoded Single Cell 3' v3 Gel Beads, a Master Mix containing cells and Partitioning Oil."); Single Cell V(D)J User Guide (10X1679-000002997) at 3013 ("It does so by partitioning thousands of cells into nanoliter-scale Gel Beads-in-emulsion (GEMs)."); *id*. ("GEMs are generated by combining barcoded Single Cell 5' Gel Beads, a Master Mix with cells, and Partitioning Oil on Chromium Chip A.").

See also Single Cell ATAC User Guide (10X1679-000001131) at 1201 ("This is achieved by transposing nuclei in bulk solution; then using a microfluidic chip, the nuclei are partitioned into nanoliter-scale Gel Beads-in-emulsion (GEMs)."); *id.* ("GEMs are generated by combining barcoded Gel Beads, transposed nuclei, a Master Mix, and Partitioning Oil on a Chromium Next GEM Chip H.").

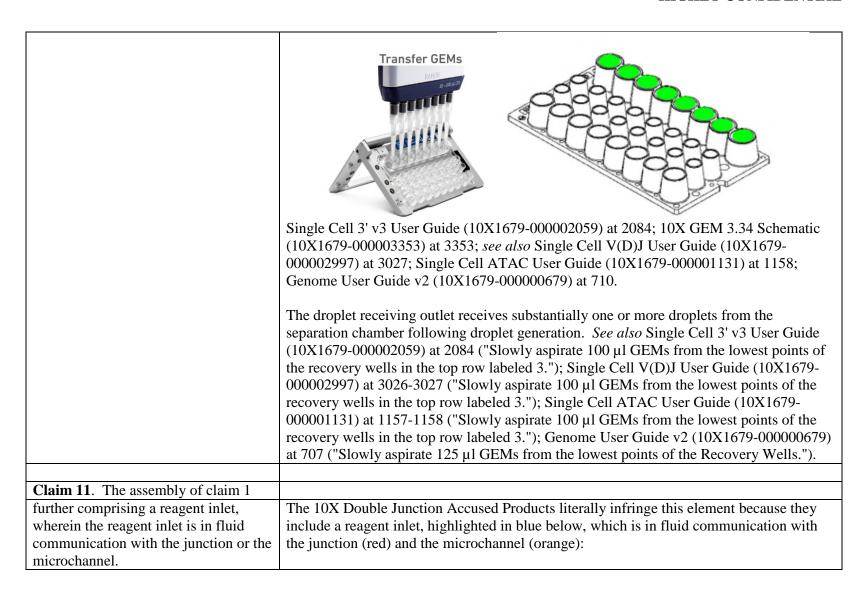
See also Genome User Guide v2 (10X1679-000000679) at 692 ("In the microfluidic Genome Chip, a library of Genome Gel Beads is combined with an optimal amount of HMW template gDNA in Master Mix and partitioning oil to create Gel Beads-in-Emulsions (GEMs)."); *id.* "Approximately 1.25-1.5 ng (~375 – 450 haploid genome equivalents) of template gDNA is partitioned across over 1 million GEMs.").

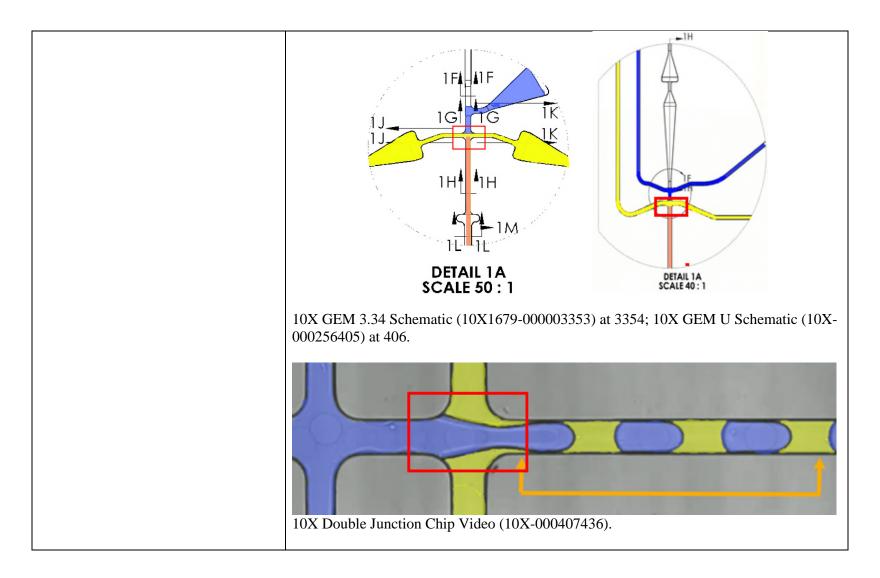
The volume in the separation chamber is sufficient to separate the plurality of droplets comprising the sample from the immiscible fluid. *See* Single Cell 3' v3 User Guide (10X1679-000002059) at 2084 ("Slowly aspirate 100 µl GEMs from the lowest points of the recovery wells in the top row labeled 3" after loading "75 µl of Master Mix + Cell Suspension," "40 µl of Gel beads," and "280 µl of Partitioning Oil"); Single Cell V(D)J User Guide (10X1679-000002997) at 3026-3027 ("Slowly aspirate 100 µl GEMs from the lowest points of the recovery wells in the top row labeled 3" after loading 90 µl of "Master Mix + Cell Suspension," "40 µl of Gel beads," and "270 µl of Partitioning Oil"); Single Cell ATAC User Guide (10X1679-000001131) at 1157-1158 ("Slowly aspirate 100 µl GEMs from the lowest points of the recovery wells in the top row labeled 3" after loading "75 µl of Master Mix + Transposed Nuclei," "40 µl of Gel beads," and "240 µl of Partitioning Oil"); Genome User Guide v2 (10X1679-000000679) at 707-709 ("Slowly

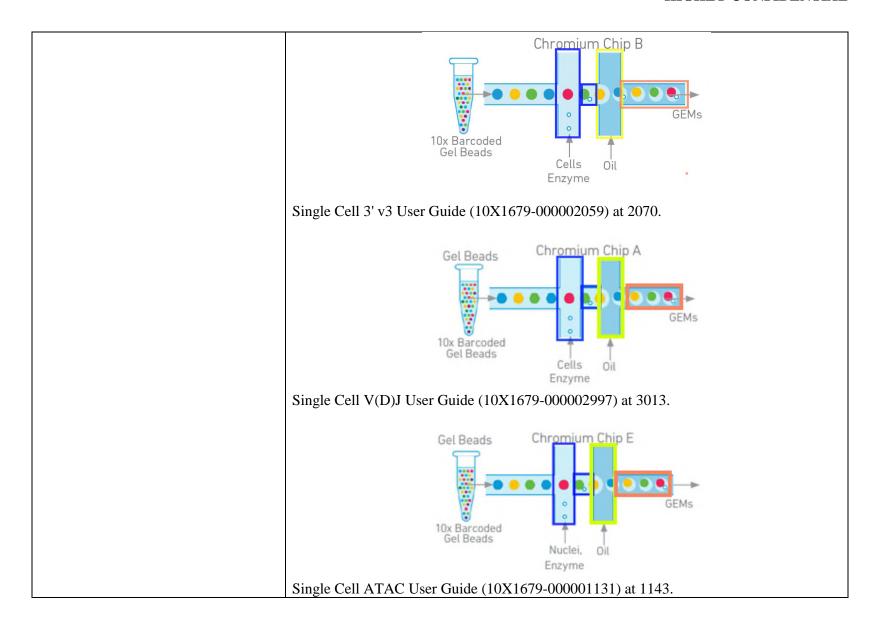
	aspirate 125 µl GEMs from the lowest points of the Recovery Wells" after loading "90 µl Sample Master Mix-denatured gDNA mixture," "85 µl Gel Beads," and "270 µl Partitioning Oil").
Claim 2. The assembly of claim 1,	See Claim 1.
wherein an aqueous fluid from the sample inlet is segmented with an immiscible fluid from the immiscible fluid inlet at the junction.	The 10X Double Junction Accused Products literally infringe this element because they include at least one droplet formation module comprising a sample inlet (blue), an immiscible fluid inlet (yellow), and a junction (red square) where aqueous fluid (shown in blue) is segmented with immiscible fluid (shown in yellow) from the immiscible fluid inlet at the junction as shown below and in element 1(c): 10X Double Junction Chip Video (10X-000407436).
Claim 4 . The assembly of claim 1,	See Claim 1.
wherein the at least one droplet receiving outlet is located on the upper portion of the separation chamber.	The 10X Double Junction Accused Products literally infringe this element because the droplet receiving outlet is located on the upper portion of the separation chamber, which is shown below as the location where the pipette is inserted to remove the "GEMs":

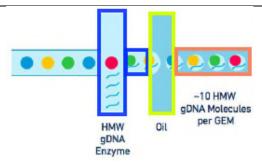
	Single Cell 3' v3 User Guide (10X1679-000002059) at 2084; 10X GEM 3.34 Schematic (10X1679-000003353) at 3353; see also Single Cell V(D)J User Guide (10X1679-000002997) at 3027; Single Cell ATAC User Guide (10X1679-000001131) at 1158; Genome User Guide v2 (10X1679-000000679) at 710.
Claim 7. The assembly of claim 1, wherein the droplet receiving outlet is coupled to a vessel.	See Claim 1. The 10X Double Junction Accused Products literally infringe this element because the droplet receiving outlet is located on the upper portion of the separation chamber, which is shown below as the location where the pipette is inserted to remove the "GEMs":

	Single Cell 3' v3 User Guide (10X1679-000002059) at 2084; 10X GEM 3.34 Schematic (10X1679-000003353) at 3353; see also Single Cell V(D)J User Guide (10X1679-000002997) at 3027; Single Cell ATAC User Guide (10X1679-000001131) at 1158; Genome User Guide v2 (10X1679-00000679) at 710. The droplet receiving outlet is coupled to a tube strip, which receives the droplets after droplet generation. See Single Cell 3' v3 User Guide (10X1679-000002059) at 2084 ("Over the course of ~20 sec, dispense GEMs into the tube strip on ice."); Single Cell V(D)J User Guide (10X1679-000002997) at 3027 ("Over the course of ~20 sec, dispense GEMs into the tube strip on ice."); Single Cell ATAC User Guide (10X1679-000001131) at 1158 ("Over the course of ~20 sec, dispense GEMs into the tube strip on ice.").
Claim 9 . The assembly of claim 7, wherein the vessel is a PCR tube or a test tube.	See Claim 7. See Claim 7.
Claim 10. The assembly of claim 1, wherein the droplet receiving outlet receives substantially one or more droplets.	See Claim 1. The 10X Double Junction Accused Products literally infringe this element because the droplet receiving outlet is located on the upper portion of the separation chamber, which is shown below as the location where the pipette is inserted to remove the "GEMs":









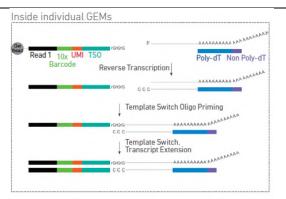
Genome User Guide v2 (10X1679-000000679) at 692.

See also Single Cell 3' v3 User Guide (10X1679-000002059) at 2070 ("GEMs are generated by combining barcoded Single Cell 3' v3 Gel Beads, a Master Mix containing cells, and Partitioning Oil into Chromium Chip B."); *id.* at 2283 ("[A]dd the appropriate volume of nuclease-free water and corresponding volume of single cell suspension to Master Mix for a total of 80 μ l in each tube."); *id.* ("Gently pipette mix the Master Mix + Cell Suspension and using the same pipette tip, dispense 75 μ l Master Mix + Cell Suspension into the bottom center of each well in row labeled 1.").

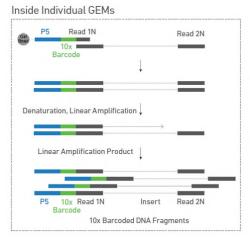
See also Single Cell V(D)J User Guide (10X1679-000002997) at 3013 ("GEMs are generated by combining barcoded Single Cell 5' Gel Beads, a Master Mix with cells, and Partitioning Oil on Chromium Chip A."); *id.* at ("[A]dd the appropriate volume of nuclease-free water first, followed by corresponding volume of single cell suspension to Master Mix for a total of $100~\mu l$ in each tube."); *id.* ("Gently pipette mix the Master Mix + Cell Suspension and using the same pipette tip, dispense $90~\mu l$ into the bottom center of each well in row labeled 1").

See also Single Cell ATAC User Guide (10X1679-000001131) at 1143 ("GEMs are generated by combining barcoded Gel Beads, transposed nuclei, a Master Mix, and Partitioning Oil on a Chromium Chip E."); id. at 1157 ("Using the same pipette tip, dispense 75 μ l Master Mix + Transposed Nuclei into the bottom center of each well in row labeled 1.").

	See also Genome User Guide v2 (10X1679-000000679) at 692 ("In the microfluidic
	Genome Chip, a library of Genome Gel beads is combined with an optimal amount of
	HMW template gDNA in Master Mix and partitioning oil to create Gel Bead-in-
	Emulsions (GEMs)."); id. at 706 ("[S]lowly transfer 90 µl Sample Master Mix-denatured
	gDNA mixture into the bottom of wells in the row labeled 1.").
Claim 12 . The assembly of claim 11,	See Claim 11
wherein the reagent inlet introduces	The 10X Double Junction Accused Products literally infringe this element because they
one or more amplification reagents to	include a reagent inlet. See Claim 11.
the junction or the microchannel.	
	The reagent inlet introduces one or more amplification reactions at the junction into each
	droplet and allows for amplification to occur within the droplets following droplet
	generation.
	Inside individual GEMs
	(Ge)
	Read 1 10x UMI Poly(dT)VN Barcode
	Poly-di Primer
	3AAAAAAAA
	Bear Team
	Reverse Transcription
	GCC
	Template Switch Oligo Priming
	TSO
	- ccc
	Template Switch, Transcript Extension
	** rororo
	See Single Cell 3' v3 User Guide (10X1679-000002059) at 2071 ("Immediately following
	GEM generation, the Gel Bead is dissolved, primers are released, and any co-partitioned
	cell is lysed. Primer containing an Illumina TruSeq Read 1 (read 1 sequencing primer), 16
	nt 10x Barcode, 12 nt unique molecular identifier (UMI), and 30 nt poly(dT) sequence are
	mixed with cell lysate and a Master Mix containing reverse transcription (RT) reagents.
	Incubation of the Gems produces barcoded, full-length cDNA from poly-adenylated
	mRNA.").



See also Single Cell V(D)J v1 User Guide (10X1679-000002997) at 3013 ("Immediately following GEM generation, the Gel Bead is dissolved and any co-partitioned cell is lysed. Oligonucleotides containing (i) an Illumina R1 sequence (read 1 sequencing primer), (ii) a 16 nt 10x Barcode, (iii) a 10 nt unique molecular identifier (UMI), and (iv) 13 nt template switch oligo (TSO) are released and mixed with the cell lysate and a Master Mix containing reverse transcription (RT) reagents and poly(dT) RT primers. Incubation of the GEMs produces 10x Barcoded, full-length cDNA from polyadenylated mRNA.").



See also Single Cell ATAC v1 User Guide (10X1679-000001131) at 1143 ("Upon GEM generation, the Gel Bead is dissolved. Oligonucleotides containing (i) an Illumina® P5

	sequence, (ii) a 16 nt 10x Barcode and (iii) a Read 1 (Read 1N) are released and mixed with DNA fragments and Master Mix. Thermal cycling of the GEMs produces 10x barcoded single-stranded DNA. After incubation, the GEMs are broken and pooled fractions are recovered.").
	See also Genome User Guide v2 (10X1679-000000679) at 692 ("Upon dissolution of the Genome Gel Bead in the GEM, primers containing (i) an Illumina R1 sequence (Read 1 sequencing primer), (ii) a 16 bp 10x Barcode, and (iii) a 6 bp random primer sequence are released. Isothermal incubation of the GEMs produces barcoded fragments ranging from a few to several hundred base pairs. After incubation, the GEMs are broken and the pooled fractions are recovered.").
Claim 13. The assembly of claim 12,	See Claim 12.
wherein the one or more amplification reagents are for a polymerase chain reaction.	See Claim 12.