

AccuPrep® PCR/Gel Purification Kit (K-3037, K-3038)**I Recommendation**

- 1) Please prepare absolute isopropanol (not provided) before purification.
- 2) We recommend to use $\leq 1\%$ concentration of agarose gel for gel purification.
- 3) For high-purity DNA extraction from >200 mg of agarose gel (max. 400 mg), please use WB1 Buffer in washing step.

II Fragment DNA purification from PCR product

- 1) Add **3 volumes of FB Buffer** to PCR product (If the PCR product is 20 μ l, add 60 μ l of FB Buffer)
- 2) Add **1 volume of absolute isopropanol** and mix immediately by using pipette or inverting.
- 3) Transfer the mixture to a binding column in a 2 ml collection tube.
- 4) Close the lid and centrifuge **at 14,000 rpm for 1 min.**
- 5) Discard the flow-through and re-assemble the binding column with the collection tube.
- 6) Add **500 μ l of W2 Buffer** without wetting the rim, close the tube, and centrifuge **at 14,000 rpm for 1 min.** Discard the flow-through and re-assemble the binding column with the collection tube.
- 7) **Repeat step 6.**
- 8) Centrifuge once more **at 14,000 rpm for 1 min** to completely remove residual ethanol, and check that there is no droplet clinging to the bottom of the binding column.
- 9) Transfer the binding column tube to a new 1.5 ml tube for elution, add **30 μ l of EA Buffer** onto the binding column tube, and wait for at least 1 min at RT (15 - 25°C).
- 10) Centrifuge **at 14,000 rpm for 1 min** to elute.

AccuPrep® PCR/Gel Purification Kit (K-3037, K-3038)**III Fragment DNA purification from Agarose gel**

- 1) Visualize the band in agarose gel stained with any nucleic acid staining chemicals and cut the gel around the DNA band of interest using a scalpel blade.
- 2) The maximum amount of gel slice per each sample is ~ **400 mg**, Add **3 volumes of FB Buffer** to weight of the gel slice (If the weight of gel slice is 200 mg, add 600 µl of FB Buffer)
- 3) Incubate **at 50°C for 10 min** and mix by inverting the tube every 2-3 min during the incubation.
- 4) Add **1 volume of absolute isopropanol** and mix immediately by using pipette or inverting.
- 5) Transfer the mixture to a binding column in a 2 ml collection tube.
- 6) Close the lid and centrifuge **at 14,000 rpm for 1 min**.
- 7) Discard the flow-through and re-assemble the binding column with the collection tube.
- 8) **(Optional)** Add **500 µl of WB1 Buffer** without wetting the rim, close the tube, and centrifuge **at 14,000 rpm for 1 min**. Discard the flow-through and re-assemble the binding column with the collection tube.
- 9) Add **500 µl of W2 Buffer** without wetting the rim, close the tube, and centrifuge **at 14,000 rpm for 1 min**. Discard the flow-through and re-assemble the binding column with the collection tube.
- 10) **Repeat step 9.**
- 11) Centrifuge once more **at 14,000 rpm for 1 min** to completely remove residual ethanol, and check that there is no droplet clinging to the bottom of the binding column.
- 12) Transfer the binding column tube to a new 1.5 ml tube for elution, add **30 µl of EA Buffer** onto the binding column tube, and wait for at least 1 min at RT (15 - 25°C).
- 13) Centrifuge **at 14,000 rpm for 1 min** to elute.