International Journal of Pharmaceutics, 75 (1991) 97-115
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ADONIS 037851739100307L

LJP 02512

Invited Reviews

Peptide and protein drugs: I. Therapeutic applications, absorption and parenteral administration

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(Received 20 February 1991)

(Modified version received 4 May 1991)

(Accepted 10 May 1991)

Key words: Peptide delivery; Protein delivery; Stability; Bioavailability; Absorption barrier; Proteolytic activity; Absorption enhancer; Proteinase inhibitor; Liposome

Summary

In this first part of a two-part review of peptide and protein drugs, the pertinent terminology is introduced and the therapeutic applications of those drugs summarised. Their absorption and the methodology commonly used for study on it are discussed. Approaches to optimising delivery of the peptide and protein drugs are highlighted.

Introduction

With the recent advances in recombinant DNA technology, the commercial production of proteins and peptides for pharmaceutical purpose is now routine. The list of available therapeutic agents produced by this technology is expanding rapidly to include interferon, macrophage activation factors, tissue plasminogen activator, neuropeptides and experimental agents that may have potential in cardiovascular disease, inflammation, contraception and so on. Unfortunately, protein and peptide drugs possess some chemical and

physical properties, including molecular size, susceptibility to proteolytic breakdown, rapid plasma clearance, immunogenicity and denaturation, which make them unsuitable for delivery using the normal absorption routes and in particular, the oral route. In part one of this review protein and peptide drugs are considered with particular emphasis on their pharmacological profiles, potential routes of delivery and their associated problems.

Recent major reviews on the subject include the general article by Gardner (1984) on the intestinal absorption of intact peptides and proteins and that by Humphrey and Ringrose (1986) on the absorption, metabolism and excretion of peptide and related drugs. In a further review, Lee (1988) discussed enzymic barriers to peptide and protein absorption. Banga and Chien (1988)

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broadened the scope and considered systemic delivery of those agents in general.

Terminology

Peptide or protein drugs are derived from amino acids by peptide bond linkages. Proteins are large peptides. Peptides containing less than eight amino acid residues are called small pentides. Peptide drugs in this group include enalapril, lisinopril and thyroid releasing hormone analogues. The term polypeptide drugs refers to peptide drugs with eight or more amino acid residues and includes cyclosporin, leuproline and luliberin. Polypeptide drugs containing from about 50 to as many as 2500 amino acid residues are named protein drugs. These include insulin, growth hormone and interferons. Some protein drugs, such as insulin or IgG containing two or more polypeptide chains, are called oligomeric proteins and their component chains are termed subunits or protomers.

Therapeutic Uses of Peptide and Protein Drugs

Peptide and protein drugs can be conveniently classified according to their activity profiles as follows:

Enzymes

Some exogenous enzymes have been used as enzyme replacement therapy in the treatment of enzyme deficiency diseases such as lysosomal storage and mannosidosis (Table 1). Because enzyme deficiency in humans is usually genetic in origin, enzyme replacement is often the only available therapy. Some exogenous enzymes have also been utilized in the treatment of diseases other than inborn enzyme deficiency. Good examples include t-PA (tissue plasminogen activators), urokinase and streptokinase. These enzymes activate circulating plasminogen and fibrin clot-associated plasminogen equally well and, because of this, they have been marketed in the U.K. and U.S.A. (Robinson and Sobel, 1986; British National Formulary, 1989). Thrombin-like enzymes of snake venoms have also been developed for dissolving blood clots through enhanced release of fibrinopeptides from fibrinogen (Kornalik, 1985).

Hormones

Hormones represent the largest class of protein or peptide drugs used in medical therapy. All hormones have 'target cells' on which they act and these may be located in a specific organ or be more widely distributed in the body. Some hor-

TABLE 1
Therapeutic application of some enzymes

Enzymes	Therapeutic application	Reference
Adenosine deaminase	Enzyme deficiency	Hershfield et al. (1987)
Dextranase	Lysosomal storage	Colley and Ryman (1974)
β-Fructofuranosidase	Storage disease	Gregoriadis and Ryman (1972b)
α-Mannosidase	Mannosidosis	Patel and Ryman (1974)
		Fishman and Citri (1975)
L-Asparaginase	Cancer	Abuchowski et al. (1984)
β-Glucosidase	Adult Gaucher's disease	Braidman and Gregoriadis (1976)
Tissue plasminogen activators	Thrombosis	Robinson and Sobel (1986)
Urokinase	Thrombosis	Robinson and Sobel (1986)
Streptokinase	Thrombosis	Robinson and Sobel (1986)
Thrombin-like enzymes of snake venoms	Thrombosis	Kornalik (1985)

mones like luliberin (luteinizing hormone releasing hormone, LHRH) function solely to bring about the release of other hormones from different endocrine glands. It is also well known that many hormones act by means of a second messenger and quite often this is cyclic AMP (cAMP) which is formed from ATP. On reaching its receptor in the cell membrane, the hormone causes the release of cAMP, which is the actual regulator of the metabolic process. In this way, the physiological effect of one molecule of the hormone is amplified many times (Wills, 1985). Because hormones are very specific and a tiny amount can produce large pharmacological effects, they are ideal for biotechnological development which is more suitable for relatively small outputs. Perhaps the best known hormone drug is insulin which has been used as an endocrinotherapeutic agent since the 1920's (Banting and Best, 1922).

Enzyme inhibitors

Enzyme inhibitors have been used as drugs for a long time. These include proteins such as aprotinin, and peptide drugs such as enalapril and lisinopril. Captopril is an inhibitor of angiotensin converting enzyme (ACE), which catalyses in vivo generation of angiotensin II from the decapeptide, angiotensin I, to constrict arterioles and increase cardiac output, leading to hypertension in man. Captopril is now a widely used antihypertensive agent (Romankiewicz et al., 1983). Enalapril and lisinopril are subsequent developments which are also becoming widely adopted for the treatment of hypertension and congestive heart failure (Todd and Heel, 1986; Lancaster and Todd, 1988).

Antimicrobial agents

A number of antimicrobial agents are peptide drugs, for example, the penicillins, cephalosporins, polymyxin B sulphate, actinomycin and bleomycin. Structurally, these drugs are small peptides, mostly containing a non-peptide moiety. All of these antimicrobial drugs are microbial metabolites.

Immunomodulating peptides and proteins

Endogenous immunomodulating agents

These agents are now produced by molecular genetic approaches. Well-known examples are the interferons (IFNs) which are families of inducible secretory proteins produced by eukaryotic cells in response to viral and other stimuli. Interferons are not directly antiviral but they act prophylactically by inducing antiviral proteins. These protect cells from viral infection by inhibiting virus-directed translation and transcription (Moore and Dawson, 1989). Another example is interleukin-2 (IL-2) which exerts its biological effect through cell surface receptors on activated T and B cells and on NK cells (natural killer cell). Interleukin-2 has been administered clinically in attempts to restore immunocompetence in patients suffering from the acquired immunodeficiency syndrome (AIDS), and to improve the immunocompetence of cancer patients (Dawson and Moore, 1989).

Exogenous immunomodulating agents

Some exogenous immunomodulating agents are also used to promote immunocompetence in man. For example, cyclosporin (CS-4), a cyclic undecapeptide which is isolated from *Tolypocladium inflatum* Gams, is widely used as an immunosuppressive (Calne et al., 1978; Cantarovich et al., 1987; Mehta et al., 1988; Borel, 1989), whereas muramyl dipeptide has been used as an immunological adjuvant (Kreuger et al., 1984; Bomford, 1989).

Vaccines

Vaccines derived from the infective microorganisms are introduced into the mammalian body to induce antibody formation against the pathogens. Well-known examples include measles vaccine and polio vaccine. It is anticipated that an increasing number of such vaccines will be biotechnologically produced, to give more specific and pronounced antigenic responses.

Absorption of Peptide and Protein Drugs

Analytical problems

Several methods have been employed for studying the absorption of peptide and protein drugs. However, high molecular weight proteins and polypeptides present some unique difficulties. Techniques such as gel filtration and ion-exchange HPLC usually have to be used. Even so, it is still very difficult to assay them in the presence of body fluids such as blood and urine. In such cases, radioassays or radioimmunoassays are often the most appropriate and hence, these techniques have been widely used in the measurement of the bioavailability of peptide or protein drugs. However, radioassays may be non-specific, and many chemical assay procedures may by themselves influence the conformation of protein

drugs, thereby causing the loss of their biological activities. The entity being chemically assayed may not be the biologically active moiety and in such cases, in vitro or in vivo bioassays are often used during absorption studies. For protein/peptide hormones, the measurement of pharmacological responses may be the assay method of choice. For enzymes or enzyme inhibitors, specific enzyme reactions may be the best analytical method. The bioavailability of immunomodulating and antimicrobial agents may be evaluated using some specific animal models and indicator microorganisms. For example, the prophylactic

TABLE 2
Instability of protein and peptide drugs

Effect factor	Protein or peptide drugs	Reference		
Physical instability				
Aggregation	Interferon-y	Hsu and Arakawa (1985)		
		Arakawa et al. (1987)		
	Bovine growth hormone	Brems et al. (1986)		
		Brems et al. (1988)		
Precipitation	Insulin	Brennan et al. (1985)		
		Loughced et al. (1980)		
Chemical instability				
β Elimination	Lysozyme	Nashef et al. (1977)		
	Phosvitin	Sen et al. (1977)		
Deamidation	Bovine growth hormone	Lewis and Cheever (1965)		
	Human growth hormone	Lewis et al. (1970)		
		Becker et al. (1988)		
	Insulin	Berson and Yalow (1966)		
		Fisher and Porter (1981)		
	r-Immunoglobulin	Minta and Painter (1972)		
	Epidermal growth factor	Diaugustine et al. (1987)		
	Prolactin	Graf et al. (1970)		
	Gastrin releasing peptide	McDonald et al. (1983)		
	ACTH	Graf et al. (1971)		
		Bhatt et al. (1990)		
Disulphide exchange	Lysozyme	Volkin and Klibanov (1987)		
	Ribonuclease A	Zale and Klibanov (1986)		
Racemization	ACTH	Geiger and Clarke (1987)		
		Meinwald et al. (1986)		
Oxidation	Corticotropin	Dedman et al. (1961)		
	α -, β -Melanotropins	Dixon (1956)		
	Parathyroid hormone	Tashjian et al. (1964)		
	Gastrin	Morley et al. (1965)		
	Calcitonin	Riniker et al. (1968)		
	Corticotropin releasing factor	Vale et al. (1981)		

TABLE 3
Liposomes as peptide and protein carrier

Liposome composition	Peptide or protein	Route	Animal model	Reference
Phosphatidyl- choline: cholesterol 7:2	semipurified glucocerebroside β-glucosidase	i.v.	man	Belchetz et al. (1977)
Phosphatidyl- choline: cholesterol 7:7	highly purified glucocerebroside β-glucosidase	i.v.	man	Gregoriadis et al. (1982)
Phosphatidyl- choline: cholesterol: phosphatidic acid 7:2:1	bacterial amyloglucosidase	i.v.	man	Tyrell et al. (1976)
Dimyristoyl phosphatidyl- choline: choles- terol: dicetyl phosphate 1:0.75:0.11	cholera toxin human malaria sporozoite antigen	i.v.	rabbit	Alving et al. (1986)
Phosphatidyli- nositol	insulin	i.v.	mouse rat	Dapergolas and Gregoriadis (1976)
Phosphatidyl- choline: choles- terol: dicetyl phosphate 10:2:1	insulin	oral	rat	Patel and Ryman (1976)
Phosphatidyl- choline: choles- terol: dicetyl phosphate 3:9:1	insulin	oral	rat	Tanaka et al. (1975)
Phosphatidyl- choline: phos- phatidylserine 7:3	muramyl peptide	i.v.	mouse guinea-pig	Fidler et al. (1985)
Phosphatidyl- choline: choles- terol: dicetyl phosphate 7:1:2	lysozyme			Sessa and Weissmann (1970)
Phosphatidyl- choline: choles- terol: phospha- tidic acid 20:1.5:0.2	adenovirus type 5 hexon protein	i.v.	mouse	Six et al. (1988)
Phosphatidyl- choline: choles- terol: phospha- tidic acid 7:1:2	lysozyme			Sessa and Weissmann (1970)

(continued)

TABLE 3 (continued)

Liposome composition	Peptide or protein	Route	Animal model	Reference
Microcapsules	catalase	i.s.	mouse	Chang and Poznansky (1968)
Phosphatidyl- choline : choles- cerol : dicetyl chosphate 7 : 2 : 1	amylo g lucosidase	i.v.	rat	Gregoriadis and Ryman (1972a)
Phosphatidyl- holine:choles- erol:phospha- idic acid :2:1	yeast invertase	i.v.	rat	Gregoriadis and Ryman (1972b)
Phosphatidyl- choline: choles- erol: phospha- idic acid 7:2:1	neuraminidase	i.v.	rat	Gregoriadis et al. (1974a)
Phosphatidyl- choline: choles- erol: phospha- idic acid 7:2:1	dextranase	i.v.	rat	Colley and Ryman (1974)
Phosphatidyl- choline: choles- erol: phospha- idic acid 7:2:1	lpha-mannosidase	i.v.	rat	Patel and Ryman (1974)
Dipalmitoyl- phosphatidyl- pholine	α-amylase		amoeba	Batzri and Korn (1975)
Phosphatidyl- choline: choles- erol: phospha- idic acid 7:2:1	horseradish peroxidase		rat	Wisse and Gregoriadis (1975)
Phosphatidyl- choline: choles- cerol: Phospha- idic acid 7:2:1	asparaginase	i.v.	mouse	Neerunjun and Gregoriadis (1976)
Phosphatidyli- nositol	glucose oxidase	i.v.	mouse	Dapergolas et al. (1976)
Phosphatidyl- choline : choles- terol 7 : 2	albumin	i.v.	rat	Gregoriadis and Neerunjun (1974)
Phosphatidyl- choline:choles- terol:phospha- tidic acid 7:2:1	albumin	i.v.	man	Gregoriadis et al. (1974b)

TABLE 3 (continued)

Liposome composition	Peptide or protein	Route	Animal model	Reference
Phosphatidyl- choline: choles- terol: dicetyl phosphate 6:6:2	albumin	i.v.	mouse	Heath et al. (1976)
Phosphatidyl- choline: choles- terol: dicetyl phosphate 7:2:1	diphtheria toxoid	i.v.	mouse	Gregoriadis and Alison (1974)
Phosphatidyl- choline: choles- terol: phospha- tidic acid 7:2:1	fetuin	i.v.	rat	Gregoriadis and Neerunjun (1975)
Phosphatidyl- choline: choles- terol: phospha- tidic acid 4:2:1	anti-α- glucosidase	i.v.	rat	De Barsy et al. (1975)
Phosphatidyl- choline: choles- terol: phosphati- dylethanolamine 10:10:1	monoclonal anti-Thyi IgG1	i.v.	mouse	Debs et al. (1987)
Distearoylphos- phatidylcholine: (2-puridyldithio)- propionol-dipal- mitoylphospha- tidylcholine: cholesterol 0.99:0.01:1	monoclonal anti-Thy1 IgG1	i.v.	mouse	Wolff and Gregoriadis (1984)
^a Liposomes	TRH	i.v.	cat	Kumashiro et al. (1986)
Phosphatidyl- choline: phos- phatidylserine 7:3	superoxide dismutase	intratracheal injection	rat	Padmanabhan et al. (1985)
Phosphatidylcholine: phosphatidic acid 15.3:0.1	factor VIII	oral	man	Sakuragawa et al. (1985)
Dipalmitoyl- phosphatidyl- choline: cholesterol: <i>m</i> -ma- leimidobenzoyl- (dipalmitoyl- phosphatidyl)- ethanolamine 25:17.5:2.5	subunits of monoclonal IgM	i.p.	mouse	Hashimoto et al. (1986)

^a The composition of liposomes was not mentioned in the paper.

usefulness of intranasal IFN- β against rhinovirus infection was determined using healthy volunteers or animals (Higgins et al., 1986).

Assay methods available for monitoring the absorption of small peptide drugs are freely available and routine methods include reverse-phase HPLC, TLC, and fluorescence techniques.

Stability

Irrespective of which dosage form is used, peptide or protein decomposition may be a problem. Drug breakdown can take place both in the formulation and when present in tissue fluids. First pass metabolism and enzymic breakdown are discussed in greater detail further on. Non-enzymic breakdown may be of two types: chemical and physical changes. Physical changes include aggregation and precipitation and are usually induced by high concentrations of co-solvents which may be used in some formulations or by injudicious choice of ionic strengths. Loss of conformation not only leads to poor absorption but also to loss of activity. Chemical changes include β -elimination, deamidation, disulphide exchange, racemization and oxidation. Examples of peptides and proteins which have been reported to be unstable are shown in Table 2 along with some of the reported mechanisms of breakdown.

Parenteral routes of delivery

For systemic delivery of peptide and protein drugs, parenteral administration is currently almost universally required in order to achieve consistent therapeutic activities. This is because of the drugs' susceptibility to breakdown by gastric acid and the proteolytic enzymes in the gastrointestinal tract. In addition, peptides and proteins are high-molecular-weight substances and thus do not easily cross the intestinal mucosa. Therefore, the oral bioavailabilities of most intact peptides and proteins are very low.

Of the parenteral routes, only intravenous (i.v.) administration is usually efficient in delivering protein and peptide drugs to the systemic circulation. For example, optimal blood levels of protein

or peptide drugs, such as γ -globulin (Buckley, 1982), can be achieved by the intravenous route. Generally, intramuscular or subcutaneous injections are less efficient due to the absorption and diffusion barriers presented by the muscle mass and connective tissues under the skin. However, insulin can be efficiently administered by subcutaneous injections (Nora et al., 1964; Koivisto and Felig, 1978) although hydrolysis is still significant (Berger et al., 1979).

While most peptide/protein drugs can be efficiently delivered to the systemic circulation by parenteral injections, poor disposition profiles lead to sub-optimal therapeutic benefits without high dosing frequencies. Such frequent injections, besides being unpleasant to the patients, also lead to usual complications such as thrombophlebitis and tissue necrosis.

In attempts to improve the disposition profile and the efficiency of delivery of parenterally administered peptides and protein drugs, many investigators have reported on liposomal systems. Examples of enzymes and monoclonal antibodies which have been formulated as liposomal systems for intravenous administration are shown in Table 3. Also included are some liposomal systems intended for oral administration. Despite the extensive evaluation of such systems and experimental results (Gregoriadis, 1976; Goosen, 1987) indicating that insulin absorption is greatly enhanced in animals by liposomal encapsulation of the hormone, no insulin liposomal system is currently in commercial use.

One biodegradable implant in current use in humans in goserelin acetate formulated in a biodegradable matrix of lactide-glycolide co-polymer. Systems which are designed with an enzymically controlled feed-back mechanism have also been described. Fischel-Ghodsian et al. (1988), for example, reported on an insulin system consisting of insulin and glucose oxidase dispersed in an ethylene/vinyl acetate polymer matrix. In the presence of glucose oxidase, glucose is converted into gluconic acid. This acid lowers the pH and increases the solubility of entrapped insulin which is then released faster. Consequently, some feedback control between glucose and insulin is thereby established.

General Approaches to Optimizing Absorption and Disposition

To optimize the absorption of high-molecularweight protein and peptide drugs across absorption barriers, several approaches are available: (i) inhibition of their enzymic degradation; (ii) increasing their permeability across the relevant membrane; and (iii) improving their resistance to breakdown by structural modification.

Inhibitors of proteolytic enzymes

Protease inhibitors have been known for several years to increase the absorption of protein drugs (Laskowski et al., 1958). Table 4 lists the different protease inhibitors which have been used in investigations of the delivery of peptide and protein drugs.

Aprotinin, a bovine pancreatic kallikrein inhibitor, consists of a single-chain polypeptide containing 58 amino acid residues with a molecular weight of 6500 (Kassell et al., 1965). It has been used to inhibit plasmin, trypsin, chymotrypsin and various intracellular proteases (Trautschold et al., 1967). It was demonstrated, in an early study, that when insulin and aprotinin

were injected together into a loop of the jejunum, a significant drop in blood glucose was observed. In contrast, no significant drop in blood glucose was found when the insulin was injected alone (Laskowski et al., 1958). Similar results have also been reported by several other workers (Berger et al., 1980; Fredenberg et al., 1981; William et al., 1983; Dandona et al., 1985; Linde and Gunnarson, 1985; Owens et al., 1988). However, some recent studies provided conflicting results, at least with respect to insulin and calcitonin absorption by nasal administration (Hanson et al., 1986; Aungst and Rogers, 1988). When the effects of laureth-9, sodium salicylate, Na₂EDTA and aprotinin on insulin absorption via the rectal, nasal and buccal tissues were examined by Aungst and Rogers (1988), aprotinin was found to be ineffective, either alone or in combination with laureth-9. Hanson and his co-workers (1986) examined the effects of several protease inhibitors, including bile salt, fatty acid derivative, aprotinin, kallikrein inhibitor, RG-1, bestatin, fusidic acid, chemostatin, benzamidine, chymotrypsin inhibitor, trypsin inhibitor III-0 and leupeptin on intranasal delivery of calcitonin, and found that aprotinin in vitro did not inhibit proteolytic activity of nasal extracts. In vivo the inhibitor did not

TABLE 4
Inhibitors of proteolytic enzymes used in investigation of the delivery of peptide and protein drugs

Compound	Route	Peptide studied	Animal model	Reference
Aprotinin	intestinal	insulin	rat	Ziv and Kidron (1987),
				Laskowski et al. (1958)
		RNase	rat	Ziv and Kidron (1987)
	s.c. a	insulin	man	Owens et al. (1988),
				Linde and Gunnarsson (1985),
				Berger et al. (1980)
Soybean	intestinal	insulin	rat	Ziv and Kidron (1987)
trypsin inhibitor		RNase	rat	Ziv and Kidron (1987)
FK-448 b (chymotrypsin inhibitor)	intestinal	insulin	rat	Yokoo et al. (1988)
Boroleucine c	nasal	Leu-enkephalin	rat	Hussain et al. (1989)
Borovaline c	nasal	Leu-enkephalin	rat	Hussain et al. (1989)

^a Subcutaneous delivery.

^b 4-(4-Isopropylpiperazinocarbonyl)phenyl-1,2,3,4-tetrahydro-1-naphthoate methanesulphonate.

^c α-Aminoboronic acid derivatives.

enhance the serum calcium drop observed. These results are supported by the study of Deurloo et al. (1989). The co-administration of sodium taurodihydrofusidate with aprotinin also failed to increase significantly insulin bioavailability in rabbits via the nasal route. Clearly, further studies are required to define better the effects of aprotinin on the absorption of peptide and protein drugs.

More recently, α -aminoboronic acid derivatives, such as boroleucine, which are potent and reversible inhibitors of aminopeptidase, have been used to stabilize peptide drugs during their intranasal absorption (Hussain et al., 1989). When these inhibitors were compared with other known peptidase inhibitors, bestatin [an inhibitor of leucine aminopeptidase, aminopeptidase B, and aminopeptidase N (Suda et al., 1976)] and puromycin [an inhibitor of aminopeptidase B and N but not leucine aminopeptidase (McDonald et al., 1964)], using leucine enkephalin as substrate in rat nasal perfusate, it was found that bestatin and puromycin were less effective than boroleucine, even at concentrations 100- and 1000times higher, respectively.

Absorption enhancers

The use of absorption enhancers has been studied extensively, particularly with respect to insulin absorption. These enhancers can be divided into several groups as listed in Table 5.

Despite extensive use, it is very difficult to make a judgement about the relative efficacy of these bioenhancers in promoting peptide or protein absorption because the results were obtained in different laboratories using different assay methods and different experimental conditions. However, it is clear that the bioavailability of most peptide and protein drugs administered by any non-parenteral route may be significantly enhanced by some of these compounds (see Tables in part II of this review).

The value of a particular enhancer depends on the route of administration used. For example, the bioavailability of ocular insulin was found to be significantly enhanced by saponin, whereas enhancement by glycocholate, which was a potentially good enhancer for nasal and rectal peptide and protein drug absorption, was found to be only slight (Chiou and Chuang, 1989).

The mechanisms of action of the peptide absorption enhancers are not clearly known, but several possibilities have been postulated. The first is increased solubility of the drugs brought about by the enhancers because proteins and peptides usually form aggregates in aqueous solutions. In the presence of enhancers, dissociation takes place to form monomers which are better absorbed. A second mechanism is the protection of the peptide and protein drugs from potential proteolytic hydrolysis. Both bile salts (Hirai et al., 1981b; Hanson et al., 1986; Zhou and Li Wan Po, 1991) and derivatives of fusidic acid (Deurloo et al., 1989) are known to inhibit proteolytic degradation of the drugs by nasal homogenates. Thirdly, binding between peptide or protein and enhancer to produce a better-absorbed entity may be a possibility. Although the effects of absorption enhancers such as glycocholate (Hirai et al., 1981b) and sodium cholate (Zhou and Li Wan Po, 1991) on the absorption of insulin by nasal delivery are thought to be partly due to inhibition of protease, recent work suggests that compared to aminopeptidase inhibitors such as bestatin and amastatin, cholate and its analogues are not very efficient (Hanson et al., 1986). Cholate and its analogues may also enhance the absorption of peptide and protein drugs by binding to insulin (Zhou and Li Wan Po, 1991). This would prevent the formation of enzyme-substrate complex to undergo the necessary conformational change which aligns the catalytic site on the protease with the susceptible bond of the substrate. Cholate and its analogues may possibly also promote the absorption of proteins by selectively denaturing the enzymes, although this is unlikely as it is difficult to identify the basis for the necessary selectivity.

Chemical modification

Chemical modification is an important approach for enhancing the absorption of peptides and protein drugs, especially for peptides with fewer than ten amino acid residues. Chemical modification usually results in denaturation of

TABLE 5
Absorption enhancers for peptides and proteins

Compound	Route	Peptide	Animal model	Reference
Fatty acid	37			
MCFC ^a				
Caprylate	nasal	insulin	rat	Mishima et al. (1987)
Caprate	nasal	insulin	rat	wishina ct al. (1967)
Laurate	nasal	insulin	rat	
	nusui	mount	Tat	
LCFC b				
Oleate in PAGB ^c	rectal	insulin	rat	Morimoto et al. (1983)
Linoleate in PAGB	rectal	insulin	rat	
Linolenate in PAGB	rectal	insulin	rat	
Oleic acid	vaginal	leuprolide	rat	Okada et al. (1982)
Bile salts				
Taurocholate	nasal	insulin	rat	Hirai et al. (1981a)
Cholate	nasal	insulin	rat	Moses et al. (1983)
Deoxycholate	nasal	insulin	rat	
Glycocholate	nasal	insulin	rat	Mishima et al. (1987)
Chenodeoxycholate	nasal	insulin	rat	
Deoxycholate (aerosol)	nasal	insulin	man	Moses et al. (1984)
Glycocholate	vaginal	leuprolide	rat	Okada et al. (1982)
Cholate	rectal	insulin	rat	Ziv et al. (1981)
Deoxycholate	rectal	insulin	rat	Lit of all (1901)
Cholate	intestinal	insulin	rat	Ziv et al. (1987)
	intestinal	RNase	rat	211 et al. (1707)
Glycocholate	nasal	insulin	rat	Aungst et al. (1988)
,	rectal	insulin	rat	rungst et al. (1700)
	buccal	insulin	rat	
	sublingual	insulin	rat	
Ethylacetoacetate enamine of sodium D-glycine	glycine rectal	insulin	rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium p-glycine Ethylacetoacetate		insulin	rabbit	Kim et al. (1983)
enamine of sodium D-glycine Ethylacetoacetate enamine of sodium	rectal			Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine		insulin insulin	rabbit rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate	rectal			Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium	rectal	insulin	rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine	rectal			Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate	rectal	insulin	rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium	rectal rectal	insulin insulin	rabbit rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine	rectal	insulin	rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate Ethylacetoacetate Ethylacetoacetate	rectal rectal	insulin insulin	rabbit rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium	rectal rectal rectal	insulin insulin insulin	rabbit rabbit rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-phenylalanine	rectal rectal	insulin insulin	rabbit rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-phenylalanine Ethylacetoacetate	rectal rectal rectal	insulin insulin insulin	rabbit rabbit rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-phenylalanine Ethylacetoacetate enamine of sodium	rectal rectal rectal	insulin insulin insulin	rabbit rabbit rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-phenylalanine Ethylacetoacetate enamine of sodium D-phenylalanine	rectal rectal rectal rectal	insulin insulin insulin insulin	rabbit rabbit rabbit rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-phenylalanine Ethylacetoacetate enamine of sodium D-phenylalanine in gelatin	rectal rectal rectal	insulin insulin insulin	rabbit rabbit rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-phenylalanine Ethylacetoacetate enamine of sodium D-phenylalanine Ethylacetoacetate entamine of sodium D-phenylalanine in gelatin Ethylacetoacetate	rectal rectal rectal rectal	insulin insulin insulin insulin	rabbit rabbit rabbit rabbit	Kim et al. (1983)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-phenylalanine Ethylacetoacetate eriamine of sodium D-phenylalanine Ethylacetoacetate eriamine of sodium D-phenylalanine in gelatin Ethylacetoacetate enamine of sodium	rectal rectal rectal rectal rectal	insulin insulin insulin insulin	rabbit rabbit rabbit dog	
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-phenylalanine Ethylacetoacetate enamine of sodium D-phenylalanine in gelatin Ethylacetoacetate enamine of sodium D-phenylalanine in gelatin Ethylacetoacetate enamine of sodium D-phenylalylicate	rectal rectal rectal rectal	insulin insulin insulin insulin	rabbit rabbit rabbit rabbit	Kim et al. (1983) Kamada et al. (1981)
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-phenylalanine Ethylacetoacetate enamine of sodium D-phenylalanine in gelatin Ethylacetoacetate enamine of sodium D-phenylalanine in gelatin Ethylacetoacetate enamine of sodium D-phenylglycinate Ethylacetoacetate	rectal rectal rectal rectal rectal	insulin insulin insulin insulin	rabbit rabbit rabbit dog	
Ethylacetoacetate enamine of sodium D-glycine Ethylacetoacetate enamine of sodium D-alanine Ethylacetoacetate enamine of sodium D-leucine Ethylacetoacetate enamine of sodium D-isoleucine Ethylacetoacetate enamine of sodium D-phenylalanine Ethylacetoacetate enamine of sodium D-phenylalanine in gelatin Ethylacetoacetate enamine of sodium D-phenylalanine in gelatin Ethylacetoacetate enamine of sodium D-phenylalylicate	rectal rectal rectal rectal rectal	insulin insulin insulin insulin	rabbit rabbit rabbit dog	

rued)

TABLE 5 (continued)

Compound	Route	Peptide	Animal model	Reference
Ethylacetoacetate				
enamine of sodium				
D-phenylglycine	rectal	lysozyme	rabbit	
Estan truns				
Ester type Glycerine-1,				
3-diacetoacetate	rectal	lysozyme	rabbit	Miyake et al. (1984)
1,2-Isopropylidene-	roctur	iyso2yine	140011	mijane et an (1501)
glyceryl-3-				
acetoacetate	rectal	lysozyme	rabbit	
Ethylaceto-				
acetylglycolate	rectal	łysozyme	rabbit	
Polyoxyethylene		• •		
10-monolaurate	nasal	insulin	rat	Hirai et al. (1981a)
Glyceryl esters of				
acetoacetic acid	rectal	insulin	rabbit	Nishihata et al. (1983)
Ether type				
Polyoxyethylene				
9-lauryl ether				
in PAGB	rectal	calcitonin	rat	Morimoto et al. (1985)
Polyoxyethylene				
9-lauryl ether	rectal	insulin	dog	Shichiri et al. (1978)
Polyoxyethylene			•	
24-cholesteryl				
ether	oral	ergot peptide alkaloids	rat	Urbancic-Smerkolj et al. (1987)
Polyoxyethylene				
5-octyl ether	nasal	insulin	rat	Hirai et al. (1981a)
Polyoxyethylene		,		
10-octyl ether	nasal	insulin	rat	
Polyoxyethylene		*		
5-lauryl ether	nasal	insulin	rat	
Polyoxyethylene	nasal	insulin	rat	
9-lauryl ether Polyoxyethylene	nasai	Houtili	iai	
10-lauryl ether	nasal	insulin	rat	
Polyoxyethylene	iiasai	mount	iai	
20-lauryl ether	nasal	insulin	rat	
Polyoxyethylene	nusui	and Wash		
10-cetyl ether	nasal	insulin	rat	
Polyoxyethylene				
20-cetyl ether	nasal	insulin	rat	
Polyoxyethylene				
10-stearyl ether	nasal	insulin	rat	
Polyoxyethylene				
20-stearyl ether	nasal	insulin	rat	
Polyoxyethylene				
10-nonylphenyl ether	nasal	insulin	rat	
Polyoxyethylene	_			
10-octylphenyl ether	nasal	insulin	rat	
Polyoxyethylene		1 Po.		
24-cholesteryl ether	nasal	insulin	rat	

(continued)

TABLE 5 (continued)

Compound	Route	Peptide	Animal model	Reference
Polyoxyethylene				
20-cetyl ether				
(Cetomacrogol 1000)	rectal	insulin	rat	Bar-on et al. (1981)
Polyoxyethylene				
9-lauryl alcohol ether	rectal	insulin	rabbit	Ichikawa et al. (1980)
Polyoxyethylene				
10-nonylphenyl ether	rectal	insulin	rabbit	
Salicylates				
3,5-Diiodosalicylate	rectal	insulin	rat	Hauss and Ando (1988)
5-Methoxysalicylate	rectal	insulin	rat	
5-Methoxysalicylate	rectal	insulin	rat	Nishihata et al. (1981)
5-Methoxysalicylate	rectal	gastrin	rat	Yoshioka et al. (1982)
, , , , , , , , , , , , , , , , , , , ,		pentagastrin	rat	•
5-Methoxysalicylate		- -		
in PAGB (pH 5.5,				
M.W. 1250000)	rectal	calcitonin	rat	Morimoto et al. (1984)
Salicylate	rectal	human growth	rat	Moore et al. (1986)
		hormone		
Organic acids				
Citric acid	vaginal	leuprolide	rat	Okada et al. (1982, 1983)
Chine dold	vaginal	insulin	rat	Onada ot an (1302, 1305)
Succinic acid	vaginal	leuprolide	rat	
Tartaric acid	vaginal	leuprolide	rat	
Malonic acid	vaginal	leuprolide	rat	
Glycoside	•			
Saponin	nasal	insulin	rat	Hirai et al. (1981a)
Saponin	ocular	insulin	rat	Chiou and Chuang (1989)
Daniel de Unit d				
Peptide lipid Surfactin	nasal	insulin	rat	Hirai et al. (1981a)
Surfactili	iiasai	msum	lat	fillal et al. (1961a)
Amine				
Polyoxyethylene				
5-oleylamine	rectal	insulin	rabbit	Ichikawa et al. (1980)
Anion				
Sodium lauryl				
sulphate	rectal	insulin	rabbit	Ichikawa et al. (1980)
Monosodium	rectar	Mount	labolt	icilikawa ci al. (1700)
N-lauroyl				
L-glutamate	rectal	insulin	rabbit	
_				
Derivatives of fusidic acid				
sodium tauro-				
dihydrofusidate	nasal	insulin	rabbit	Deurloo et al. (1989)
Sodium tauro-				
dihydrofusidate	nasal	insulin	sheep	Longenecker et al. (1987)
Glycyrrhetinic acid derivatives				
Glycyrrhizinic acid	nasal	insulin	rat	Mishima et al. (1989)
Glycyrrhetinic acid				
hydrogen succinate	nasal	insulin	rat	

^a Medium-chain fatty acids.

b Long-chain fatty acids.
c Polyacrylic acid gel base.

Fig. 1. Structure of cyclosporin (CS-A). MeBmt *, (4R)-4-[(E)-2-butenyl]-4-methyl-L-threonine.

the bioactive protein or polypeptide. However, a modified luteinizing hormone releasing hormone (LHRH), a decapeptide, and a modified tripeptide, thyrotrophin releasing hormone (TRH), were found to be active when given orally to rat and man. Both of these hypothalamic regulatory hormones contain a pyroglutamyl residue at the amino end, which renders them relatively resistant to enzymic hydrolysis (Masson et al., 1979). The possibility of formation of pyroglutamyl residues by cyclization of the N-terminal glutamine warrants more attention, because the pyroglutamyl residue is hydrolysis-resistant and may promote the gastrointestinal entry of intact peptide hormone into the systemic circulation. Peptides such as 1-deamino-8-D-arginine vasopressin and cyclosporin (Fig. 1) containing D-amino acids may lead to sequences which are relatively stable to enzymatic hydrolysis. Peptides with a high proline content or with a phosphate residue are also relatively resistant to enzymatic hydrolysis (Gardner, 1984).

Some peptide drugs can be converted into prodrugs. A good example is enalapril, which was found to be orally well absorbed and is metabolized to the active form, enalaprilat, in the liver. In contrast, the parent drug, enalaprilat is very poorly absorbed via the oral route (Ip and Brenner, 1987).

Conclusion

Peptide and protein drugs will form an increasingly important part of the range of therapeutic agents available for the control of human diseases. The problems associated with the need to use the parenteral route for most of those drugs present formidable challenges for the formulation scientist. Some promising avenues have been discussed in this review. Alternatives to the parenteral route are discussed in greater detail in part II of this review.

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